

Diabetes, Non-enzymatic glycation, and aging**Carolina Reato Marçon, MD**¹ Department of Dermatology, Santa Casa de Misericórdia de São Paulo, São Paulo, SP, Brazil***Corresponding Author: Carolina Reato Marçon, MD**, Rua Piauí 305, Apto 114, Higienópolis, São Paulo, SP, Brazil 01241-001; Phone: 11 99934-7114, Email: carolrmarcon@hotmail.com**Citation:** Diabetes, Non-enzymatic glycation, Sci J of Der and Ven. 2020; 3(1): 01-29.**Submitted:** 20 January 2020; **Approved:** 22 January 2020; **Published:** 24 January 2020**Summary:**

First described in the context of diabetes, advanced glycation end products (AGEs) are formed through a type of non-enzymatic reaction called glycation. Protein glycation and formation of advanced glycation end products (AGEs) play an important role in the pathogenesis of diabetic complications like retinopathy, nephropathy, neuropathy, cardiomyopathy along with some other diseases such as rheumatoid arthritis, osteoporosis, and, recently, skin aging. Glycation of proteins interferes with their normal functions by disrupting molecular conformation, altering enzymatic activity, and interfering with receptor functioning. AGEs form intra- and extracellular cross linking not only with proteins, but with some other endogenous key molecules including lipids and nucleic acids. Recent studies suggest that AGEs interact with plasma membrane localized receptors for AGEs (RAGE) to alter intracellular signaling, gene expression, release of pro-inflammatory molecules and free radicals. Characteristic findings of aging skin, including decreased resistance to mechanical stress, impaired wound healing, and distorted dermal vasculature, can be in part attributable to glycation. Multiple factors mediate cutaneous senescence, and these factors are generally characterized as endogenous (e.g., telomere shortening) or exogenous (e.g., ultraviolet radiation exposure). Interestingly, AGEs exert their pathophysiological effects from both endogenous and exogenous routes. The former entails the consumption of sugar in the diet, which then covalently binds an electron from a donor molecule to form an AGE. The latter process mostly refers to the formation of AGEs through cooking. Results of several studies in animal models and humans show that the restriction of dietary AGEs has positive effects on wound healing, insulin resistance and cardiovascular diseases. Moreover, several dietary compounds have emerged as promising candidates for the inhibition of glycation-mediated aging. This article summarizes the evidence supporting the critical role of glycation in skin aging and highlights preliminary studies on dietary strategies that may be able to combat this process.

Keywords: advanced glycation end-products, diabetic complications, inflammation, oxidative stress, anti-AGE therapies, skin aging, Maillard reaction**Introduction**

Societal obsession with the process of aging dates back to ancient history, and myths related to the conservation of youth—ranging from a bathing fountain that confers eternal youth to a philosopher's stone that could be used to create an elixir of life—populate both past and contemporary folklore. However, it is only within recent years that aging has been

investigated from an empirical approach, as it continues to garner increasing attention from the scientific community. While several hypotheses have been proposed to explain the pathophysiology responsible for senescence, no single theory accounts for the diverse phenomena observed. Rather, aging appears to be a multifactorial process that results from a complex interplay of several factors and mechanisms.¹

Cite this article: Citation: Diabetes, Non-enzymatic glycation, Sci J of Der and Ven. 2020; 3(1): 01-29.

Aging is characterized by a decline of anatomical integrity and function across multiple organ systems and a reduced ability to respond to stress. The multisystem decline is associated with increasing pathology, disease, and progressively higher risk of death. Although the true mechanisms that drive the aging process are still a mystery, there is evidence that both genetic and environmental factors may affect the rate of appearance of phenotypes characteristic of the aging process. Thus, aging appears in part to be modulated by a genetic–environmental interaction.² Studies of gene expression across species and tissues have consistently observed that old age is associated with progressive impairment of mitochondrial function, increased oxidative stress, and immune activation.³ Interestingly, all these processes can be influenced by modification of nutritional intake. For example, studies of animal species have found that caloric restriction reduces oxidative stress and is associated with longer life expectancy. Recent studies have cast doubts on whether humans may be able to maintain a long-term regimen of caloric restriction without unacceptable psychological consequences and whether even caloric restriction may have overall positive health effects in humans.³ It has been suggested that changes in the quality of the diet could have positive effects on health and longevity and could be more easily implemented compared with caloric restriction.

Nevertheless, stratification of factors and mechanisms contributing to senescence is critical for the development of initial strategies in combating the aging process. The skin is an excellent paradigm for studying aging, in large part due to its easy accessibility. Moreover, in addition to its vulnerability to internal aging processes because of its diverse role in cellular processes, such as metabolism and immunity, the skin is subject to a variety of external stressors as the chief barrier between the body and the environment.

Aging factors can generally be classified as exogenous or endogenous. As ultraviolet (UV) radiation exposure is so strongly associated with a host of age-related skin diseases, endogenous and exogenous factors can theoretically be studied somewhat independently in the skin by differentiating between

UVprotected and UVexposed sites.⁴ Endogenously aged skin displays characteristic morphological features with resultant alterations in functionality. These include epidermal, dermal, and extracellular matrix atrophy leading to increased fragility, diminished collagen and elastin resulting in fine wrinkle formation, and marked vascular changes disrupting thermoregulation and nutrient supply. Endogenously aged skin also displays

decreased mitotic activity, resulting in delayed wound healing, as well as decreased glandular function, resulting in disturbed re-epithelialization of deep cutaneous wounds. Also seen is a reduction of melanocytes and Langerhans cells manifesting as hair graying and higher rates of infection, respectively.⁴⁻¹³

Exogenously aged skin, in which environmental factors such as UV radiation act in concert with endogenous processes, shares many of the characteristics of endogenously aged skin. In addition, exogenously aged skin displays a thickened epidermis and aggregation of abnormal elastic fibers in the dermis (i.e., solar elastosis).⁴

Among the many mechanisms thought to underlie aging, glycation has emerged in recent years as one of the most widely studied processes. Testament to the rapidly growing attention from the scientific community, a cursory literature search will yield thousands of articles related to glycation, the majority of them published in the last decade. Glycation refers to the nonenzymatic process of proteins, lipids, or nucleic acids covalently bonding to sugar molecules, usually glucose or fructose. The lack of enzyme mediation is the key differentiator between glycation and glycosylation. Glycosylation occurs at defined sites on the target molecule and is usually critical to the target molecule's function. In contrast, glycation appears to occur at random molecular sites and generally results in the inhibition of the target molecule's ability to function. The products of glycation are called advanced glycation end products (AGEs).

Advanced glycation end-products (AGEs) were first identified in cooked food as end-products from a non enzymatic reaction between sugars and proteins called the Maillard reaction.¹⁴ The Maillard reaction

(non enzymatic glycation or browning) in foods has been well studied by the food industry to control food quality. However, it is only 40 years ago that a similar glycation process was recognized in human body by the observation of increased formation of glycosylated haemoglobins in diabetic patients¹⁵ and this would lead to the formation of detrimental advanced glycation endproducts (AGEs) in humans.¹⁶

The body also produces AGEs naturally as it processes sugars. The formation of AGEs is a part of normal metabolism, but if excessively high levels of AGEs are reached in tissues and the circulation they can become pathogenic.¹⁷

The pathologic effects of AGEs are related to their ability to promote oxidative stress and inflammation by binding with cell surface receptors or cross-linking with body proteins, altering their structure and function.¹⁸ The formation and accumulation of AGEs is a characteristic feature of tissues in aged people, especially in patients with diabetes mellitus, and these products have also been strongly implicated in the pathogenesis of age-related and diabetic complications. Many chronic diseases, including heart diseases, diabetes and both osteoarthritis and rheumatoid arthritis are associated with inflammation. It has been reported that AGEs are involved in musculoskeletal diseases such as osteoarthritis (OA), which is the most common chronic disabling disorder for aged people. Accumulation of AGEs increases stiffness of the collagen network in the bone as well, which may explain some of the age-related increase in skeletal fragility and fracture risk.¹⁹ AGEs can be particularly dangerous for diabetics, as the increased availability of glucose in diabetes patients accelerates the formation of AGEs. Apart from their presence in a wide variety of body tissues, AGEs have also been identified from exogenously derived sources such as tobacco²⁰ and certain foods.²¹

Advanced glycation end products (AGEs) are a heterogeneous group of macromolecules that are formed by the nonenzymatic glycation of proteins, lipids, and nucleic acids. Humans are exposed to two main sources of AGEs: exogenous AGEs that are ingested in foods and endogenous AGEs that are formed in the body. The Western diet is rich in AGEs.

AGEs are formed when food is processed at elevated temperatures, such as during deep-frying, broiling, roasting, grilling; high temperature processing for certain processed foods such as pasteurized dairy products, cheeses, sausages, and processed meats; and commercial breakfast cereals. Endogenous AGEs are generated at higher rates in diabetics due to altered glucose metabolism. AGEs, by increasing oxidative stress and through other mechanisms, may accelerate the multisystem decline that occurs with aging and, therefore, reducing intake and circulating levels of AGEs may promote healthy aging and greater longevity.²

Increased accumulation of AGEs was first directly correlated to the development of diabetic complications. Since then, AGEs have been implicated in a host of other pathologies, including atherosclerosis, end stage renal disease, and chronic obstructive pulmonary disease.²² (It should be noted that AGE levels have been shown to vary by race and gender, and until larger studies are done to create ethnic- and gender-specific reference values, increased accumulation of AGEs should be defined as levels that are elevated for all demographic groups.²³) Not coincidentally, many of the pathologies associated with AGEs, including diabetic sequelae, are closely related to senescence.

This extends to aging skin, as methods of AGE detection, such as immunostaining, have demonstrated the prevalence of glycation in aged skin. Glycation results in characteristic structural, morphological, and functional changes in the skin, a process colloquially known as "sugar sag." With glucose and fructose playing such a prominent role in the mechanism, it is not surprising that diet plays a critical role in glycation and thus aging skin.¹

Perhaps more surprising, studies have shown that consumption of AGEs is not only tied to the sugar content of food, but is also affected by the method of cooking. Furthermore, as the connection between diet and aging is more clearly characterized, a host of dietary compounds have surfaced as potential therapeutic candidates in the inhibition of AGE-mediated changes.¹

Until today, more than 300 theories of aging have been proposed, among them the theory of cellular senescence, decreased proliferative capacity and telomere shortening, mitochondrial DNA single mutations, the free radical theory and others, none of which can fully explain all changes observed in aging.²⁴⁻²⁸ According to the inflammatory theory of aging, a common characteristic of skin aging factors is their ability to induce or maintain proinflammatory changes and trigger a local inflammatory response which through subsequent immune responses, matrix metalloproteinase (MMP) activation and proinflammatory cytokine production contributes to the structural changes observed in aged skin.²⁹

In the recent years, the role of advanced glycation end products (AGEs) has been increasingly discussed in skin aging, and the potential of anti-AGE strategies has received high interest from pharmaceutical companies for the development of novel anti-aging cosmeceutical compounds.³⁰ The study of AGE represents one of the most promising areas of research today. Although the initial chemistry behind their formation has been known since the early 1900s, it is only in the last 20 years or so that important work has been done to elaborate on this. The chemical processes and pathways that ultimately lead to AGE formation have, however, yet to be fully clarified.³¹ As our knowledge of AGE chemistry increases it is becoming apparent that not all AGE have been isolated, whereas as those that have been characterized are both complex and heterogenous. Thus, the discovery and investigation of AGE inhibitors would offer a potential therapeutic approach for the prevention of diabetic or other pathogenic complications.¹⁶

Biochemistry of AGEs

Glycation is the non-enzymatic reaction between reducing sugars, such as glucose, and proteins, lipids or nucleic acids.³² Glycation has to be distinguished from glycosylation, which is an enzymatic reaction. Since its first description by Maillard in 1912 and its involvement in food browning during thermal processing by Hodge 50 years later, its presence in living systems and involvement in various pathologies of the human body, including aging and diabetes,

have been an intensive field of research.^{33,34}

First described over a century ago, glycation entails a series of simple and complex non-enzymatic reactions. In the key step, known as the Maillard reaction, electrophilic carbonyl groups of the sugar molecule react with free amino groups of proteins, lipids, or nucleic acids, leading to the formation of a Schiff base. This nonstable Schiff base contains a carbon-nitrogen double bond, with the nitrogen atom connected to an aryl or alkyl group. The Schiff base rapidly undergoes re-arrangement to form a more stable ketoamine, termed the Amadori product. At this juncture, the Amadori product can: (1) undergo the reverse reaction; (2) react irreversibly with lysine or arginine functional groups to produce stable AGEs in the form of protein adducts or protein cross-links; or (3) undergo further breakdown reactions, such as oxidation, dehydration, and polymerization, to give rise to numerous other AGEs.³² AGE formation is accelerated by an increased rate of protein turnover, hyperglycemia, temperatures above 120° C (248° F), and the presence of oxygen, reactive oxygen species, or active transition metals.³⁵

AGEs comprise a highly heterogenous group of molecules. The first, and perhaps most well-known, physiological AGE to be described was glycated hemoglobin (hemoglobin A1C), now widely used to measure glycemic control in diabetes. However, the most prevalent AGE in the human body, including the skin, is carboxymethyl-lysine (CML), which is formed by oxidative degeneration of Amadori products or by direct addition of glyoxal to lysine. In the skin, CML is found in the normal epidermis, aged and diabetic dermis, and photoaging-actinic elastosis.³⁶⁻³⁸ Other AGEs detected in skin include pentosidine, glyoxal, methylglyoxal, glucosepane, fructoselysine, carboxyethyl-lysine, glyoxal-lysine dimer, and methylglyoxal-lysine dimer.³⁹

Formation of AGEs is a complicated molecular process involving simple and more complex multistep reactions. During the classical Maillard reaction electrophilic carbonyl groups of glucose or other reactive sugars react with free amino groups of amino acids (especially of basic lysine or arginine residues),

forming a non-stable Schiff base.⁴⁰ Further rearrangement leads to formation of a more stable ketoamine (Amadori product).^{32,40} Schiff bases and Amadori products are reversible reaction products. However, they can react irreversibly with amino acid residues of peptides or proteins to form protein adducts or protein crosslinks.^{32,40} Alternatively, they can undergo further oxidation, dehydration, polymerization and oxidative breakdown reactions to give rise to numerous other AGEs.^{32,35} Oxygen, reactive oxygen species (ROS) and redox active transition metals accelerate AGE formation. When an oxidative step is involved, the products are called advanced glycoxidation end products.^{32,35}

AGEs are a very heterogeneous group of molecules. Since the discovery of the first glycosylated protein, glycosylated hemoglobin in diabetes, numerous other AGEs have been detected. Some of them have characteristic autofluorescent properties, which simplifies their identification in situ or in vivo.³² To date, numerous AGEs have been identified.^{35-38,41-48}

Carboxymethyl-lysine (CML) was first described by Ahmed and represents the most prevalent AGE in vivo.^{49,50} It is a non-fluorescent protein adduct. Mechanisms of its formation include oxidative degradation of Amadori products or direct addition of glyoxal to lysine. It seems to be the major epitope of the commonly used polyclonal anti-AGE antibodies.⁵⁰

Pentosidine was first isolated and characterized by Sell and Monnier. It is composed of an arginine and a lysine residue crosslinked to a pentose.⁵¹ Pentosidine is a fluorescent glycoxidation product and forms protein-protein crosslinks.⁴⁰ Dicarbonyl compounds like 3-deoxyglucosone, methylglyoxal and glyoxal derive from oxidative degradation or autooxidation of Amadori products and other pathways.^{32,52} These dicarbonyl compounds are very reactive molecules leading to protein crosslinks.³² Other in vivo characterized AGEs include glucosepane, carboxymethyl-hydroxy-lysine, carboxyethyl-lysine (CEL), fructose lysine, methylglyoxal-derived hydroimidazolones and pyrraline, which form non-fluorescent protein adducts, while glyoxal-lysine dimer (GOLD) and methylglyoxal-lysine dimer (MOLD) form non-fluorescent protein crosslinks.^{32,35}

