

Special Report

Open Access

Medical-Scientific Rationale for a Randomized, Placebo-Controlled, Phase 2 Study of Trabedersen/OT-101 in COVID-19 Patients with Hypoxemic Respiratory Failure

Fatih M. Uckun¹ and Vuong Trieu¹

¹ Immuno-Oncology Program, Oncotelic Inc., Agoura Hills, CA 91301, USA.

***Corresponding Author: Fatih M. Uckun, MD, PhD,** Worldwide Clinical Trials, 480 E. Swedesford Road, Wayne, PA 19087. E-mail: fatih.uckun@worldwide.com

Citation: Medical-Scientific Rationale for a Randomized, Placebo-Controlled, Phase 2 Study of Trabedersen/OT-101 in COVID-19 Patients with Hypoxemic Respiratory Failure. Ann Pulm Crit Care Med..2020; 3(1); 01-09.

Submitted: 20 April 2020; Approved: 22 April 2020; Published: 24 April 2020

ABSTRACT

TGF-β was identified as the exclusive master regulator of the epithelial sodium channel (ENaC). ENAC internalization by alveolar epithelial cells and its upregulation in ARDS causes an ENAC trafficking defect with marked reduction in the cell-surface abundance of ENaC on lung epithelial cells thereby rapidly and substantially impairing alveolar fluid reabsorption in ARDS patients and contributing to the persistence of their pulmonary edema. A significant negative correlation existed between TGF-β levels in bronchoalveolar lavage fluid (BAL) samples from ARDS patients and ventilator-free days and ICU-free days. Furthermore, lower TGF-β levels correlated with better survival outcome indicating that that patients with higher TGF-β levels may have a higher and faster case mortality. The anti-TGF-β RNA therapeutic Trabedersen/OT-101 exhibited a favourable clinical safety profile in cancer patients and it also exhibited nanomolar in vitro potency against SARS-CoV-2. The correlation of lower BAL fluid TGF-β levels with improved survival of ARDS patients taken together with the potent anti-SARS-CoV-2 activity of Trabedersen/OT101 support the concept of reducing TGF-β levels with Trabedersen/OT-101 in COVID-19 patients with ARDS.

Introduction

After infection with SARS-CoV-2, up to one third of COVID-19 patients develop an acute pulmonary inflammation with a fulminant progression to acute respiratory distress syndrome (ARDS) with a high fatality rate in high risk patient populations despite best available supportive care [1-3]. In these patients, the "burst release" of proinflammatory cytokines in massive amounts and in succession causes a severe form of systemic capillary leak syndrome with pulmonary edema, that can cause hypoxic injury and dysfunction of multiple organs, ultimately leading to an irreversible and fatal multi-organ failure [1-3]. This hyperacute inflammatory process is reminiscent of the cytokine release syndrome (CRS) observed in cancer

patients treated with immunotherapeutic modalities such as CAR-T cells (e.g. tisagenlecleucel) or bispecific T-cell engagers (e.g. blinatumomab) that may hyperactivate elements of both the innate and acquired immune system [4-8]. Although an intensive research effort is underway globally to identify effective antiviral drugs or drug combinations against SARS-CoV-2, it is unlikely that such treatments, including treatments employing drugs such as Remdesivir (ClinicalTrials.gov Identifier: NCT04280705), (ClinicalTrials.gov Favipiravir Identifier: NCT04310228), Hydroxychloroquine (Clinical-Trials.gov Identifier: NCT04318444), albeit of very high clinical impact potential for post-exposure prophylaxis in early treatment settings, could effectively reverse or ameliorate the

pulmonary and systemic inflammation in COV-ID-19 patients with evolving or established ARDS. Therefore, there is an urgent and unmet medical need for treatments that can effectively reduce the risk of ARDS or its mortality rate in COVID-19 patients with pneumonia.