AGEs can be exogenously ingested (through food consumption) or be endogenously produced. Endogenous AGE formation is increased in diabetes; however, AGEs are also formed at lower rates by normal metabolic processes of the organism.⁵³

Environmental factors, such as diet and smoking influence the rate of AGE formation.⁵⁴ Moreover, it seems that the level of circulating AGEs levels are genetically determined, as shown in a cohort study of healthy monozygotic and heterozygotic twins.⁵⁵

The content of AGEs in the organism is not only defined by the rate of their formation but also by the rate of their removal. Many cells have developed intrinsic detoxifying pathways against accumulation of AGEs.⁵⁶ The glutathione-dependent glyoxalase system, comprising of glyoxalase (Glo) I and II, has a key role in the defense against glycation.⁵⁷ This system uses reduced glutathione (GSH) to catalyze the conversion of glyoxal, methylglyoxal and other α -oxoaldehydes to the less toxic D-lactate.⁵⁷ Other enzymatic systems include fructosyl-amine oxidases (FAOXs) and fructosamine kinases, relatively new classes of enzymes which recognize and break Amadori products.⁵⁸ However, FAOXs or "amadoriases" have been found to be expressed only in bacteria, yeast and fungi but not in mammals. They oxidatively break Amadori products but act mostly on low molecular weight compounds.⁵⁹ On the contrary, fructosamine kinases are expressed in various genomes including humans.⁵⁸ These intracellular enzymes phosphorylate and destabilize Amadori products leading to their spontaneous breakdown.⁵⁹ Fructosamine-3-kinase (FN3K), one of the most studied enzymes in this system, is almost ubiquitarily expressed in human tissues including the skin. Thus, it plays an important role in the intracellular breakdown of Amadori products.⁶⁰

Receptors for AGEs

AGEs not only exert their deleterious actions due to their biological properties per se, but also through their interaction with specific receptors. Receptor for AGEs (RAGE) is a multiligand member of the immunoglobulin superfamily of cell surface receptors, encoded by a gene on chromosome 6 near the major histocompatibility complex III. It is a pattern recognition receptor binding in addition to AGEs various other molecules such as S-100/calgranulins, high motility group protein B1 (amphoterine), β -amyloid peptides and β -sheet fibrils.^{53,61} The binding of ligands to RAGE stimulates various signaling pathways including the mitogen-activated protein kinases (MAPKs) extracellular signal-regulated kinases (ERK) 1 and 2, phosphatidylinositol 3 kinase, p21Ras, stress-activated protein kinase/c-Jun-N-terminal kinase and the janus kinases.^{53,61} Stimulation of RAGE results in activation of the transcription factor nuclear factor kappa-B (NF κ B) and subsequent transcription of many proinflammatory genes.⁶¹ Interestingly, RAGE-induced activation of NF κ B is characterized by a sustained and self-perpetuating action, through induction of positive feedback loops and overwhelming of the autoregulatory negative feedback loops. RAGE activation leads to new synthesis of the transcriptionally active subunit p65, which overwhelms the newly synthesized inhibitor I κ B α . Moreover NF κ B increases further expression of RAGE, which itself further stimulates NF κ B, forming a vicious cycle of self-renewing and perpetuating proinflammatory signals.⁶¹ RAGE activation can directly induce oxidative stress by activating nicotinamide adenine dinucleotide phosphate (NADPH)-oxidase (NOX), decreasing activity of superoxide dismutase (SOD), catalase and other pathways, and indirectly by reducing cellular antioxidant defenses, like GSH and ascorbic acid.⁶¹⁻⁶³ The reduction of GSH leads furthermore to decreased activity of Glo I, the major cellular defense system against methylglyoxal, therefore supporting further production of AGEs.³⁷ RAGE is almost ubiquitarily expressed in the organism, typically at low levels, and its expression is upregulated under various pathological conditions.^{61,64} In the skin, RAGE expression was observed in both epider-

mis and dermis, and it was increased in sun-exposed compared with UV irradiation-protected areas. Keratinocytes, fibroblasts, dendritic cells and to a lesser extent endothelial cells and lymphocytes express RAGE.⁶⁴ Not only in vivo, but also in vitro, various skin cells types have been shown to express RAGE.^{62,64-69}

RAGE is the most studied receptor for advanced glycation end products. Another group of cell surface receptors, AGER1, AGER2 and AGER3 seem to regulate endocytosis and degradation of AGEs, thus counteracting the effects of RAGE.⁷⁰ AGER1 has been further shown to counteract AGEs-induced oxidative stress via inhibition of RAGE signaling.^{71,72} Soluble RAGE (sRAGE) is a truncated splice variant of RAGE containing the ligand-binding domain but not the transmembrane domain and has been found in plasma. sRAGE is a soluble extracellular protein without signaling properties and it is considered as a natural decoy receptor of RAGE.⁷³

Toxicity of advanced glycation end-products (AGEs)

The possible pathophysiological role of AGEs has become a topic of increasing interest over the past few years. With the continued research on the Maillard reaction, it was demonstrated that the Maillard reaction also occurs in vivo and the term “glycation” was introduced as a synonym for “non-enzymatic glycosilation”, in order to distinguish this from the well known enzymatic glycosilation of proteins.⁷⁴ Protein modifications called “Advanced glycation end-products” (AGEs), which are formed during aging, diabetes and in renal failure via comparable chemical pathways as described for heated foods, nowadays are generally accepted to play a pivotal pathophysiological role in several diseases.⁷⁵ In vitro studies using humanderived endothelial cells exhibited the food-derived AGEs have same protein cross-linking and intracellular oxidant stress actions as their endogenous counterparts. In animal studies like in mice, reduction of dietary AGE intake is accompanied by significant reduction of circulating AGEs levels as well as reduction of diseases related to inflammation and oxidative stress. A low-AGE diet has been associated with a significant increase in mouse lifespan.

The human relevance of the *in vitro* and animal data discussed in a number of studies found independent correlate of the circulating AGEs with the dietary AGEs intake. Moreover, the effect of a low and a high-AGE diet on the inflammatory mediators was also studied by using a group of diabetic subjects. The low-AGE diet significantly reduced serum AGE levels as well as markers of inflammation and endothelial dysfunction. Thus, all these studies demonstrate the associated toxicity of AGEs.^{16,76}

AGEs and the Skin

AGEs accumulate in various tissues as a function, as well as a marker, of chronological age.⁷⁷ Proteins with slow turnover rates, such as collagen, are especially susceptible to modification by glycation. Collagen in the skin, in fact, has a half-life of approximately 15 years and thus can undergo up to a 50% increase in glycation over an individual's lifetime.⁷⁸

Collagen is critical not only to the mechanical framework of the skin but also to several cellular processes, and is impaired by glycation in multiple ways. First, intermolecular cross-linking modifies collagen's biomechanical properties, resulting in increased stiffness and vulnerability to mechanical stimuli.⁷⁹ Second, the formation of AGEs on collagen side chains alters the protein's charge and interferes with its active sites, thereby distorting the protein's ability to interact properly with surrounding cells and matrix proteins.⁸⁰ Third, the ability to convert L-arginine to nitric oxide, a critical cofactor in the crosslinking of collagen fibers, is impaired.⁸¹ Finally, glycated collagen is highly resistant to degradation by matrix metalloproteinases (MMPs). This further retards the process of collagen turnover and replacement with functional proteins.⁸²

Other cutaneous extracellular matrix proteins are functionally affected by glycation, including elastin and fibronectin. This further compounds dermal dysfunction^{39,83} as glycation crosslinked collagen, elastin, and fibronectin cannot be repaired like their normal counterparts.

Interestingly, CML-modified elastin is mostly found in sites of solar elastosis and is nearly absent in sun-protected skin. This suggests that UV-radiation can mediate AGE

formation in some capacity or, at the least, render cells more sensitive to external stimuli.⁸⁴ It is hypothesized that UV-radiation accomplishes this through the formation of superoxide anion radicals, hydrogen peroxide, and hydroxyl radicals. This induces oxidative stress and accelerates the production of AGEs.⁸⁵ AGEs themselves are very reactive molecules and can act as electron donors in the formation of free radicals. Occurring in conjunction with the decline of the enzymatic system that eliminates free radicals during the aging process, these properties lead to a "vicious cycle" of AGE formation in the setting of UV exposure.

Formed both intracellularly and extracellularly, AGEs can also have an effect on intracellular molecular function. In the skin, the intermediate filaments of fibroblasts (vimentin) and keratinocytes (cytokeratin 10) have been shown to be susceptible to glycation modification.⁴² Analogous to the diverse role of collagen in the skin, intermediate filaments are essential to both the maintenance of cytoskeletal stability and the coordination of numerous cellular functions. Fibroblasts with glycated vimentin demonstrate a reduced contractile capacity, and these modified fibroblasts are found to accumulate in skin biopsies of aged donors.⁴²

In fact, general cellular function may be compromised in the presence of high concentrations of AGEs. *In vitro*, human dermal fibroblasts display higher rates of premature senescence and apoptosis, which likely explains the decreased collagen and extracellular matrix protein synthesis observed in both cell culture and aged skin biopsies.^{66,86} Similarly, keratinocytes exposed to AGEs express increased levels of pro-inflammatory mediators, suffer from decreased mobility, and also undergo premature senescence in the presence of AGEs.⁸⁷

In addition to intermediate filaments, proteasomal machinery and DNA can undergo glycation. Proteasomal machinery, which functions to remove altered intracellular proteins, decline functionally *in vitro* when treated with glyoxal.⁸⁸ Similar *in vitro* findings were observed when human epidermal keratinocytes and fibroblasts were treated with glyoxal, leading to accumulation of CML in histones, cleavage of DNA, and, ultimately, arrest of cellular growth.⁸⁹

Beyond the modification of host molecular physicochemistry, AGEs also exert detrimental effects through the binding to specialized cellular surface receptors, called the Receptor for AGEs (RAGE). How cited before, RAGE is a multiligand protein that, when activated, can trigger several cellular signaling pathways, including the mitogen-activated protein kinases (MAPKs), extracellular signal-regulated kinases (ERK), phosphatidylinositol-3-kinase (PI3K), and nuclear factor kappa-beta (NFκ-β) pathways.⁶¹ These pathways are known to mediate various pathogenic mechanisms through the alteration of cell cycle regulators, gene expression, inflammation, and extracellular protein synthesis.⁶¹ Not surprisingly, RAGE is found to be highly expressed in the skin and is present at even higher levels in both UV-exposed anatomical sites and aged skin.⁶⁴

Role of AGEs in the Skin Aging Process

Cutaneous accumulation of AGEs is a feature of skin aging. As mentioned above, AGEs can be directly formed in the organism or be exogenously ingested. Accumulation of AGEs has been detected in various tissues during aging and diabetes, including articular collagen, skeletal and smooth vascular muscles or glomerular basement membranes.⁹⁰⁻⁹² Accordingly, deposited AGEs in these tissues have been implicated in various diabetes or age-associated pathologies such as diabetic angiopathy, age and diabetes-associated macular degeneration and osteoarthritis.^{82,90-95}

Skin, due to its easy accessibility, offers an excellent opportunity for minimal invasive or even non-invasive investigation of glycation, taking advantage of the characteristic autofluorescent properties of AGEs. Accumulation of AGEs in the skin has been therefore thoroughly studied and is detected not only in diabetes as expected but also during chronological aging.^{38,96,97} Glycation-associated skin autofluorescence was shown to correlate with chronological aging in a large number of healthy subjects.⁹⁸

It is a general perception today that AGE accumulation is dependent on protein turnover rate; therefore long-lived proteins are thought to be mainly modified by glycation.⁹⁰ Collagen types I and IV, exhibiting a slow turnover rate of about 10 y, and other dermal long-lived

proteins like fibronectin mainly suffer from glycation during intrinsic chronological aging.^{37,38} The appearance of glycated collagen is first observed at the age of 20. It accumulates with a yearly rate of about 3.7% reaching a 30–50% increase at 80 y of age.^{38,78} CML was recently histochemically detected in human epidermis from healthy donors.³⁶ The upper epidermal layers were mostly involved (stratum spinosum, granulosum and corneum) and the authors identified cytokeratin 10 (CK10) (expressed by differentiated keratinocytes) as a target protein for CML modification. The amount of CML in younger donors seemed to be weak in comparison to the older ones. The latter study had restrictions, as the size of the sample was small and heterogeneous, but indicates a potential involvement of AGEs in epidermal physiology and a possible involvement of more short-lived proteins in glycation chemistry. Moreover, in an *in vitro* reconstructed organ skin model, both epidermis and dermis, as well as their functions, were modified by glycation.⁹⁹ Moreover, smoking, a typical aggravating factor of skin aging, accelerates formation of AGEs and increases their deposition in various tissues including skin.^{83,100} Another important environmental factor for aging is diet. The content of AGEs in food is highly dependent on the method of preparation, like cooking time and temperature. Fried food contains in general far higher amounts of AGEs than boiled or steamed food.¹⁰¹ Dietary AGEs directly correlate with serum levels of AGEs and inflammatory markers in healthy human subjects, respectively.¹⁰²

It has been widely accepted that AGEs, once formed, can be only removed when the modified proteins degrade. However it has now become apparent that in the organism various enzymatic systems seem to be involved in the degradation or removal of AGEs. As mentioned above, Glo I is an enzyme responsible for the removal of reactive α-dicarbonyl compounds. Interestingly, decreased activity of such defense systems against AGEs has been reported during aging.⁶³ These age-related changes may further increase the extent of deposited AGEs in a living organism over time.

Consequences of AGE deposition in skin. AGEs can be formed intracellularly and extracel

lularly. Their presence in biological molecules modifies their biomechanical and functional properties. Proteins, lipids and nucleic acids can be targets of advanced glycation, modifying enzyme-substrate interactions, protein-DNA interactions, protein-protein interactions, DNA regulation and epigenetic modulation, thus interfering with numerous physiological functions of the organism. Moreover, AGEs are themselves reactive molecules which through interaction with their receptors activate various molecular pathways in vivo, thus becoming involved in inflammation, immune response, cell proliferation and gene expression.

1. Extracellular matrix proteins:

Extracellular matrix (ECM) proteins have been regarded as one of the major target structures for glycation. The most abundant collagen type in the skin is type I, whereas collagen IV is being found in the basal membrane. Collagen is one of the strongest proteins. In the skin, it is not only used as a supportive framework for mechanical support for cells and tissues, but represents an active component being able to interact with cells and affect various cellular functions such as migration, differentiation and proliferation.