Clinical Safety, Pharmacokinetics, and Activity of Trabedersen/OT-101

Antisense oligodeoxynucleotides are short strings of DNA that are designed to downregulate gene expression by interfering with the translation of a specific encoded protein at the mRNA level. Trabedersen (OT-101; also known as OT101) is a synthetic TGF β 2-specific 18-mer phosphorothioate antisense oligodeoxynucleotide (S-ODN) that was originally designed to reduce the level of TGF^{β2} protein within the tumor microenvironment (TME) of cancer patients in an effort to overcome the TGF-\beta-mediated immune-suppression, invasive tumor growth and generation of new blood vessels supplying tumor tissue (viz.: neovascularization) that contribute to progression of disease [9]. The safety and single agent anti-cancer activity of this RNA therapeutic was evaluated in Phase I and Phase 2 studies which demonstrated its high clinical impact potential for aggressive and difficult-to-treat advanced cancers, including high grade brain tumors, such as glioblastoma multiforme and Grade 3 anaplastic astrocytoma, and pancreas cancer [10-13]. Notably, in a randomized Phase IIb study (Clinicaltrial.gov identifier: NCT00431561), intratumoral delivery of OT-101 via extended convection-enhanced delivery (CED) in the absence of other therapeutic agents or radiation resulted in >3.5 year OS in more than half of a recurrent/refractory (R/R)high-grade glioma patient population [12, 13]. OT-101 showed remarkable single agent activity with more than a third of patients (26 of 77) receiving the intended 4-11 cycles of therapy achieving durable complete responses, partial responses, or prolonged stable disease and a median overall survival of >4.5 years.

Likewise, in a Phase I/II study (ClinicalTrials.gov identifier: NCT00844064) promising clinical activity and better than expected overall survival was observed in metastatic solid tumor patients, including a Stage IV pancreatic cancer patient who achieved a sustained complete response lasting >5 years [10, 11]. In the latter study, a total of 61 patients with Stage III/IV pancreatic cancer (n=37), malignant melanoma (n=19), or colorectal carcinoma (n=5) were treated with continuous intravenous (IV) infusion of OT-101 as 2nd to 4th-line therapy in escalating doses (40 mg/m²/day up to 330 mg/m²/ day) of two treatment schedules (initial schedule: 7-days on, 7-days off (n = 17); modified schedule: 4-days on, 10-days off (n = 44); two cycles as core study and up to 10 cycles for expanded study). A summary of the number of patients per cohort and indication, together with the median number of cycles and daily trabedersen dose is provided in Table 1.

Table 1: Number of Patients Participating in the Core Study, by Indication, Schedule, and Cohort (FAS)

	No. of Patients					
Treatment Schedule Cohort	Pancreatic cancer	Melanoma	Col- orectal cancer	Total		
Total	37	19	5	61		
7-days-on, 7-days-off	11	2	4	17		
Cohort 1: 40 mg/m²/day	4	0	0	4		
Cohort 2: 80 mg/m²/day	2	1	0	3		
Cohort 3: 160 mg/m²/day	3	1	2	6		
Cohort 4: 240 mg/m²/day	2	0	2	4		
4-days-on, 10-days-off	26	17	1	44		
Cohort A1: 140 mg/m²/day	5	0	0	5		
Cohort A2: 190 mg/m²/day	2	1	0	3		
Cohort A3: 250 mg/m²/day	4	1	0	5		
Cohort A4: 330 mg/m²/day	1	1	1	3		
Last cohort: 140 mg/m ² /day	14	14	0	28		

The planned number of 3 patients per cohort of the dose-escalation period was reached for all cohorts: the number of patients in these cohorts varied between 3 to 6. Fourteen additional pancreatic cancer patients and 14 additional melanoma patients were enrolled in the last cohort. Consequently, the number of patients per cohort of the 7-days-on, 7-daysoff and 4-days-on, 10-days-off dose escalation schedules varied between 3 and 6 patients. The last cohort treated with 140 mg/m²/day with the 4 days on, 10-days-off schedule contained 28 patients (14 each with pancreatic cancer and melanoma). The median number of cycles per cohort varied between 1.5 and 5.0. The median trabedersen dose per day during the core study period reflected the respective dose cohort,

with the highest median dose per day seen for cohort 4 of the 7-days-on, 7-days-off schedule $(167.9 \text{ mg/m}^2/\text{day})$ (Table 2).