Collagen glycation impairs its function in various ways. Intermolecular crosslinks of adjacent collagen fibers change its biomechanical properties leading to stiffness and decreased flexibility, thus increasing its susceptibility to mechanical stimuli.⁷⁹ The change of its charge and the formation of AGEs on side chains of collagen affect its contact sites with cells and other matrix proteins and inhibit its ability to react with them.⁸⁰ The precise aggregation of monomers into the triple helix may be affected as well as the association of collagen IV with laminin in the basal membrane.⁴⁰ Modified collagen resists degradation by MMPs, thus inhibiting its removal and replacement by newly synthesized and functional one.⁸² Accordingly, tissue permeability and turnover is impaired.^{40,103}

Other extracellular matrix proteins suffering from advanced glycation are elastin and fibronectin, contributing further to dermal dysfunction.^{37,38,43} Of note, CMLmodified elastin has been found almost exclusively in sites of actinic elastosis and not in sun-protected skin, underlining its potential role in photoaging. Indeed,

UV irradiation stimulates glycation of elastin in the presence of sugars. Moreover, CMLmodified elastin assembled in large and irregular structures, has decreased elasticity and is resistant to proteolytic degradation.⁸⁴

It has been shown that in vitro glycated skin samples have impaired biomechanical properties.¹⁰⁴ In vivo, decreased skin elasticity characterizes diabetic subjects in comparison to healthy controls.¹⁰⁵

2. Intracellular proteins

Intermediate filaments such as vimentin in fibroblasts and CK10 in keratinocytes have been found to be modified by AGEs.^{36,42} Cytoskeletal proteins are important in providing stability of the cytoskeleton and are crucially involved in numerous cellular functions such as migration and cellular division. Various other intracellular proteins including enzymes and growth factors may be targets of non-enzymatic modification by sugars. Glycated basic fibroblast growth factor (bFGF) displays impaired mitogenic activity in endothelial cells.¹⁰⁶ Glycation of enzymes of the ubiquitin-proteasome system and of the lysosomal proteolytic system has been shown to inhibit their action.¹⁰⁷ Antioxidant and other protective enzymes such as Cu-Zn-SOD can be inactivated.¹⁰⁸ Other intracellular components, such as DNA and lipids can be glycated with detrimental effects on their function.³²

3. Receptors for AGEs: RAGE.

AGEs do not only act by altering the physicochemical properties of glycated proteins. As mentioned above, AGEs may bind to their cell surface receptor, RAGE, initiating a cascade of signals influencing cell cycle and proliferation, gene expression, inflammation and extracellular matrix synthesis (reviewed in Bierhaus et al.).⁶¹ Interestingly, RAGE is broadly expressed in human skin and in epidermal keratinocytes, dermal fibroblasts and endothelial cells in vitro. It is highly found in sites of solar elastosis, and its expression is induced by advanced glycation end products and proinflammatory cytokines like TNF α .⁶⁴ In skin cells RAGE has been shown to decrease cell proliferation, induce apoptosis and increase MMPs production. Many of these effects involve NF κ B signaling.⁶⁶

4. Effects of AGEs on resident skin cells:

AGEs have been shown to affect various functions of skin cells in vitro. They decrease proliferation and enhance apoptosis of human dermal fibroblasts, an effect which is at least partly RAGE dependent and correlates with the activation of NF κ B and caspases.⁸⁶ In keratinocytes, AGEs decrease cell viability and migration and induce the expression of proinflammatory mediators.⁸⁷ Moreover, AGEs are able to induce premature senescence in human dermal fibroblasts and in normal human keratinocytes in vitro.^{109,110} Collagen and ECM protein synthesis have been also found to be decreased, while the expression of MMPs is induced.⁶⁶ Dicarboxyls such as glyoxal and methylglyoxal impair the signaling of epidermal growth factor receptor (EGFR), a receptor controlling various cellular functions such as proliferation, differentiation, motility and survival, by formation of EGFR crosslinks, blocking of phosphorylation and impaired activation of ERKs and phospholipase C.¹¹¹ Various other growth factors or proteins significant for cellular functions, like bFGF, may be glycosylated inhibiting their functions.¹⁰⁶ In the context of extrinsic aging, AGEs seem to render cells more sensitive to external stimuli, as UVA irradiated fibroblasts and keratinocytes exhibit decreased viability after exposure to AGEs.¹¹²

5. The role of oxidative stress:

Oxidative stress has been widely accepted to mediate the deleterious effects of solar radiation in the skin during photoaging. Interestingly, in vitro exposure of AGEs to UVA irradiation leads to formation of ROS, such as superoxide anion, hydrogen peroxide and hydroxyl radicals.⁸⁵ AGEs can lead to ROS formation in cells by various ways. They can stimulate NOX to induce production of superoxide anion or they can compromise cellular antioxidant defense systems, e.g. inactivation of Cu-Zn-SOD by cross-linking and site-specific fragmentation of this molecule.¹⁰⁸ Moreover, AGEs are themselves very reactive molecules. As early as during their crosslinking reactions they can act as electron donors leading to formation of superoxide anions.¹¹³ Glycation of proteins creates active enzyme-like centers (cation-radical sites of crosslinked proteins) able to catalyze one-electron oxidation-reduction reactions leading

to ROS generation with or without presence of oxygen or transition metals such as iron and copper.¹¹³⁻¹¹⁵ Finally, autofluorescent AGEs, such as pentosidine, can act as endogenous photosensitizers leading to increased ROS formation after UVA irradiation of human skin. UV irradiation of human keratinocytes and fibroblasts in the presence of AGEs led to increased ROS formation and decreased proliferation in vitro.¹¹²

6. Skin AGEs as biomarkers of aging:

As AGEs have been etiologically implicated in aging and aging-related pathologies, the idea of using them as biomarkers is appealing. AGEs in the skin have been initially measured by western blots (WB) with polyclonal antibodies or by autofluorescence measurements of skin biopsies, thus restricting the wide use of these measurements. An AGE-Reader (DiagnOptics B.V., Groningen, The Netherlands) has been introduced some years ago as a new, noninvasive method to measure in vivo the skin content of AGEs based on their characteristic autofluorescence.¹¹⁶⁻¹¹⁸ Until now it has been shown that skin autofluorescence positively correlates with various diabetes- and age-related complications such as micro and macrovascular complications, renal disease, cardiovascular events, overall mortality, age-related macular degeneration and chronic renal disease.^{117,119,120} Skin glycation has been proposed as a prognostic factor for the development of diabetic complications.¹²¹ Lately it was shown that skin autofluorescence increases with chronological aging and correlates with skin deposition of AGEs, making this method a potential tool in investigating the effect of various anti-aging products of the cosmetic industry.¹²²

Dietary advanced glycation end-products (d-AGEs)

A large database of different food items and their AGE contents has been created by measuring CML with ELISA.^{18,101} In general the reported CML contents are correlated with corresponding levels of MG-derivatives.¹⁸ AGE content of foods as determined by CML and MG levels shows a highly significant linear correlation ($r=0.8$, $P=0.0001$) prepared by different cooking techniques. The highly significant internal correlation between two chemically distinct AGEs (CML and MG) in a variety of foods

prepared by different methods validates the methodology applied and supports the choice of CML levels as a useful marker of d-AGE content.

As with CML, foods high in protein and fat contained higher amounts of MG than did carbohydrate-rich foods. Recent studies indicate that the meat group contains the highest levels of AGEs because meats are served in larger portions as compared to fats which tend to contain more dAGE per gram of weight. When items in the meat category prepared by similar methods were compared, the highest dAGE levels were observed in beef and cheeses followed by poultry, pork, fish, and eggs. Lamb ranked relatively low in dAGEs compared to other meats.

Higher-fat and aged cheeses, such as full-fat American and Parmesan, contained more dAGEs than lower-fat cheeses, such as reduced-fat mozzarella, 2% milk cheddar, and cottage cheese. Whereas cooking is known to drive the generation of new AGEs in foods, it is interesting to note that even uncooked, animal-derived foods such as cheeses can contain large amounts of dAGEs. This is likely due to pasteurization and/or holding times at ambient room temperatures (e.g., as in curing or aging processes). Glycation-oxidation reactions, although at a slower rate, continue to occur over time even at cool temperatures, resulting in large accumulation of dAGEs in the long term. High-fat spreads, including butter, cream cheese, margarine, and mayonnaise, was also among the foods highest in dAGEs, followed by oils and nuts. As with certain cheeses, butter and different types of oils are AGE-rich, even in their uncooked forms. This may be due to various extraction and purification procedures involving heat, in combination with air and dry conditions, however mild they are. The type of cooking fat used for cooking led to the production of different amounts of dAGEs.¹⁶

In comparison to the meat and fat groups, the carbohydrate group generally contained lower amounts of AGEs due to the higher water content or higher level of antioxidants and vitamins in these foods, which may diminish new AGE formation. The highest dAGE level per gram of food in this category was found in dry-heat processed foods such as crackers, chips, and cookies.

This is likely due to the addition of ingredients such as butter, oil, cheese, eggs, and nuts, which during dry-heat processing substantially accelerate dAGE generation. Although AGEs in these snack types of food remain far below those present in meats, they may represent an important health hazard for people who consume multiple snacks during the day or as fast meals.¹²³

Grains, legumes, breads, vegetables, fruits, and milk were among the lowest items in dAGE, unless prepared with added fats. For instance, biscuits had more than 10 times the amount of dAGEs found in low-fat breads, rolls, or bagels.¹⁸ Nonfat milk had significantly lower dAGEs than whole milk. Whereas heating increased the dAGE content of milk, the values were modest and remained low relative to those of cheeses. Likewise, milk-related products with a high moisture index such as yogurt, pudding, and ice cream were also relatively low in AGEs.¹⁸

Factors affecting the rate of dietary AGEs (d-AGEs) formation during cooking

The rate of formation and the diversity of the generated AGEs in food depend on factors such as composition, availability of precursors, presence of transition metals, and availability of pro and antioxidants. Reaction time, processing temperature, concentrations of reactants, availability of water, and pH are particularly well known to have a decisive effect on the rate of the Maillard reaction.¹²⁴ As a rule of thumb, the rate of the Maillard reaction at least doubles when the temperature is increased by 10°C. If browning is used to measure the progress of the Maillard reaction, then four weeks at 20 °C, 3 h at 100 °C, and 5 min at 150 °C give approximately the same result.¹²⁵ Factors like pH^{126,127} and water activity greatly affect the rate of formation of Maillard reaction products (MRPs).

The rate of the Maillard reaction is considered to be low at acidic pH, but increases with increasing pH until a maximum is reached around pH 10.²¹ At higher moisture levels, a decrease in reaction rate is observed due to dilution of the reactants in the aqueous phase. Water is a product of the reaction and it is probable that the law of mass action also leads to a decreased rate of reaction at high moisture levels.¹²⁸ Dry heat cooking has been found to promote formation of dietary AGEs as

determined by immunological methods. However, AGE formation seems to be reduced by heating in an oven at high humidity, shorter cooking times, lower cooking temperatures, or by the use of acidic ingredients, such as lemon juice or vinegar.¹⁶

Absorption and bioavailability

Early animal studies reported that MRPs are at least partially absorbed, and those low molecular weights (LMW) MRPs are absorbed to a higher degree than high molecular weight (HMW) MRPs.¹²⁹ The absorption of AGEs into the circulation in humans measured by a non-specific ELISA method was estimated to be about 10 % of ingested AGEs.¹³⁰ HMWAGEs need to be degraded by gut proteases before the LMW products are liberated. The bioavailability of the partially degraded HMW AGEs will depend on the size of the associated peptide, type of diet, gut environment, and duration of their presence in the gut. Heat-induced changes in proteins can decrease their susceptibility to degradation by gastrointestinal enzymes, and protein and mineral bioavailability have been shown to be influenced negatively by a heat-treated diet.¹³¹⁻¹³³ Oral bioavailability is thought to be low (10 %), secondary to poor absorption from the gastrointestinal tract, as AGE cross link formation is resistant to enzymatic or chemical hydrolysis.¹³⁰ The water solubility and amphoteric properties makes LMWAGEs to be absorbed to extracellular and intracellular compartments than HMWAGEs.

The in vivo distribution of CML and CEL after an intravenous injection in rats showed a temporary accumulation in the liver,¹³⁴ indicating that they may have high affinity to some specific hepatic proteins. In the study of ¹⁴C labeled AGEs, it was observed that 60 % of the absorbed AGEs were bound in liver and kidney after 72 h, but radioactivity was also observed in lung, heart, and spleen indicating more global distribution and tissue binding.¹³⁵ Several animal studies have shown a correspondence between dietary AGE content and serum and tissue AGE levels.^{136,137} Any deterioration in renal function results in AGE accumulation which can lead to endothelial perturbation and hence vascular disease.¹³⁸ In vitro studies have proposed that insulin also contributes to

AGE elimination from the plasma via the IRS and phosphatidylinositol-3-OH kinase (PI3 kinase) pathway.¹³⁹ This pathway is thought to be vasculo-protective, leading to a rise in nitric oxide as well as facilitating insulin-mediated glucose transport in adipocytes and skeletal muscle. Recent human studies revealed that about 10 % of diet-derived AGEs were absorbed, two-thirds of which remained in the body and only one-third of the absorbed AGEs was excreted into the urine within 3 days from ingestion.^{16,130,135}

Dietary advanced glycation end-products (d-AGEs) and their health implications

Nutrient composition, temperature and method of cooking can affect the formation of AGEs in foods. Fats or meat-derived products processed by high heat such as broiling and oven frying contain more AGEs than carbohydrates boiled for longer periods.^{101,140} That is, in the absence of lipids and proteins or heat, sugar content does not necessarily correlate with AGE values in the food. And, the absence of sugars does not necessarily predict low AGE content, as in preparations containing preformed AGE-like caramel additives.¹³⁰

Food-derived AGEs induce protein cross-linking and intracellular oxidant stress similar to their endogenous counterparts when tested in vitro using human-derived endothelial cells.¹⁴¹ These pro oxidant and pro inflammatory properties are also found in the circulating AGE fractions derived from these exogenous AGEs. Experiments performed in different animal models have established a significant role for dietary AGEs in inducing type 1 diabetes mellitus in nonobese diabetic (NOD).¹³⁶ In a group of diabetic subjects, dietary AGE restriction was associated with significant reduction of two markers of inflammation, plasma C reactive protein (CRP) and peripheral mononuclear cell TNF- α , as well as of VCAM-1, a marker of endothelial dysfunction.⁹³ These observations were later extended to chronic renal failure patients on maintenance peritoneal dialysis, in whom dietary AGE restriction was associated with a parallel reduction of serum AGEs and CRP.¹⁴² The parallel changes of serum AGEs and CRP following dietary AGE modifications are highly suggestive of a role for dietary AGEs in inducing inflammation.¹⁶

Role of food-derived AGEs in vascular complications in diabetic animals

With regards to complications of diabetes, several different animal models have been used to examine the role of dietary AGEs in the development of kidney disease. In diabetic mouse models, there has been reports of both protective¹⁴³ and disparate effects¹⁴⁴ of diets low in AGEs in development of diabetic nephropathy. In remnant kidney models in rats, proteinuria increased during feeding with high AGE diets^{145,146} Furthermore, high AGE diets were shown to accelerate progression of renal fibrosis.¹⁴⁵ In addition, in a mouse model of obesity, renal impairment developed when high AGEs and a high fat diet were combined. An AGE-poor diet that contained four- to five-fold lower AGE contents for 2 months also decreased serum levels of AGEs and markedly reduced tissue AGEs and RAGE expression, numbers of inflammatory cells, tissue factor, VCAM-1, and MCP-1 levels in diabetic apolipoprotein E deficient mice.¹⁴⁸

Role of food-derived AGEs in ageing

Ageing is associated with increased oxidative stress generation and AGE formation.⁷² A life-long restriction of AGE containing diet reduces oxidative stress generation and AGE accumulation which are associated with RAGE and p66 suppression, resulting in extension of lifespan in mice.⁷² Oral intake of AGE containing foods also determines the effects of calorie restriction on oxidant stress, age related diseases, and lifespan.¹⁴⁹ These observations suggest that restriction of AGE-rich diet may be a novel therapeutic target for prevention of age associated various disorders.

In food analyses, CML has been the most widely used marker for AGEs.¹⁵⁰ The CML content of the same food item can be increased up to 200-fold by increasing the temperature and conditions used in cooking. The CML concentrations of various foods vary widely from about 0.35–0.37 mg CML/kg food for pasteurized skimmed milk and butter to about 11 mg CML/kg food for fried minced beef and 37 mg CML/kg food for white bread crust. Fried meat, sausage, and cookies are high in CML.¹⁵¹ Other foods that are high in AGEs include many commercial breakfast cereals,¹⁵² roasted nuts and seeds,¹⁵³ ice cream,¹⁵⁴ and barbecue sauces.¹⁵⁵ High concentrations of methylglyoxal,

an intermediate product of the Maillard reaction, are found in commercial soft drinks that contain high fructose corn syrup.¹⁴⁴ Methylglyoxal is reactive and readily modifies lysine or arginine residues of proteins to form carboxyethyllysine and hydroimidazolones. Pasteurized milk and sterilized milk contain much higher CML concentrations than raw milk.³² Evaporated whole milk contains high concentrations of CML, probably due to the high temperatures used in processing the milk. Infant formula contains high concentrations of AGEs.¹⁵⁶ Commercial infant formulas contain a 70- fold higher level of CML than human breast milk, and infants fed infant formula had significantly high plasma CML than breast-fed infants.¹⁵⁷ Foods that are either eaten raw or cooked at lower temperatures are relatively low in AGEs, and such foods include raw fruits and vegetables, raw fish, raw nuts, yoghurt, tofu, pasta, boiled rice, boiled potatoes, and other boiled or simmered foods.

Other processes, besides the formation of AGEs, also take place in food during cooking. It is well-known and described in the literature that heating of food induces degradation and oxidation of heat-sensitive compounds, including vitamins and other bioactive compounds.^{158-160A} high versus low AGE diet made by differences in heat treatment will, therefore, have dissimilar content of such compounds and this has also been confirmed when it has been measured in intervention studies.¹⁶¹ This is a problem, because effects of high AGE diets cannot be directly related only to the AGE content. It cannot be ruled out that a lower content of a range of heat-sensitive nutrients in the diet, e.g., vitamin C, E, and thiamine, could also contribute to these negative effects. Accordingly, AGE levels in body fluids might be markers of the inflammatory and oxidative burden. For example, marginal thiamine deficiency has been shown to increase both markers of oxidative stress and of reactive dicarbonyls¹⁶², and vitamin B6 can also affect AGE formation. Furthermore, extensive heat processing of food can generate Maillard-derived antinutritional and toxic compounds^{164,165} Such compounds include acrylamide,^{166,167} heterocyclic aromatic amines¹⁶⁸ and 5-hydroxymethylfurfural,¹⁶⁹ all of which are suspected carcinogens. Thus,

simply referring the effects of a less heat-treated diet to effects of AGEs is problematic; the consequences of cooking for the concentrations of AGEs as well as other heat-derived compounds are not tested in the majority of the dietary AGE studies. Only one study has reported the content of acrylamide and 5 hydroxymethylfurfural and they were found to be significantly higher in the high AGE diet.¹⁷⁰

Nevertheless, this shows there is a large range of potentially harmful compounds generated by heat and points to the essential problem with identifying the active compounds. Harmful effects of high AGE diets cannot be directly related to the AGE content. Studies with well-defined compounds outside a complex food matrix (e.g., synthetically produced AGEs) are needed to identify individual effects. Moreover, AGEs are often investigated and discussed as a whole, even though they are a large and heterogeneous group of compounds. The heterogeneity of AGEs makes it difficult to conclude which of these compounds are biologically active and exert which specific effects in vivo. Within the large range of MRPs, not only AGEs have been identified, but also compounds with potential beneficial effects have been described. Melanoidins have been associated with health benefits in some studies and antioxidative properties of MRPs have been observed in a human intervention study.^{16,28}

Anti-AGE Strategies: Current Knowledge and Future Perspectives

Since the emergence of AGEs as an important pathogenetic factor in diabetes and aging the development of strategies against AGEs has been in the center of scientific interest. Substances able to prevent or inhibit formation of AGEs, as well as agents able to break already formed AGEs or those antagonizing their signaling have been identified. Some of them are already being tested in clinical trials.^{171,172}

1. Substances preventing or inhibiting AGE formation.

Aminoguanidine was one of the first substances identified limiting the formation of AGEs.¹⁷³ Aminoguanidine is a nucleophilic hydrazine and its anti-AGE properties result from trapping of early glycation products such as carbonyl intermediate compounds.

It has no effects on more advanced stages of glycation. Despite its potential effects in attenuating various diabetes- and age-related complications in animal models, its use in clinical practice is limited due to adverse effects in clinical trials with diabetic patients.¹⁷⁴ In an in vitro skin aging model it could attenuate collagen glycation, however its effects against AGE induced collagen modification in vivo have been contradictory.¹⁷⁵⁻¹⁷⁷ Studies on topical application of aminoguanidine in the skin are lacking.

Different AGE inhibitors suppress AGE formation at different stages of glycation. For example, aspirin (acetylsalicylic acid) is known to inhibit glycation by acetylating free amino groups of a protein, thereby blocking the attachment of reducing sugars^{178,179} at the early stage of the glycation process. The inhibitory activities against AGE formation of various vitamin B1 and B6 derivatives such as pyridoxamine^{180,181,182} and thiamine pyrophosphate¹⁸³ have mainly been attributed to their abilities to scavenge reactive carbonyl compounds.^{32,182}

Pyridoxamine, a naturally occurring vitamin B6 isoform, seems to be another tool in the fight against AGEs. Pyridoxamine traps reactive carbonyl intermediates, scavenges ROS and in addition inhibits post-Amadori stages of AGE formation.¹⁸⁴ It has shown promising results in a phase II clinical trial against diabetic nephropathy.¹⁸⁵ Oral intake of pyridoxamine resulted in potent inhibition of skin collagen CML formation in diabetic rats. In addition, penicillamine could reduce the level of AGEs through decreasing the formation of Amadori products.^{177,186,187} However, its potential against skin aging remains to be shown.

2. "AGE breakers."

Chemical substances and enzymes able to recognize and break the Maillard reaction crosslinks have been identified. Such chemical AGE breakers are dimethyl-3phenacyl-thiazolium chloride (ALT-711), N-phenacylthiazolium and N-phenacyl-4,5-dimethylthiazolium.¹⁸⁵ They have been developed to chemically break the prototypical Maillard reaction crosslink via a thiazolium structure.¹⁸⁵ Promising results against cardiovascular complications in diabetes and aging have been reported, although their actual ability to cleave

existing protein crosslinks in tissues has been questioned.¹⁸⁶⁻¹⁸⁹

Interference with intrinsic AGE-de-toxifying enzymes like FAOXs, FN3K and the enzymatic system of Glo is another interesting strategy to remove AGEs, as enzymes recognize specific substrates and may be associated with fewer side effects.^{58,190,191} There are a lot of data supporting the significance of these enzyme systems in aging. As noted above decreased Glo I activity and increased accumulation of AGEs with age have been shown in many tissues and animals.⁵⁷ Overexpression of Glo I significantly inhibits hyperglycemia-induced intracellular formation of AGEs in bovine aortic endothelial cells and in mouse mesangial cells by reduction of intracellular oxidative stress and apoptosis.^{192,193} A potential in vivo beneficial effect of Glo I against AGEs could be also shown in transgenic rats.¹⁹⁴ Interestingly, it has been recently shown that Glo I is transcriptionally controlled by Nrf2, and that pharmacological Nrf2 activators increase Glo I mRNA and protein levels as well as its activity.⁵⁷ The pharmacological induction of such enzymes could represent a novel future strategy against AGEs. Fructosamine phosphokinases are relatively new enzymes and currently under investigation, and until now no inductors or activators of their expression have been found.⁴⁰ FAOXs, on the other hand, are not expressed in mammals, and their potential use in humans by enzymatic engineering remains to be discovered.⁵⁹

3. Nutraceuticals.

Since oxidation steps are crucially involved in formation of many AGEs, substances with antioxidative or metal chelating properties, may also have antiglycating activities.¹⁹⁵ Thus, a lot of interest has been directed to nutrients and vitamins, so called "nutraceuticals," as natural tools against AGEs.^{172,196}

Accordingly, an increasing list of natural antioxidants and chelating agents such as ascorbic acid, α -tocopherol, niacinamide, pyridoxal, sodium selenite, selenium yeast, trolox, rivoflavin, zinc and manganese has been shown to inhibit glycation of albumin in vitro.¹⁹⁷ Alpha-lipoic acid was able to reverse tail tendon collagen glycation in fructose-fed rats, an effect which was attributed to its endogenous antioxidant action, its ability to recycle

ascorbic acid, α -tocopherol and GSH as well as to its positive influence on glucose uptake and glycaemia.¹⁹⁸ Green tea, vitamins C and E and a combination of N-acetylcystein with taurine and oxerutin could inhibit skin collagen glycation in mice.^{196,199} Another compound, the green tea-derived polyphenol and flavonoid epigallocatechin-3-gallate revealed also promising in vitro effects by antagonizing AGE-induced proinflammatory changes.²⁰⁰ In healthy human subjects, supplementation of vitamin C significantly decreased serum protein glycation.²⁰¹ Many spices and herbs were shown to inhibit glycation of albumin in vitro, among them ginger, cinnamon, cloves, marjoram, rosemary and tarragon.²⁰² Their protective effects correlated with their phenolic content. Recently, in vivo beneficial effects of some of these compounds were shown in zebrafish.²⁰³ Other promising compounds include blueberry extract and naturally occurring flavonoids, such as luteolin, quercetin and rutin, which can inhibit various stages of AGE formation.^{204,205} Blueberry extract, an AGE-inhibitor and C-xyloside, a glycosaminoglycan synthesis stimulator, were tested for 12 weeks in female diabetic subjects. This treatment resulted in significant improvement of skin firmness, wrinkles and hydration although it failed to show a significant decrease in the cutaneous content of AGEs.²⁰⁴ In one of the few human studies successfully conducted on anti-AGE therapeutics, L-carnitine supplementation for 6 months in hemodialysis patients significantly decreased levels of AGEs in the skin.²⁰⁶ L-carnitine, which is naturally abundant in meat, poultry, fish, and dairy products, is an antioxidant. Furthermore, it may function synergistically to neutralize oxidative stress when given with α -lipoic acid.²⁰⁷

As a well-known nutraceutical product, grape seed extract (GSE) is an abundant source of catechins and proanthocyanidins with a strong antioxidant and free radical scavenging activity.²⁰⁸ Peng et al. (2010) studied the effects of GSE on the formation of N ϵ - (carboxy-methyl) lysine (CML) in bread. Besides introducing antioxidant activity to bread, GSE also appeared to attenuate CML content in bread crust. In particular, adding 600 and 1000 mg of GSE to bread (500 g) led to over 30 % and 50 % reduction in bread crust CML content, respectively.

Strong antioxidant activities of catechins and proanthocyanidins abundant in GSE may contribute to the reduction of CML in GSE-fortified bread.²⁰⁹ On the other hand, catechins and proanthocyanidins proved to be able to scavenge the intermediate dicarbonyls (such as methylglyoxal, glyoxal)^{210,211} in the glycation process, which may also decrease the CML content of GSE-fortified bread.

4. Caloric restriction and dietary measures

As nutrition is an important factor in skin aging, dietary caloric restriction may be effective in preventing accumulation of AGEs in the human body. In mice restriction of caloric intake increases lifespan and delays many age-related dysfunctions by altering stress response and influencing the expression of various metabolic and biosynthetic genes.²¹² Dietary restriction could significantly decrease the levels of AGEs in rat and mice skin collagen.^{213,214} Skin collagen glycation and glyoxidation inversely correlated with lifespan whereas caloric restriction led to decreased accumulation of AGEs and increased lifespan.²¹⁵ Dietary restriction may not be a pragmatic option in humans; however a restriction in intake of dietary “glycotoxins” may be more feasible. As outlined above these dietary glycotoxins derive from nutrition. In humans dietary glycotoxins significantly increase concentrations of systemic inflammatory mediators like TNF α , interleukin (IL)-6 and C-reactive protein and are thus considered as diabetogenic, nephrotoxic and proatherogenic.^{93,216} Dietary intake of AGEs correlates with serum AGEs and can induce systemic oxidative stress, increase RAGE expression, decrease antioxidant levels and shorten lifespan in mice.¹⁴⁹ A diet with a low content in AGEs could reduce circulating AGEs and inflammatory biomarkers in patients with diabetes and renal failure thus seeming to be an important supportive therapy in diabetes.^{217,218} In mice low dietary AGEs had beneficial effects in wound healing and other diabetes mellitus-associated pathologies.¹³⁶ There are no studies investigating the effects of AGE-poor diets on skin aging in humans. However, it has been shown that skin collagen glycation positively correlates with blood glucose levels in diabetes and that intensive treatment can reduce the levels of skin glycation,

implicating that a diet low in AGEs may have a beneficial effect on skin glycation.^{219,220}

5. Targeting RAGE.

Another potential strategy against excessive accumulation of AGEs could be the antagonism of RAGE.²²¹ Possible approaches include gene knock-down of RAGE by siRNA or anti-sense and antagonism of RAGE with putative small molecular inhibitors against RAGE-induced signaling.^{68,221} Promising effects in various systems have been shown in vitro and in vivo with neutralizing anti-RAGE antibodies.⁶¹ Since serum concentrations of sRAGE negatively correlate with AGE-induced pathologies, neutralization of AGEs by these decoy receptors of RAGE may be considered as another anti-AGE strategy. Potential protective effects of sRAGE have been shown in various diabetes and inflammatory models.^{61,63,64,222} Interestingly, sRAGE could also attenuate impaired wound healing in diabetic mice. Therefore, studies will be needed to investigate an analogous effect on skin aging.²²³

6. Others

Molecular chaperones like carnosine have lately shown promise in improving skin appearance in various studies at least in part by reducing the amounts of skin AGEs.²²⁴⁻²²⁶

Combating AGE with Diet

Nearly 70 years ago, Urbach and Lentz reported that the level of sugar both in the blood and in the skin is decreased with a diet low in sugar.²²⁷ Although its significance was not appreciated at the time, this finding demonstrated a quintessential connection between diet and skin health. We now understand that food is a source of both monosaccharides that, in high amounts, catalyze the production of AGEs in the body, and preformed AGEs.²²⁸

Preformed AGEs are absorbed by the gut with approximately 10-0% efficiency. They can then enter the circulation, where they may induce protein cross-linking, inflammation, and intracellular oxidative stress. The end result is the amplification of a similar “vicious cycle”, which may be as detrimental as the consumption of excess dietary sugar.²²⁹ Interestingly, preformed AGEs largely result from exogenous synthesis mediated by the food cooking process. Grilling, frying, deep fat frying,

and roasting methods are all known to produce higher levels of AGEs in food. In contrast, methods of preparation that are water-based, such as boiling and steaming, produce a logarithmically lower amount of AGEs.²¹

A diet low in AGEs correlated with a reduction in inflammatory biomarkers (i.e., tumor necrosis factor-alpha, interleukin-6, and C-reactive protein) in diabetic human patients, as well as an improvement in wound healing and other diabetes-associated sequelae in mice.^{136,218} Other authors have cited the relatively youthful appearance that is often associated with the elderly Asian population as evidence of the long-term impact of employing water-based cooking practices, which are characteristic of Asian cooking.²²⁸

The varying conditions of water and heat play a significant role in the production of dAGE content. As scrambled eggs prepared in an open pan over medium-low heat had about one half the dAGEs of eggs prepared in the same way but over high heat. Similarly, poached or steamed chicken had less than one fourth the dAGEs of roasted or broiled chicken. Thus, frying, broiling, grilling and roasting yielded more dAGEs compared to boiling, poaching, stewing, and steaming. Moreover, microwaving also did not raise dAGE content to the same extent as other dry heat cooking methods for the relatively short cooking times (6 min or less) that were tested. In nut shell, higher temperature and lower moisture levels coincided with higher dAGE levels.¹⁸ Tight glycemic control over a 4-month period can result in a reduction of glycated collagen formation by 25%.^{228,229} Consumption of a low-sugar diet prepared through waterbased cooking methods would limit both the consumption of preformed exogenous AGEs and endogenous production through physiological glycation. Avoiding foods that result in higher levels of AGEs, such as donuts, barbecued meats, and dark-colored soft drinks, can be an effective strategy for slowing "sugar sag."