Treatment Schedule	No. of patients	No. of cycles	Daily dose core study period			
Cohort	Total	Median	(mg/m²/day)			
	(PanCa/MM/CRC)	(min; max)	Median (Q1, Q3)			
7-days-on, 7-days-off						
Cohort 1: 40 mg/m²/day	4 (4/0/0)	2.0 (1; 4)	23.1 (20.2, 34.3)			
Cohort 2:80 mg/m²/day	3 (2/1/0)	5.0 (4; 10)	40.0 (39.9, 40.7)			
Cohort 3: 160 mg/m²/day	6 (3/1/2)	2.5 (1; 4)	69.1 (50.4, 83.4)			
Cohort 4: 240 mg/m²/day	4 (2/0/2)	1.5 (1; 2)	167.9 (133.6, 186.3)			
4-days-on, 10-days-off						
Cohort A1: 140 mg/m²/day	5 (5/0/0)	4.0 (1; 10)	33.0 (21.5, 40.8)			
Cohort A2: 190 mg/m ² /day	3 (2/1/0)	4.0 (2; 8)	56.3 (56.1, 57.5)			
Cohort A3: 250 mg/m²/day	5 (4/1/0)	4.0 (1; 10)	59.8 (58.2, 76.0)			
Cohort A4: 330 mg/m²/day	3 (1/1/1)	2.0 (2; 10)	79.7 (75.3, 82.2)			
Last cohort (4-days-on, 10-da	ys-off)					
140 mg/m²/day	28 (14/14/0)	4.0 (1; 8)	38.1 (31.8, 40.1)			
CRC = colorectal cancer; MM = malignant melanoma; No. = number; PanCa = pancreatic cancer; Q1 = first quartile; Q3 = third quartile						

Table 2: Extent of Exposure (FAS)

On average, patients in the first treatment schedule were exposed to study medication for 2.7 cycles, corresponding to a treatment period of 1.2 months. The total estimated average exposure was 2248 mg/m². In the second treatment schedule, patients on average were exposed for 4.4 cycles, corresponding to a treatment period of 2 months. The total estimated average exposure was 2734 mg/m².

Overall, Trabedersen was well-tolerated. The majority of the AEs occurring during treatment with OT-101 were most likely related to the underlying disease or associated symptoms rather than the study medication itself. A total of 30 patients (49.2%) reported 87 TEAEs considered possibly related to trabedersen by the Investigator, with slightly higher frequency in the 4-days-on, 10-days-off schedule group (24 patients [54.5%]) than in the 7-days-on, 7-daysoff schedule group (6 [35.3%]) (Table 3).

Six patients (9.8%) overall reported 21 TEAEs considered related to trabedersen by the Investigator, 2 patients (11.8%) with the 7-days-on, 7-days-off schedule and 4 (9.1%) with the 4-days-on, 10-days-off schedule (Table 3, Table 4).

The only related TEAE reported by > 1 patient was thrombocytopenia that occurred in 2 patients of each schedule group. All cases of thrombocytopenia and decreased platelet count reported during the study were considered related or possibly related to trabedersen by the Investigator (Table 3, Table 4).