²¹ Beans are recommended as suitable foods for diabetic patients in the past mainly for their high fibre and protein contents. Four kinds of beans including mung bean (*Vigna radiata*) black bean (*Phaseolus vulgaris* L.), soybean (*Glycine max*) and cowpea (*Vigna unguiculata*) were investigated for trapping of methylglyoxal,

a key intermediate compound for the formation of AGEs. The aqueous alcohol extracts of all beans examined have showed significant inhibitory activities at a concentration of 500 ppm with 80.4 % inhibition for mung bean, 72.1 % for black bean, 70.1 % for soybean, and 67.3 % for cowpea extract, respectively.²¹⁰ Various phenolic antioxidants from plant extracts have been found to inhibit the formation of AGEs, and their inhibition of free radical generation in the glycation process and subsequent inhibition of modification of proteins have been considered as the major mechanisms for mediating their anti-glycation activities. Total phenolics were determined and it was found that mung bean extract had the highest phenolic content and anti-glycation activities of these beans were highly correlated with their total phenolic contents ($R^2=0.95$). Two major phenolic compounds from mung bean, vitexin and isovitexin were studied for their activities in direct reaping of methylglyoxal, a key intermediate compound for the formation of AGEs.²¹⁰

Low or acidic pH also arrests the new AGE development. For example, beef that was marinated for 1 h in lemon juice or vinegar formed less than half the amount of AGEs during cooking than the untreated samples.¹⁸ Green tea is known well for diabetic people in several ways. It reduces blood glucose level; improves sensitivity to insulin and enhances antioxidant defenses.^{230,231} Furthermore, green tea inhibits the formation of AGEs in an in vitro bovine serum albumin (BSA)/glucose system and in the collagen of aged rats and diabetic rats.

Of interest, several culinary herbs and spices are believed to be capable of inhibiting the endogenous production of AGEs (specifically fructose-induced glycation). These include cinnamon, cloves, oregano, and allspice.^{197,198,202} Other dietary compounds that have been linked to inhibition of AGE formation based on in vitro data and preliminary animal models include ginger, garlic, α -lipoic acid, carnitine, taurine, carnosine, flavonoids (e.g., green tea catechins), benfotiamine, α -tocopherol, niacinamide, pyridoxal, sodium selenite, selenium yeast, riboflavin, zinc, and manganese.^{197,198,202} The cosmetic industry has taken notice of this data, and several have recently released topical products containing carnosine and α -lipoic acid,

with claims related to anti-AGE formation.²²⁹ However, data is lacking as to whether topical administration of these compounds is as effective as dietary delivery in slowing the aging process.

Since glycation is accelerated in the presence of reactive oxygen species, antioxidants should theoretically be effective in limiting the production of new AGEs. They may also impact AGE-induced tissue damage. One intriguing study looked at the effects of the antioxidant resveratrol. Popularly known for its abundance in red wine, resveratrol is a natural phenol produced by several plants in response to injury and is found in the skin of grapes, blueberries, raspberries, and mulberries. In one study, resveratrol inhibited AGE-induced proliferation and collagen synthesis activity in vascular smooth muscle cells belonging to stroke-prone rats.²³² Another study found that it decreased the frequency of DNA breaks in methylglyoxal treated mouse oocytes. Although resveratrol does not appear to reverse the glycation process itself, these studies suggest that it can reduce AGE-induced tissue damage.²³³ While these findings are promising, to our knowledge these laboratory results have not yet been demonstrated in human studies.

Numerous traditional herbal infusions, including Luobuma (*Apocynum venetum* L.), Nagarmotha (*Cyperus rotundus*), Mate (*Ilex paraguariensis*) and Guava (*Psidium guajava* L.) exhibit potent anti-glycation capacities.²³⁴⁻²³⁷ All herbal infusions inhibited the glucose-mediated formation of fluorescent AGEs in a dose-dependent manner at dilutions of 10-fold to 40-fold. At a ten-fold dilution, balm, mint, black tea, green tea and sage almost completely inhibited the formation of fluorescent AGEs. At a 20-fold dilution, only balm retained its capacity to inhibit totally the formation of fluorescent AGEs. Accordingly, comparing the antiglycation capacities of different herbal infusions based on the experimental results obtained from a 40-fold dilution seems logical. At a 40-fold dilution, the anti-glycation capacity of herbal infusions followed the order, balm (89.8 %)>mint (47.8 %)>black tea (38.0 %)>green tea (35.4 %), sage (33.4 %) and common verbena (30.4 %)>rosemary (18.8 %)>lemongrass(3.0 %).²³⁸

Based on the current evidence, individuals with diabetes and/ or kidney disease seem to be the population groups deriving most benefit from an AGE-restricted diet and potentially from inhibition of AGE-formation and its associated actions in the body.^{16,196,230,239}

How to win the battle against AGEs/ fight against AGEs in kitchen

As modern diets are largely heat processed, they are more prone to contain high levels of advanced glycation end products (AGEs).¹⁶ On an average, the intake of dAGE in a cohort of healthy adults from the New York city areas was found to be 14,700 ±680 AGE kU/day.¹⁰² By smart food selection and by changing the way of cooking, the level of AGEs could be lowered in the diet. Overall, moving away from foods high in fat, red meat and processed and fast foods and toward a diet focused more on fruits and vegetables, whole grains and lean meats and fish will not only reduce the AGE intake but help to meet other important nutritional goals as well.

Reducing dAGE may be especially important for people with diabetes, who generate more endogenous AGEs than those without diabetes and for those with renal disease, who have impaired AGE clearance from the body.¹³⁰ Recently there has been heightened interest in therapeutic diets that are higher in protein and fat and lower in carbohydrate for weight loss, diabetes and cardiovascular diseases. This type of dietary pattern may substantially raise dAGE intake and thus contribute to health problems over the long term. A safe and optimal dAGE intake for the purposes of disease prevention has yet to be established.

Some tips to win the battle against AGEs in kitchen are as:

- Use of lower cooking temperatures over high cooking temperatures;
- Steaming, stewing and poaching are better cooking methods than frying, grilling and roasting;
- Be wary of browning;
- Higher temperature and lower moisture levels in food during cooking increase dAGE levels;
- Phenolic antioxidants (e.g., in beans) can inhibit the formation of AGEs;

- Addition of acids (e.g., vinegar, lemon juice) lowers AGE levels;
- Green tea inhibits formation of AGEs;
- Cook fresh foods as possible;
- Eat more often at home.¹⁶

Conclusion

Current evidence from many different disciplines lends strong support to the idea that AGEs contribute to the multisystem decline that occurs with aging. AGEs contribute to inflammation and tissue damage through AGE-RAGE binding. AGEs cross-link collagen and other proteins and thus increase the stiffness of tissues such as the major arteries, heart, bone, and muscle.

There is clearly an abundance of in vitro data and a handful of in vivo animal findings that support various options for dietary therapy directed against “sugar sag.” However, studies in humans are limited by logistical, ethical, and inherent study design issues. Nevertheless, the role of diet in skin aging is undeniable. As our understanding of how accumulation of AGEs affects a rapidly growing number of pathologies, it is inevitable that our research methods will evolve to better address the challenges that currently seem so discouraging.

In the meantime, awareness of the critical impact of AGE formation in both diabetics and non-diabetics must be extended to all patients, regardless of their current health status. That task begins with clinicians. Dietary counseling should be incorporated into our regular interactions with patients, alongside essential discussions about UV-protection and avoidance of tobacco. After all, these are the three most important known exogenous aging factors. Their common grouping is reflective of their interconnected nature and their action in concert to disturb homeostasis.

Finally, there is ample evidence that AGEs play an important role in skin aging. There are also numerous studies investigating potential substances against excessive accumulation of AGEs in tissues. Some of these studies have already shown protective effects against diabetic complications. Modification of intake and circulating levels of AGEs may be a possible strategy to promote health in old age, especially because most Western foods are processed at high temperature and are rich in AGEs.

As controlled human studies investigating the effects of these anti-AGE strategies against skin aging are largely missing, this is a hot field for future researches.

References

1. Nguyen HP, Katta R. Sugar Sag: Glycation and the Role of Diet in Aging Skin. *Skin Therapy Lett.* 2015 Nov;20(6):1-5.
2. Semba RD, Nicklett EJ, Ferrucci L. Does accumulation of advanced glycation end products contribute to the aging phenotype? *J Gerontol A Biol Sci Med Sci.* 2010 Sep;65(9):963-75.
3. Tosato M, Zamboni V, Ferrini A, Cesari M. The aging process and potential interventions to extend life expectancy. *Clin Interv Aging.* 2007;2:401-412.
4. Zouboulis CC, Boschnakow A. Chronological ageing and photoageing of the human sebaceous gland. *Clin Exp Dermatol.* 2001 Oct;26(7):600-7.
5. Elias PM, Ghadially R. The aged epidermal permeability barrier: basis for functional abnormalities. *Clin Geriatr Med.* 2002 Feb;18(1):103-20.
6. Ye J, Garg A, Calhoun C, et al. Alterations in cytokine regulation in aged epidermis: implications for permeability barrier homeostasis and inflammation. I. IL1 gene family. *Exp Dermatol.* 2002 Jun;11(3):209-16.
7. Tsutsumi M, Denda M. Paradoxical effects of beta-estradiol on epidermal permeability barrier homeostasis. *Br J Dermatol.* 2007 Oct;157(4):776-9.
8. Fimmel S, Kurfurst R, Bonte F, et al. Responsiveness to androgens and effectiveness of antisense oligonucleotides against the androgen receptor on human epidermal keratinocytes is dependent on the age of the donor and the location of cell origin. *Horm Metab Res.* 2007 Feb;39(2):157-65.
9. Makrantonaki E, Vogel K, Fimmel S, et al. Interplay of IGF-I and 17betaestradiol at age-specific levels in human sebocytes and fibroblasts in vitro. *Exp Gerontol.* 2008 Oct;43(10):939-46.
10. Ashcroft GS, Horan MA, Ferguson MW. The effects of ageing on wound healing: immunolocalisation of growth factors and their receptors in a murine incisional model. *J Anat.* 1997 Apr;190 (Pt 3):351-65.
11. Bhushan M, Cumberbatch M, Dearman RJ, et al. Tumour necrosis factoralpha-induced migration of human Langerhans cells: the influence of ageing. *Br J Dermatol.* 2002 Jan;146(1):32-40.
12. Zouboulis CC, Makrantonaki E. Clinical aspects and molecular diagnostics of skin aging. *Clin Dermatol.* 2011 Jan-Feb;29(1):3-14.
13. Chung JH, Yano K, Lee MK, et al. Differential effects of photoaging vs intrinsic aging on the vascularization of human skin. *Arch Dermatol.* 2002 Nov;138(11):1437-42.

14. Cordain L, Eaton SB, Sebastian A, Mann N, Lindeberg S, Watkins BA, O'Keefe JH, Brand-Miller J. Origins and evolution of the western diet: health implications for the 21st century. *Am J Clin Nutr* 2005;81:341-354.
15. Rahbar S, Blumenfe O, Ranney HM. Studies of an unusual hemoglobin in patients with diabetes mellitus. *Biochem Biophys Res Commun* 1969;36:838-843.
16. Sharma C, Kaur A, Thind SS, Singh B, Raina S. Advanced glycation Endproducts (AGEs): an emerging concern for processed food industries. *J Food Sci Technol* 2015;52(12):7561-7576.
17. Ulrich P, Cerami A. Protein glycation, diabetes and aging. *Recent Prog Horm Res* 2001;56:21.
18. Uribarri J, Woodruff S, Goodman S, Cai W, Chen X, Pyzik R, Yong A, Striker GE, Vlassara H. Advanced glycation end products in foods and a practical guide to their reduction in the diet. *J Am Diet Assoc* 2010;110(6):911-16.
19. Vashishth D, Gibson GJ, Khoury JI, SchafflerMB, Kimura J, Fy hrie DP. Influence of nonenzymatic glycation on biomechanical properties of cortical bone. *Bone* 2001;28:195-201.
20. Nicholl ID, Stitt AW, Moore JE, Ritchie AJ, Archer DB, Bucala R. Increased levels of advanced glycation endproducts in the lenses and blood vessels of cigarette smokers. *Mol Med* 1998; 4:594-601.
21. O'Brien J, Morrissey PA. Nutritional and toxicological aspects of the Maillard browning reaction in foods. *Crit Rev Food Sci Nutr*. 1989 28(3):211-48.
22. Van Puyvelde K, Mets T, Njemini R, et al. Effect of advanced glycation end product intake on inflammation and aging: a systematic review. *Nutr Rev*. 2014 Oct;72(10):638-50.
23. Mook-Kanamori MJ, Selim MM, Takiddin AH, et al. Ethnic and gender differences in advanced glycation end products measured by skin autofluorescence. *Dermatoendocrinol*. 2013 Apr 1;5(2):325-30.
24. Medvedev ZA. An attempt at a rational classification of theories of ageing. *Biol Rev Camb Philos Soc* 1990; 65:375-98.
25. Dimri GP, Lee X, Basile G, Acosta M, Scott G, Roskelley C, et al. A biomarker that identifies senescent human cells in culture and in aging skin in vivo. *Proc Natl Acad Sci U S A* 1995; 92:9363-7.
26. Allsopp RC, Vaziri H, Patterson C, Goldstein S, Younglai EV, Futcher AB, et al. Telomere length predicts replicative capacity of human fibroblasts. *Proc Natl Acad Sci U S A* 1992; 89:10114-8.
27. Michikawa Y, Mazzucchelli F, Bresolin N, Scarlato G, Attardi G. Agingdependent large accumulation of point mutations in the human mtDNA control region for replication. *Science* 1999; 286:774-9.
28. Harman D. The free radical theory of aging. *Antioxid Redox Signal* 2003; 5:557-61.
29. Giacomoni PU, Rein G. Factors of skin ageing share common mechanisms. *Biogerontology* 2001; 2:219-29.
30. Paraskevi Gkogkolou P, Böhm M. Advanced glycation end products Key players in skin aging? *Dermato-Endocrinology* 2012; 4(3):259-270.
31. JohnWG, Lamb EJ. Themaillard or browning reaction in diabetes. *Eye* 1993;7:230-237.
32. Ahmed N. Advanced glycation end products-role in pathology of diabetic complications. *Diabetes Res Clin Pract* 2005; 67:3-21.
33. Maillard LC. Action des acides amines sur les sucres: formation des melanoidines par voie methodique. *C R Acad Sci (Paris)* 1912; 154:66-8.
34. Hodge JE. Dehydrated foods, chemistry of browning reactions in model systems. *J Agric Food Chem* 1953;1:928-43.
35. Thorpe SR, Baynes JW. Maillard reaction products in tissue proteins: new products and new perspectives. *Amino Acids*. 2003 Dec;25(3-4):275-81.
36. Kawabata K, Yoshikawa H, Saruwatari K, et al. The presence of N(epsilon)(carboxymethyl) lysine in the human epidermis. *Biochim Biophys Acta*. 2011 Oct;1814(10):1246-52.
37. Dyer DG, Dunn JA, Thorpe SR, et al. Accumulation of Maillard reaction products in skin collagen in diabetes and aging. *J Clin Invest*. 1993 Jun;91(6):2463-9.
38. Jeanmaire C, Danoux L, Pauly G. Glycation during human dermal intrinsic and actinic ageing: an in vivo and in vitro model study. *Br J Dermatol*. 2001 Jul;145(1):10-8.
39. Gkogkolou P, Bohm M. Advanced glycation end products: Key players in skin aging? *Derma-toendocrinol*. 2012 Jul 1;4(3):259-70.
40. Paul RG, Bailey AJ. Glycation of collagen: the basis of its central role in the late complications of ageing and diabetes. *Int J Biochem Cell Biol* 1996; 28:1297-310.
41. Fan X, Sell DR, Zhang J, Nemet I, Theves M, Lu J, et al. Anaerobic vs aerobic pathways of carbonyl and oxidant stress in human lens and skin during aging and in diabetes: A comparative analysis. *Free Radic Biol Med* 2010; 49:847-56.
42. Kueper T, Grune T, Prahl S, Lenz H, Welge V, Biernoth T, et al. Vimentin is the specific target in skin glycation. Structural prerequisites, functional consequences, and role in skin aging. *J Biol Chem* 2007; 282:23427-36.

43. Mizutani K, Ono T, Ikeda K, Kayashima K, Horiuchi S. Photo enhanced modification of human skin elastin in actinic elastosis by N(epsilon)(carboxymethyl)lysine, one of the glycoxidation products of the Maillard reaction. *J Invest Dermatol* 1997; 108:797-802.
44. Yu Y, Thorpe SR, Jenkins AJ, Shaw JN, Sochaski MA, McGee D, et al. Advanced glycation end-products and methionine sulphoxide in skin collagen of patients with type 1 diabetes. *Diabetologia* 2006; 49:2488-98.
45. Taneda S, Monnier VM. ELISA of pentosidine, an advanced glycation end product, in biological specimens. *Clin Chem* 1994; 40:1766-73.
46. Sell DR, Biemel KM, Reihl O, Lederer MO, Strauch CM, Monnier VM. Glucosepane is a major protein cross-link of the senescent human extracellular matrix. Relationship with diabetes. *J Biol Chem* 2005; 280:12310-5.
47. Ahmed MU, Brinkmann Frye E, Degenhardt TP, Thorpe SR, Baynes JW. Nepsilon-(carboxyethyl) lysine, a product of the chemical modification of proteins by methylglyoxal, increases with age in human lens proteins. *Biochem J* 1997; 324:565-70.
48. Frye EB, Degenhardt TP, Thorpe SR, Baynes JW. Role of the Maillard reaction in aging of tissue proteins. Advanced glycation end product-dependent increase in imidazolium cross-links in human lens proteins. *J Biol Chem* 1998; 273:18714-9.
49. Ahmed MU, Thorpe SR, Baynes JW. Identification of N epsilon-carboxymethyllysine as a degradation product of fructoselysine in glycated protein. *J Biol Chem* 1986; 261:4889-94.
50. Reddy S, Bichler J, Wells-Knecht KJ, Thorpe SR, Baynes JW. N epsilon(carboxymethyl)lysine is a dominant advanced glycation end product (AGE) antigen in tissue proteins. *Biochemistry* 1995; 34:10872-8.
51. Sell DR, Monnier VM. Isolation, purification and partial characterization of novel fluorophores from aging human insoluble collagen-rich tissue. *Connect Tissue Res* 1989; 19:77-92.
52. Thornalley PJ, Langborg A, Minhas HS. Formation of glyoxal, methylglyoxal and 3-deoxyglucosone in the glycation of proteins by glucose. *Biochem J* 1999; 344:109-16.
53. Fleming TH, Humpert PM, Nawroth PP, Bierhaus A. Reactive metabolites and AGE/RAGE-mediated cellular dysfunction affect the aging process: a mini-review. *Gerontology* 2011; 57:435-43.
54. Cerami C, Founds H, Nicholl I, Mitsuhashi T, Giordano D, Vanpatten S, et al. Tobacco smoke is a source of toxic reactive glycation products. *Proc Natl Acad Sci U S A* 1997; 94:13915-20.
55. Leslie RD, Beyan H, Sawtell P, Boehm BO, Spector TD, Snieder H. Level of an advanced glycated end product is genetically determined: a study of normal twins. *Diabetes* 2003; 52:2441-4.
56. Thornalley PJ. The enzymatic defence against glycation in health, disease and therapeutics: a symposium to examine the concept. *Biochem Soc Trans* 2003; 31:1341-2.
57. Xue M, Rabbani N, Thornalley PJ. Glyoxalase in ageing. *Semin Cell Dev Biol* 2011; 22:293-301.
58. Wu X, Monnier VM. Enzymatic deglycation of proteins. *Arch Biochem Biophys* 2003; 419:16-24.
59. Van Schaftingen E, Collard F, Wiame E, Veiga-da-Cunha M. Enzymatic repair of Amadori products. *Amino Acids* 2012; 42:1143-50.
60. Conner JR, Beisswenger PJ, Szwegold BS. Some clues as to the regulation, expression, function, and distribution of fructosamine-3-kinase and fructosamine-3kinase-related protein. *Ann NY Acad Sci* 2005; 1043:824-36.
61. Bierhaus A, Humpert PM, Morcos M, Wendt T, Chavakis T, Arnold B, et al. Understanding RAGE, the receptor for advanced glycation end products. *J Mol Med (Berl)* 2005; 83:876-86.
62. Loughlin DT, Artlett CM. Precursor of advanced glycation end products mediates ER-stress-induced caspase-3 activation of human dermal fibroblasts through NAD(P)H oxidase 4. *PLoS One* 2010; 5:e11093.
63. Ramasamy R, Vannucci SJ, Yan SS, Herold K, Yan SF, Schmidt AM. Advanced glycation end products and RAGE: a common thread in aging, diabetes, neurodegeneration, and inflammation. *Glycobiology* 2005; 15:16R-28R.
64. Lohwasser C, Neureiter D, Weigle B, Kirchner T, Schuppan D. The receptor for advanced glycation end products is highly expressed in the skin and upregulated by advanced glycation end products and tumor necrosis factor-alpha. *J Invest Dermatol* 2006; 126:291-9.
65. Fujimoto E, Kobayashi T, Fujimoto N, Akiyama M, Tajima S, Nagai R. AGE modified collagens I and III induce keratinocyte terminal differentiation through AGE receptor CD36: epidermal-dermal interaction in acquired perforating dermatosis. *J Invest Dermatol* 2010; 130:405-14.
66. Zhu P, Ren M, Yang C, Hu YX, Ran JM, Yan L. Involvement of RAGE, MAPK and NF-κB pathways in AGEs-induced MMP-9 activation in HaCaT keratinocytes. *Exp Dermatol* 2012; 21:123-9.
67. Hilmenyuk T, Bellinghausen I, Heydenreich B, Ilchmann A, Toda M, Grabbe S, et al. Effects of glycation of the model food allergen ovalbumin on

- antigen uptake and presentation by human dendritic cells. *Immunology* 2010; 129:437-45.
68. Chen Y, Akirav EM, Chen W, Henegariu O, Moser B, Desai D, et al. RAGE ligation affects T cell activation and controls T cell differentiation. *J Immunol* 2008;181:4272-8.
69. Tanaka N, Yonekura H, Yamagishi S, Fujimori H, Yamamoto Y, Yamamoto H. The receptor for advanced glycation end products is induced by the glycation products themselves and tumor necrosis factor-alpha through nuclear factor-kappa B, and by 17beta-estradiol through Sp-1 in human vascular endothelial cells. *J Biol Chem* 2000; 275:25781-90.
70. Vlassara H. The AGE-receptor in the pathogenesis of diabetic complications. *Diabetes Metab Res Rev* 2001; 17:436-43.
71. Lu C, He JC, Cai W, Liu H, Zhu L, Vlassara H. Advanced glycation endproduct (AGE) receptor 1 is a negative regulator of the inflammatory response to AGE in mesangial cells. *Proc Natl Acad Sci U S A* 2004; 101:11767-72.
72. Cai W, He JC, Zhu L, Chen X, Wallenstein S, Striker GE, Vlassara H. Reduced oxidant stress and extended lifespan in mice exposed to a low glycotoxin diet: association with increased AGER1 expression. *Am J Pathol* 2007;170:1893-1902.
73. Ramasamy R, Yan SF, Schmidt AM. RAGE: therapeutic target and biomarker of the inflammatory response-the evidence mounts. *J Leukoc Biol* 2009; 86:505-12.
74. Brownlee M, Vlassara H, Cerami A. Nonenzymatic glycosylation and the pathogenesis of diabetic complications. *Annu Int Med* 1984;101:527-537.
75. Raj DS, Choudhury D, Welbourne TC, Levi M AGE: a nephrologist's perspective. *Am J Kidney Dis* 2000;35:365-380.
76. Uribarri J. Advanced Glycation End Products. In: Daugirdas JT. (ed) handbook of chronic kidney disease management. Lippincott Williams and Wilkins 2012, pp 152- 158.
77. Hipkiss AR. Accumulation of altered proteins and ageing: causes and effects. *Exp Gerontol*. 2006 May;41(5):464-73.
78. Dunn JA, McCance DR, Thorpe SR, et al. Age-dependent accumulation of N epsilon-(carboxymethyl)lysine and N epsilon-(carboxymethyl) hydroxylysine in human skin collagen. *Biochemistry*. 1991 Feb 5;30(5): 1205-10.
79. Avery NC, Bailey AJ. The effects of the Maillard reaction on the physical properties and cell interactions of collagen. *Pathol Biol (Paris)*. 2006 Sep;54(7):387-95.
80. Haitoglou CS, Tsilibary EC, Brownlee M, et al. Altered cellular interactions between endothelial cells and nonenzymatically glucosylated laminin/type IV collagen. *J Biol Chem*. 1992 Jun 25;267(18):12404-7.
81. Goldin A, Beckman JA, Schmidt AM, et al. Advanced glycation end products: sparking the development of diabetic vascular injury. *Circulation*. 2006 Aug 8;114(6):597-605.
82. DeGroot J, Verzijl N, Wenting-Van Wijk MJ, et al. Age-related decrease in susceptibility of human articular cartilage to matrix metalloproteinase-mediated degradation: the role of advanced glycation end products. *Arthritis Rheum*. 2001 Nov;44(11):2562-71.
83. Nowotny K, Grune T. Degradation of oxidized and glycoxidized collagen: role of collagen cross-linking. *Arch Biochem Biophys*. 2014 Jan 15;542:56-64.
84. Yoshinaga E, Kawada A, Ono K, et al. N(varepsilon)-(carboxymethyl)lysine modification of elastin alters its biological properties: implications for the accumulation of abnormal elastic fibers in actinic elastosis. *J Invest Dermatol*. 2012 Feb;132(2):315-23.
85. Masaki H, Okano Y, Sakurai H. Generation of active oxygen species from advanced glycation end-products (AGEs) during ultraviolet light A (UVA) irradiation and a possible mechanism for cell damaging. *Biochim Biophys Acta*. 1999 Jun 28;1428(1):45-56.
86. Alikhani Z, Alikhani M, Boyd CM, et al. Advanced glycation end products enhance expression of pro-apoptotic genes and stimulate fibroblast apoptosis through cytoplasmic and mitochondrial pathways. *J Biol Chem*. 2005 Apr 1;280(13):12087-95.
87. Zhu P, Yang C, Chen LH, et al. Impairment of human keratinocyte mobility and proliferation by advanced glycation end products-modified BSA. *Arch Dermatol Res*. 2011 Jul;303(5):339-50.
88. Bulteau AL, Verbeke P, Petropoulos I, et al. Proteasome inhibition in glyoxal-treated fibroblasts and resistance of glycated glucose-6-phosphate dehydrogenase to 20 S proteasome degradation in vitro. *J Biol Chem*. 2001 Dec 7;276(49):45662-8.
89. Roberts MJ, Wondrak GT, Laurean DC, et al. DNA damage by carbonyl stress in human skin cells. *Mutat Res*. 2003 Jan 28;522(1-2):45-56.
90. Verzijl N, DeGroot J, Oldehinkel E, Bank RA, Thorpe SR, Baynes JW, et al. Age-related accumulation of Maillard reaction products in human articular cartilage collagen. *Biochem J* 2000; 350:381-7.
91. Haus JM, Carrithers JA, Trappe SW, Trappe TA. Collagen, cross-linking, and advanced glycation end products in aging human skeletal muscle. *J Appl Physiol* 2007; 103:2068-76.

92. Sell DR, Carlson EC, Monnier VM. Differential effects of type 2 (non-insulindependent) diabetes mellitus on pentosidine formation in skin and glomerular basement membrane. *Diabetologia* 1993; 36:936-41
93. Vlassara H, Cai W, Crandall J, Goldberg T, Oberstein R, Dardaine V, et al. Inflammatory mediators are induced by dietary glycotoxins, a major risk factor for diabetic angiopathy. *Proc Natl Acad Sci U S A* 2002; 99:15596-601.
94. Glenn JV, Beattie JR, Barrett L, Frizzell N, Thorpe SR, Boulton ME, et al. Confocal Raman microscopy can quantify advanced glycation end product (AGE) modifications in Bruch's membrane leading to accurate, nondestructive prediction of ocular aging. 2007; 21:3542-52.
95. Stitt AW. Advanced glycation: an important pathological event in diabetic and age related ocular disease. *Br J Ophthalmol* 2001; 85:746-53.
96. Sell DR, Lane MA, Johnson WA, Masoro EJ, Mock OB, Reiser KM, et al. Longevity and the genetic determination of collagen glycoxidation kinetics in mammalian senescence. *Proc Natl Acad Sci U S A* 1996;93:485-90.
97. Schleicher ED, Wagner E, Nerlich AG. Increased accumulation of the glycoxidation product N(epsilon)-(carboxymethyl)lysine in human tissues in diabetes and aging. *J Clin Invest* 1997; 99:457-68.
98. Corstjens H, Dicanio D, Muizzuddin N, Neven A, Sparacio R, Declercq L, et al. Glycation associated skin autofluorescence and skin elasticity are related to chronological age and body mass index of healthy subjects. *Exp Gerontol* 2008; 43:663-7.
99. Pigeon H. Reaction of glycation and human skin: the effects on the skin and its components, reconstructed skin as a model. *Pathol Biol (Paris)* 2010; 58:226-31.
100. Lutgers HL, Graaff R, Links TP, Ubink-Veltmaat LJ, Bilo HJ, Gans RO, et al. Skin autofluorescence as a noninvasive marker of vascular damage in patients with type 2 diabetes. *Diabetes Care* 2006; 29:2654-9.
101. Goldberg T, Cai W, Peppia M, Dardaine V, Baliga BS, Uribarri J, et al. Advanced glycoxidation end products in commonly consumed foods. *J Am Diet Assoc* 2004; 104:1287-91.
102. Uribarri J, Cai W, Peppia M, Goodman S, Ferrucci L, Striker G, et al. Circulating glycotoxins and dietary advanced glycation endproducts: two links to inflammatory response, oxidative stress, and aging. *J Gerontol A Biol Sci Med Sci* 2007; 62:427-33.
103. DeGroot J. The AGE of the matrix: chemistry, consequence and cure. *Curr Opin Pharmacol* 2004; 4:301-5.)
104. Reihnsner R, Melling M, Pfeiler W, Menzel EJ. Alterations of biochemical and two-dimensional biomechanical properties of human skin in diabetes mellitus as compared to effects of in vitro non-enzymatic glycation. *Clin Biomech (Bristol, Avon)* 2000; 15:379-86.
105. Yoon HS, Baik SH, Oh CH. Quantitative measurement of desquamation and skin elasticity in diabetic patients. *Skin Res Technol* 2002; 8:250-4.
106. Giardino I, Edelstein D, Brownlee M. Non-enzymatic glycosylation in vitro and in bovine endothelial cells alters basic fibroblast growth factor activity. A model for intracellular glycosylation in diabetes. *J Clin Invest* 1994; 94:110-7.
107. Uchiki T, Weikel KA, Jiao W, Shang F, Caceres A, Pawlak D, et al. Glycationaltered proteolysis as a pathobiologic mechanism that links dietary glycaemic index, aging, and age-related disease (in nondiabetics). *Aging Cell* 2012; 11:1-13.
108. Ukeda H, Hasegawa Y, Ishi T, Sawamura M. Inactivation of Cu,Zn-superoxide dismutase by intermediates of Maillard reaction and glycolytic pathway and some sugars. *Biosci Biotechnol Biochem* 1997; 61:2039-42.
109. Berge U, Behrens J, Rattan SI. Sugar-induced premature aging and altered differentiation in human epidermal keratinocytes. *Ann N Y Acad Sci* 2007; 1100:524-9.
110. Ravelojaona V, Robert AM, Robert L. Expression of senescence-associated betagalactosidase (SA-beta-Gal) by human skin fibroblasts, effect of advanced glycation end-products and fucose or rhamnose-rich polysaccharides. *Arch Gerontol Geriatr* 2009; 48:151-4.
- Sejersen H, Rattan SI. Dicarbonyl-induced accelerated aging in vitro in human skin fibroblasts. *Biogerontology* 2009; 10:203-11.
111. Portero-Otín M, Pamplona R, Bellmunt MJ, Ruiz MC, Prat J, Salvayre R, et al. Advanced glycation end product precursors impair epidermal growth factor receptor signaling. *Diabetes* 2002; 51:1535-42.
112. Wondrak GT, Roberts MJ, Jacobson MK, Jacobson EL. Photosensitized growth inhibition of cultured human skin cells: mechanism and suppression of oxidative stress from solar irradiation of glycated proteins. *J Invest Dermatol* 2002; 119:489-98.
113. Yim MB, Yim HS, Lee C, Kang SO, Chock PB. Protein glycation: creation of catalytic sites for free radical generation. *Ann N Y Acad Sci* 2001; 928:48-53.
114. Lee C, Yim MB, Chock PB, Yim HS, Kang SO. Oxidation-reduction properties of methylglyoxal-modified protein in relation to free radical generation. *J Biol Chem* 1998; 273:25272-8.
115. Qian M, Liu M, Eaton JW. Transition metals bind to glycated proteins forming redox active "glycochelates": implications for the pathogenesis of certain diabetic complications. *Biochem Biophys Res Commun* 1998; 250:385-9.

116. Meerwaldt R, Links T, Graaff R, Thorpe SR, Baynes JW, Hartog J, et al. Simple noninvasive measurement of skin autofluorescence. *Ann N Y Acad Sci* 2005; 1043:290-8.
117. Mulder DJ, Water TV, Lutgers HL, Graaff R, Gans RO, Zijlstra F, et al. Skin autofluorescence, a novel marker for glycemic and oxidative stress-derived advanced glycation endproducts: an overview of current clinical studies, evidence, and limitations. *Diabetes Technol Ther* 2006; 8:523-35.
118. Tseng JY, Ghazaryan AA, Lo W, Chen YF, Hovhannisyan V, Chen SJ, et al. Multiphoton spectral microscopy for imaging and quantification of tissue glycation. *Biomed Opt Express* 2010; 2:218-30.
119. Bos DC, de Ranitz-Greven WL, de Valk HW. Advanced glycation end products, measured as skin autofluorescence and diabetes complications: a systematic review. *Diabetes Technol Ther* 2011; 13:773-9.
120. Smit AJ, Gerrits EG. Skin autofluorescence as a measure of advanced glycation endproduct deposition: a novel risk marker in chronic kidney disease. *Curr Opin Nephrol Hypertens* 2010; 19:527- 33.
121. Genuth S, Sun W, Cleary P, Sell DR, Dahms W, Malone J, et al.; DCCT Skin Collagen Ancillary Study Group. Glycation and carboxymethyllysine levels in skin collagen predict the risk of future 10-year progression of diabetic retinopathy and nephropathy in the diabetes control and complications trial and epidemiology of diabetes interventions and complications participants with type 1 diabetes. *Diabetes* 2005; 54:3103-11.
122. Beisswenger PJ, Howell S, Mackenzie T, Corstjens H, Muizzuddin N, Matsui MS. Two fluorescent wavelengths, 440(ex)/520(em) nm and 370(ex)/440(em) nm, reflect advanced glycation and oxidation end products in human skin without diabetes. *Diabetes Technol Ther* 2012; 14:285-92.
123. Story M, Hayes M, Kalina B. Availability of foods in high schools: is there cause for concern? *J Am Diet Assoc* 1996;96:123-126.
124. Vlassara H, Uribarri J. Glycooxidation and diabetic complications: modern lessons and a warning? *Rev Endocr Metab Disord* 2004;5:181-188.
125. Ledl F, Schleicher E. New aspects of the maillard reaction in foods and in the human body. *Angew Chem Int Ed* 1990;29:565-594.
126. Nursten H. Introduction. In: *The Maillard Reaction Chemistry, Biochemistry and Implications*. The Royal Society of Chemistry 2005:pp 1-4.
127. Nursten H. Recent advances. In: *The Maillard Reaction Chemistry, Biochemistry and implications*. The Royal Society of Chemistry 2005:pp 31-51.
128. Poulsen MW, Hedegaard RV, Anderson JM, Courten B, Bugel S, Nielsen J, Skibsted LH, Dragsted L. Advanced glycation endproducts in food and their effects on health. *Food Chem Toxicol* 2013. doi:10.1016/j.fct.2013.06.052.
129. Finot PA, Magnenat E. Metabolic transit of early and advanced maillard products. *Prog Food Nutr Sci* 1981;5:193-207.
130. Koschinsky T, He CJ, Mitsuhashi T, Bucala R, Liu C, Buenting C, Heitmann K, Vlassara H. Orally absorbed reactive glycation products (glycotoxins): an environmental risk factor in diabetic nephropathy. *Proc Natl Acad Sci U S A* 1997;94(12):6474-6479.
131. Delgado-Andrade C, Seiquer I, Garcia MM, Galdo G, Navarro MP. Increased maillard reaction products intake reduces phosphorus digestibility in male adolescents. *Nutr* 2011;27:86-91.
132. Garcia MM, Seiquer I, Delgado-Andrade C, Galdo G, Navarro MP. Intake of Maillard reaction products reduces iron bioavailability in male adolescents. *Mol Nutr Food Res* 2009;53:1551-1560.
133. Seiquer I, Diaz-Alguacil J, Delgado-Andrade C, Lopez-Frias M, Munoz HA, Galdo G, Navarro MP. Diets rich in maillard reaction products affect protein digestibility in adolescent males aged 11-14 y. *Am J Clin Nutr* 2006;83:1082-1088.
134. Bergmann R, Helling R, Heichert C, Scheunemann M, Mading P, Wittirsch H, Johannsen B, Henle T. Radio fluorination and positron emission tomography (PET) as a new approach to study the in vivo distribution and elimination of the advanced glycation endproducts N epsilon-carboxymethyllysine (CML) and N epsilon-carboxyethyllysine (CEL). *Nahrung* 2001;45:182-188.
135. He C, Sabol J, Mitsuhashi T, Vlassara H. Dietary glycotoxins: inhibition of reactive products by aminoguanidine facilitates renal clearance and reduces tissue sequestration. *Diabetes* 1999;48:1308-1315.
136. Peppas M, Brem H, Ehrlich P, Zhang JG, Cai W, Li Z, et al. Adverse effects of dietary glycotoxins on wound healing in genetically diabetic mice. *Diabetes* 2003; 52:2805-13.
137. Hofmann SM, Dong HJ, Li Z, Cai W, Altomonte J, Thung SN, Zeng F, Fisher EA, Vlassara H. Improved insulin sensitivity is associated with restricted intake of dietary glycooxidation products in the db/db mouse. *Diabetes* 2002;51:2082-2089.
138. Bierhaus A, Ziegler R, Nawroth PP. Molecular mechanisms of diabetic angiopathy clues for innovative therapeutic interventions. *Horm Res* 1998;50(Suppl 1):1-5.
139. Sano H, Higashi T, Matsumoto K et al. Insulin enhances macrophage scavenger receptor mediated endocytic uptake of advanced glycated end products. *J Biol Chem* 1998;273:8630-8637.

140. Uribarri J, Cai WJ, Sandu O, Peppia M, Goldberg T, Vlassara H. Diet- derived advanced glycation end products are major contributors to the body's AGE pool and induce inflammation in healthy subjects. *Ann N Y Acad Sci* 2005;1043:461–466.
141. Cai W, Cao QD, Zhu L et al. Oxidative stress-inducing carbonyl compounds from common foods: novel mediators of cellular dysfunction. *Mol Med* 2002;8:337– 346.
142. Uribarri J, Peppia M, Cai W, Goldberg Tet al. Restriction of dietary glycotoxins markedly reduces AGE toxins in renal failure patients. *J Am Soc Nephrol* 2003;14:728–731.
143. Zheng F, He C, Cai W, Hattori M, Steffes M, Vlassara H. Prevention of diabetic nephropathy in mice by a diet low in glycooxidation products. *Diabetes Metab Res Rev* 2002;18:224–237.
144. Tan AL, Sourris KC, Harcourt BE, Thallas-Bonke V, Penfold S, Andrikopoulos S, Thomas MC, O'Brien RC, Bierhaus A, Cooper ME, Forbes JM, Coughlan MT. Disparate effects on renal and oxidative parameters following RAGE deletion, AGE accumulation inhibition, or dietary AGE control in experimental diabetic nephropathy. *Am J Physiol Ren Physiol* 2010;298:763–770.
145. Feng JX, Hou FF, Liang M, Wang GB, Zhang X, Li HY, Xie D, Tian JW, Liu ZQ. Restricted intake of dietary advanced glycation end products retards renal progression in the remnant kidney model. *Kidney Int* 2007;71:901–911.
146. Sebekova K, Faist V, Hofmann T, Schinzel R, Heidland A. Effects of a diet rich in advanced glycation end products in the rat remnant kidney model. *Am J Kidney Dis* 2003;41:48–51.
147. Harcourt BE, Sourris KC, Coughlan MT, Walker KZ, Dougherty SL, Andrikopoulos S, Morley AL, Thallas-Bonke V et al. Targeted reduction of advanced glycation improves renal function in obesity. *Kidney Int* 2011;80:190–198.
148. Lin RY, Choudhury RP, Cai W, Lu M, Fallon JT, Fisher EA, Vlassara H. Dietary glycotoxins promote diabetic atherosclerosis in apolipoprotein E-deficient mice. *Atherosclerosis* 2003;168:213–220.
149. Cai W, He JC, Zhu L, Chen X, Zheng F, Striker GE, et al. Oral glycotoxins determine the effects of calorie restriction on oxidant stress, age-related diseases, and lifespan. *Am J Pathol* 2008; 173:327–36.
150. Ames JM. Determination of Ne-(carboxymethyl) lysine in foods and related systems. *Ann N Y Acad Sci* 2008;1126:20–24.
151. Hartkopf J, Pahlke C, Lüdemann G, Erbersdobler HF. Determination of Nε-carboxymethyllysine by a reserved-phase high-performance liquid chromatography method. *J Chromatogr* 1994;672:242–246.
152. Delgado-Andrade C, Rufián-Henares JA, Morales FJ. Study on fluorescence of maillard reaction compounds in breakfast cereals. *Mol Nutr Food Res* 2006;50:799– 804.
153. Yaacoub R, Saliba R, Nsouli B, Khalaf G, Birlouez-Aragon I (2008) Formation of lipid oxidation and isomerization products during processing of nuts and sesame seeds. *Journal Agric Food Chem* 2008;6:7082–7090.
154. Drusch S, Faist V, Erbersdobler HF (1999) Determination of Nε-pi-carboxymethyllysine in milk products by a modified reversed-phase HPLC method. *Food Chem* 1999;65:547–553.
155. Chao PC, Hsu CC, Yin MC. Analysis of glycative products in sauces and saucetreated foods. *Food Chem* 2009;113:262–266.
156. Birlouez-Aragon I, Pischetsrieder M, Leclère J et al. Assessment of protein glycation markers in infant formulas. *Food Chem* 2004;87:253–259.
157. Sebekova K, Saavedra G, Zumpe C, Somoza V, Klenovicsova K, Birlouez-Aragon I. Plasma concentration and urinary excretion of Ne-(carboxymethyl)lysine in breast milk- and formula-fed infants. *Ann N Y Acad Sci* 2008;1126:177–180.
158. Dhuique-Mayer C, Tbatou M, Carail M, Caris-Veyrat C, Dornier M, Amiot MJ (2007) Thermal degradation of antioxidant micronutrients in citrus juice: kinetics and newly formed compounds. *J Agric Food Chem* 2007;55:4209–4216.
159. Klopotek Y, Otto K, Bohm V. Processing strawberries to different products alters contents of vitamin C, total phenolics, total anthocyanins, and antioxidant capacity. *J Agric Food Chem* 2005;53:5640–5646.
160. Vikram VB, Ramesh MN, Prapulla SG. Thermal degradation kinetics of nutrients in orange juice heated by electromagnetic and conventional methods. *J Food Eng* 2005;69:31–40.
161. Birlouez-Aragon I, Saavedra G, Tessier FJ, Galinier A, Ait-Ameur L, Lacoste F, Niamba CN, Alt N, Somoza V, Lecerf JM. A diet based

- on high-heat-treated foods promotes risk factors for diabetes mellitus and cardiovascular diseases. *Am J Clin Nutr* 2010;91:1220–1226.
162. Depeint F, Shangari N, Furrer R, Bruce WR, O'Brien PJ. Marginal thiamine deficiency increases oxidative markers in the plasma and selected tissues in F344 rats. *Nutr Res* 2007;27:698–704.
163. Shangari N, Depeint F, Furrer R, Bruce WR, O'Brien PJ. The effects of partial thiamin deficiency and oxidative stress (i.e., glyoxal and methylglyoxal) on the levels of alpha-oxoaldehyde plasma protein adducts in Fischer 344 rats. *FEBS Lett* 2005;579:5596–5602.
164. Friedman M. Dietary impact of food processing. *Annu Rev Nutr* 1992;12:119–137.
165. Perez-Locas C, Yaylayan VA. The Maillard reaction and food quality deterioration. In: Skibsted LH, Risbo J, Andersen ML (eds) *Chemical deterioration and physical instability of food and beverages*. Woodhead Publishing, Cambridge 2010, pp 70–94.
166. Gokmen V, Senyuva HZ. Effects of some cations on the formation of acrylamide and furfurals in glucose-asparagine model system. *Eur Food Res Technol* 2012;225:815–820.
167. Tareke E, Rydberg P, Karlsson P, Eriksson S, Tornqvist M. Analysis of acrylamide, a carcinogen formed in heated foodstuffs. *J Agric Food Chem* 2002;50:4998–5006.
168. Skog KI, Johansson MA, Jagerstad MI. Carcinogenic heterocyclic amines in model systems and cooked foods: a review on formation, occurrence and intake. *Food Chem Toxicol* 1998;36:879–896.
169. Janzowski C, Glaab V, Samimi E, Schlatter J, Eisenbrand G. 5-Hydroxymethylfurfural: assessment of mutagenicity, DNAdamaging potential and reactivity towards cellular glutathione. *Food Chem Toxicol* 2000;38:801–809.
170. Pouillart P, Mauprivez H, Ait-Ameur L, Cayzele A, Lecerf JM, Tessier FJ, Birlouez-Aragon I. Strategy for the study of the health impact of dietary maillard products in clinical studies – the example of the ICARE clinical study on healthy adults. *Ann N Y Acad Sci* 2008;1126:173–176.
171. Farris PK. Innovative cosmeceuticals: sirtuin activators and anti-glycation compounds. *Semin Cutan Med Surg* 2011;30:163-6.
172. Elost A, Ghous T, Ahmed N. Natural products as anti-glycation agents: possible therapeutic potential for diabetic complications. *Curr Diabetes Rev* 2012; 8:92-108.
173. Edelstein D, Brownlee M. Mechanistic studies of advanced glycosylation end product inhibition by aminoguanidine. *Diabetes* 1992;41:26-9.
174. Reddy VP, Beyaz A. Inhibitors of the Maillard reaction and AGE breakers as therapeutics for multiple diseases. *Drug Discov Today* 2006;11:646-54.
175. Pigeon H, Técher MP, Asselineau D. Reconstructed skin modified by glycation of the dermal equivalent as a model for skin aging and its potential use to evaluate anti-glycation molecules. *Exp Gerontol* 2008; 43:584-8.
176. Sell DR, Nelson JF, Monnier VM. Effect of chronic aminoguanidine treatment on age-related glycation, glycooxidation, and collagen cross-linking in the Fischer 344 rat. *J Gerontol A Biol Sci Med Sci* 2001; 56:B405-11.
177. Degenhardt TP, Alderson NL, Arrington DD, Beattie RJ, Basgen JM, Steffes MW, et al. Pyridoxamine inhibits early renal disease and dyslipidemia in the streptozotocin-diabetic rat. *Kidney Int* 2002; 61:939-50.
178. Caballero F, Gerez E, Batlle A, Vazquez E. Preventive aspirin treatment of streptozotocin induced diabetes: blockage of oxidative status and reversion of heme enzymes inhibition. *Chem Biol Interact* 2000;126:215–225.
179. Malik NS, Meek KM. The inhibition of sugar-induced structural alterations in collagen by aspirin and other compounds. *Biochem Biophys Res Commun* 1994;99:683–686.
180. Khalifah RG, Baynes JW, Hudson BG. Amadorins: novel postamadori inhibitors of advanced glycation reactions. *Biochem Biophys Res Commun* 1999;257:251–258.
181. Metz TO, Alderson NL, Thorpe SR, Baynes JW. Pyridoxamine, an inhibitor of advanced glycation and lipoxidation reactions: a novel therapy for treatment of diabetic complications. *Arch Biochem Biophys* 2003;419:41–49.

182. Voziyan PA, Metz TO, Baynes JW, Hudson BG. A post-Amadori inhibitor pyridoxamine also inhibits chemical modification of proteins by scavenging carbonyl intermediates of carbohydrate and lipid degradation. *J Biol Chem* 2002;277:3397–3403.
183. Booth AA, Khalifah RG, Todd P, Hudson BG (1997) In vitro kinetic studies of formation of antigenic advanced glycation end products (AGEs) Novel inhibition of post-Amadori glycation pathways. *J Biol Chem* 1997;272:5430–5437.
184. Voziyan PA, Hudson BG. Pyridoxamine: the many virtues of a Maillard reaction inhibitor. *Ann N Y Acad Sci* 2005; 1043:807-16.
185. Vasan S, Foiles P, Founds H. Therapeutic potential of breakers of advanced glycation end product-protein crosslinks. *Arch Biochem Biophys* 2003; 419:89-96.
186. Candido R, Forbes JM, Thomas MC, Thallas V, Dean RG, Burns WC, et al. A breaker of advanced glycation end products attenuates diabetes-induced myocardial structural changes. *Circ Res* 2003; 92:785-92.
187. Bakris GL, Bank AJ, Kass DA, Neutel JM, Preston RA, Oparil S. Advanced glycation end-product cross-linkbreakers. A novel approach to cardiovascular pathologies related to the aging process. *Am J Hypertens* 2004; 17:23S-30S.
188. Yang S, Litchfield JE, Baynes JW. AGE-breakers cleave model compounds, but do not break Maillard crosslinks in skin and tail collagen from diabetic rats. *Arch Biochem Biophys* 2003; 412:42-6.
189. Monnier VM, Sell DR. Prevention and repair of protein damage by the Maillard reaction in vivo. *Rejuvenation Res* 2006; 9:264-73114-117 In the rat ALT-711 showed some promising results on skin hydration.
190. Xue M, Rabbani N, Momiji H, Imbasi P, Anwar MM, Kitteringham N, et al. Transcriptional control of glyoxalase 1 by Nrf2 provides a stress-responsive defence against dicarbonyl glycation. *Biochem J* 2012; 443:213-22.
191. Monnier VM, Wu X. Enzymatic deglycation with amadoriase enzymes from *Aspergillus* sp. as a potential strategy against the complications of diabetes and aging. *Biochem Soc Trans* 2003; 31:1349-53.
192. Shinohara M, Thornalley PJ, Giardino I, Beisswenger P, Thorpe SR, Onorato J, et al. Overexpression of glyoxalase-I in bovine endothelial cells inhibits intracellular advanced glycation endproduct formation and prevents hyperglycemia-induced increases in macromolecular endocytosis. *J Clin Invest* 1998; 101:1142-7.
193. Kim KM, Kim YS, Jung DH, Lee J, Kim JS. Increased glyoxalase I levels inhibit accumulation of oxidative stress and an advanced glycation end product in mouse mesangial cells cultured in high glucose. *Exp Cell Res* 2012; 318:152-9.
194. Brouwers O, Niessen PM, Ferreira I, Miyata T, Scheffer PG, Teerlink T, et al. Overexpression of glyoxalase-I reduces hyperglycemia-induced levels of advanced glycation end products and oxidative stress in diabetic rats. *J Biol Chem* 2011; 286:1374-80.
195. Price DL, Rhett PM, Thorpe SR, Baynes JW. Chelating activity of advanced glycation end-product inhibitors. *J Biol Chem* 2001; 276:48967-72.
196. Rutter K, Sell DR, Fraser N, Obrenovich M, Zito M, Starke-Reed P, et al. Green tea extract suppresses the age-related increase in collagen crosslinking and fluorescent products in C57BL/6 mice. *Int J Vitam Nutr Res* 2003; 73:453-60.
197. Tarwadi KV, Agte VV. Effect of micronutrients on methylglyoxal-mediated in vitro glycation of albumin. *Biol Trace Elem Res*. 2011 Nov;143(2):717-25.
198. Thirunavukkarasu V, Nandhini AT, Anuradha CV. Fructose diet-induced skin collagen abnormalities are prevented by lipoic acid. *Exp Diabetes Res*. 2004 OctDec;5(4):237-44.
199. Odetti P, Pesce C, Traverso N, Menini S, Maineri EP, Cosso L, et al. Comparative trial of N-acetylcysteine, taurine, and oxerutin on skin and kidney damage in long-term experimental diabetes. *Diabetes* 2003; 52:499-505.
200. Rasheed Z, Anbazhagan AN, Akhtar N, Ramamurthy S, Voss FR, Haqqi TM. Green tea polyphenol epigallocatechin-3-gallate inhibits advanced glycation end product-induced expression of tumor necrosis factor-alpha and matrix metalloproteinase-13 in human chondrocytes. *Arthritis Res Ther* 2009; 11:R71.

201. Vinson JA, Howard HB. Inhibition of protein glycation and advanced glycation end products by ascorbic acid and other vitamins and nutrients. *J Nutr Biochem* 1996; 7:659-63.
202. Dearlove RP, Greenspan P, Hartle DK, Swanson RB, Hargrove JL. Inhibition of protein glycation by extracts of culinary herbs and spices. *J Med Food* 2008; 11:275-81.
203. Jin S, Cho KH. Water extracts of cinnamon and clove exhibits potent inhibition of protein glycation and anti-atherosclerotic activity in vitro and in vivo hypolipidemic activity in zebrafish. *Food Chem Toxicol* 2011; 49:1521-9.
204. Draelos ZD, Yatskayer M, Raab S, Oresajo C. An evaluation of the effect of a topical product containing C-xyloside and blueberry extract on the appearance of type II diabetic skin. *J Cosmet Dermatol* 2009; 8:147-51.
205. Wu CH, Yen GC. Inhibitory effect of naturally occurring flavonoids on the formation of advanced glycation endproducts. *J Agric Food Chem* 2005; 53:3167-73.
206. Fukami K, Yamagishi S, Sakai K, et al. Potential inhibitory effects of L-carnitine supplementation on tissue advanced glycation end products in patients with hemodialysis. *Rejuvenation Res.* 2013 Dec;16(6):460-6.
207. Hagen TM, Liu J, Lykkesfeldt J, et al. Feeding acetyl-L-carnitine and lipoic acid to old rats significantly improves metabolic function while decreasing oxidative stress. *Proc Natl Acad Sci USA.* 2002 Feb 19;99(4):1870-5.
208. Liang CP, Wang M, Simon JE, Ho CT. Antioxidant activity of plant extracts on the inhibition of citral off-odor formation. *Mol Nutr Food Res* 2004;48:308-317.
209. Peng X, Ma J, Cheng KW, Jiang Y, Chen F, Wang M. The effects of grape seed extract fortification on the antioxidant activity and quality attributes of bread. *Food Chem* 2010;119:49-53.
210. Peng X, Cheng KW, Ma J, Chen B, Ho CT, Lo C et al. Cinnamon bark proanthocyanidins as reactive carbonyl scavengers to prevent the formation of advanced glycation endproducts. *J Agric Food Chem* 2008;56:1907-1911.
211. Lo CY, Li S, Tan D, Pan MH, Sang S, Ho CT. Trapping reactions of reactive carbonyl species with tea polyphenols in simulated physiological conditions. *Mol Nutr Food Res* 2006;50(12):1118-1128.
212. Lee CK, Klopp RG, Weindruch R, Prolla TA. Gene expression profile of aging and its retardation by caloric restriction. *Science* 1999; 285:1390-3.
213. Cefalu WT, Bell-Farrow AD, Wang ZQ, Sonntag WE, Fu MX, Baynes JW, et al. Caloric restriction decreases age-dependent accumulation of the glycoxidation products, N-epsilon-(carboxymethyl)lysine and pentosidine, in rat skin collagen. *J Gerontol A Biol Sci Med Sci* 1995; 50:B337-41.
214. Reiser KM. Influence of age and long-term dietary restriction on enzymatically mediated crosslinks and nonenzymatic glycation of collagen in mice. *J Gerontol* 1994; 49:B71-9.
215. Sell DR, Kleinman NR, Monnier VM. Longitudinal determination of skin collagen glycation and glycoxidation rates predicts early death in C57BL/6NNIA mice. *FASEB J* 2000; 14:145-56.
216. Sebeková K, Somoza V. Dietary advanced glycation endproducts (AGEs) and their health effects-PRO. *Mol Nutr Food Res* 2007;51:1079-84.
217. Yamagishi S, Ueda S, Okuda S. Food-derived advanced glycation end products (AGEs): a novel therapeutic target for various disorders. *Curr Pharm Des* 2007; 13:2832-6.
218. Vlassara H, Striker GE. AGE restriction in diabetes mellitus: a paradigm shift. *Nat Rev Endocrinol.* 2011 Sep;7(9):526-39.
219. Lyons TJ, Bailie KE, Dyer DG, Dunn JA, Baynes JW. Decrease in skin collagen glycation with improved glycemic control in patients with insulin-dependent diabetes mellitus. *J Clin Invest* 1991; 87:1910-5.
220. Monnier VM, Bautista O, Kenny D, Sell DR, Fogarty J, Dahms W, et al.; DCCT Skin Collagen Ancillary Study Group. Skin collagen glycation, glycoxidation, and crosslinking are lower in subjects with long-term intensive versus conventional therapy of type 1 diabetes: relevance of glycated collagen products versus HbA1c as markers of diabetic complications. DCCT Skin Collagen Ancillary Study Group. *Diabetes Control and Complications Trial.* *Diabetes* 1999; 48:870-80.
221. Hudson BI, Bucciarelli LG, Wendt T, Sakaguchi T, Lalla E, Qu W, et al. Blockade of receptor

- or for advanced glycation endproducts: a new target for therapeutic intervention in diabetic complications and inflammatory disorders. *Arch Biochem Biophys* 2003; 419:80-8.
222. Yan SF, Ramasamy R, Schmidt AM. Soluble RAGE: therapy and biomarker in unraveling the RAGE axis in chronic disease and aging. *Biochem Pharmacol* 2010; 79:1379-86.
223. Goova MT, Li J, Kislinger T, Qu W, Lu Y, Bucciarelli LG, et al. Blockade of receptor for advanced glycation end-products restores effective wound healing in diabetic mice. *Am J Pathol* 2001; 159:513-25.
224. Babizhayev MA, Deyev AI, Savel'yeva EL, Lankin VZ, Yegorov YE. Skin beautification with oral non-hydrolyzed versions of carnosine and carbinine: Effective therapeutic management and cosmetic skincare solutions against oxidative glycation and free-radical production as a causal mechanism of diabetic complications and skin aging. *J Dermatolog Treat* 2012 ;23(5):345-84.
225. Babizhayev MA, Nikolayev GM, Nikolayeva JG, Yegorov YE. Biologic activities of molecular chaperones and pharmacologic chaperone imidazole-containing dipeptide-based compounds: natural skin care help and the ultimate challenge: implication for adaptive responses in the skin. *Am J Ther* 2012; 19:e69-89.
226. Babizhayev MA, Yegorov YE. Therapeutic uses of drug-carrier systems for imidazole-containing dipeptide compounds that act as pharmacological chaperones and have significant impact on the treatment of chronic diseases associated with increased oxidative stress and the formation of advanced glycation end products. *Crit Rev Ther Drug Carrier Syst* 2010; 27:85-154.
227. Urbach E, Lentz JW. Carbohydrate metabolism and the skin. *Arch Derm Syphilol*. 1945 Nov-Dec;52:301-16.
228. Danby FW. Nutrition and aging skin: sugar and glycation. *Clin Dermatol*. 2010 Jul-Aug;28(4):409-11.
229. Draelos ZD. Aging skin: the role of diet: facts and controversies. *Clin Dermatol*. 2013 Nov-Dec;31(6):701-6.
230. Wu LY, Juan CC, Ho LT, Hsu YP, Hwang LS. Effect of green tea supplement on insulin sensitivity in Sprague-Dawley rats. *J Agric Food Chem* 2004;52:643-648. 231. Babu PVA, Sabitha KE, Shyamaladevi CS. Effect of green tea extract on advanced glycation and cross-linking of tail tendon collagen in streptozotocin induced diabetic rats. *Food Chem Toxicol* 2008;46:280-285.
232. Mizutani K, Ikeda K, Yamori Y. Resveratrol inhibits AGEs-induced proliferation and collagen synthesis activity in vascular smooth muscle cells from stroke-prone spontaneously hypertensive rats. *Biochem Biophys Res Commun*. 2000 Jul 21;274(1):61-7.
233. Liu Y, He XQ, Huang X, et al. Resveratrol protects mouse oocytes from methylglyoxal-induced oxidative damage. *PLoS One*. 2013 8(10):e77960. 234. Yokozawa T, Nakagawa T. Inhibitory effects of Luobuma tea and its components against glucose-mediated protein damage. *Food Chem Toxicol* 2004;42:975-981.
235. Ardestani A, Yazdanparast R. *Cyperus rotundus* suppresses AGE formation and protein oxidation in a model of fructose-mediated protein glycooxidation. *Int J Biol Macromol* 2007;41:572-578.
236. Gugliucci A, Markowicz Bastos DH, Schulze J, Ferreira Souza MF. Caffeic and chlorogenic acids in *ilex paraguariensis* extracts are the main inhibitors of AGE generation by methylglyoxal in model proteins. *Fitoterapia* 2009;80:339-344. 237. Hsieh CL, Lin YC, Ko WS, Peng CH, Huang CN, Peng RY. Inhibitory effect of some selected nutraceutical herbs on LDL glycation induced by glucose and glyoxal. *J Ethn* 2005;102:357-363.
238. Ho SC, Wu SP, Lin SM, Tang YL. Comparison of anti-glycation capacities of several herbal infusions with that of green tea. *Food Chem* 2010;122:768-774.
239. Nakagawa T, Yokozawa T, Terasawa K, Shu S, Juneja LR. Protective activity of green tea against free radical- and glucose-mediated protein damage. *J Agric Food Chem* 2002;50:2418-2422.