Table 3: Possibly Related and Related Adverse Events, by Preferred Term and Schedule and Sorted by Frequency (FAS)

	No. (%) of patients with TEAE / No. of TEAEs							
	Pos	sibly related TE	AEs	Related TEAEs				
Preferred Term	7-days- on, 7-days-	4-days-on, 10-days-off schedule	Total (N=61)	7-days-on, 7-days-off schedule	4-days- on, 10-days-	Total (N=61)		
	off schedule (N=17)	(N=44)		(N=17)	off schedule (N=44)			
Any TEAE	6 (35.3)/12	24 (54.5)/75	30 (49.2)/87	2 (11.8)/2	4 (9.1)/19	6 (9.8)/21		
Fatigue	1 (5.9)/1	5 (11.4)/6	6 (9.8)/7	0	1 (2.3)/1	1 (1.6)/1		
Nausea	0	5 (11.4)/8	5 (8.2)/8	0	1 (2.3)/4	1 (1.6)/4		
Thrombocytpenia	0	5 (11.4)/5	5 (8.2)/5	2 (11.8)/2	2 (4.5)/2	4 (6.6)/4		
Headache	0	4 (9.1)/7	4 (6.6)/7	0	1 (2.3)/1	1 (1.6)/1		
Vomiting	1 (5.9)/1	3 (6.8)/4	4 (6.6)/5	0	1 (2.3)/1	1 (1.6)/1		
Dyspnea	0	3 (6.8)/4	3 (4.9)/4	0	0	0		
Pyrexia	0	2 (4.5)/3	2 (3.3)/3	0	1 (2.3)/4	1 (1.6)/4		
Asthenia	0	2 (4.5)/2	2 (3.3)/2	0	0	0		
Constipation	0	2 (4.5)/2	2 (3.3)/2	0	0	0		
ppetite	0	2 (4.5)/2	2 (3.3)/2	0	0	0		
Diarrhea General physical health deterio-	0	2 (4.5)/2	2 (3.3)/2	0	0	0		
ration	0	2 (4.5)/2	2 (3.3)/2	0	0	0		
Pain in extremity	1 (5.9)/1	1 (2.3)/1	2 (3.3)/2	0	0	0		
Platelet count								
decreased	1 (5.9)/1	1 (2.3)/1	2 (3.3)/2	0	1 (2.3)/3	1 (1.6)/3		
Pruritus	0	2 (4.5)/2	2 (3.3)/2	0	0	0		
Chills	0	1 (2.3)/1	1 (1.6)/1	0	1 (2.3)/1	1 (1.6)/1		
Metastasis	0	0	0	0	1 (2.3)/1	1 (1.6)/1		
Skin Inflamation	0	0	0	0	1 (2.3)/1	1 (1.6)/1		

Table 4.Grade ≥3 Adverse Events Assessed as Related or Possibly Related to Trabedersen (OT-101) by Investigator

- 0					
No. of AE	Preferred Term of AE	Grade	Seriousness	Outcome	Patient No.
3	Thrombocytopenia	3	Nonserious	Recovered	PC 1017/4 MM 1027/A3 MM 1055/LC
1	Thrombocytopenia	3	Nonserious	Recovered with sequelae	CC 1016/4
1	ALAT increased	3	Nonserious	Recovered	PC 1022/A1
1	GGT increased	3	Nonserious	AE still present	PC 1005/2
1	Haemoglobin decreased	3	Nonserious	Recovered	MM 1055/LC
1	Rash maculo-pap- ular	3	Nonserious	Recovered	CC 1014/4
1	Gastrointestinal hemorrhage	3	Serious	Recovered	PC 1018/A1

Within Preferred Terms events are listed by seriousness, grade and outcome. Events are sorted by descending frequency.

AE = adverse event; CC = colorectal cancer; MM = malignant melanoma; Pat. No. = patient number; PC = pancreatic cancer

A total of 4 patients experienced a DLT in the dose escalation part of the study (core study) as per regulations applied during the study conduct, with three of these patients treated at 240 mg/m²/day (Cohort 3, Table 1) in the 7 days on, 7 days off schedule (2 Grade 3 thrombocytopenias, 1 Grade 3 maculo-papular rash). Therefore, a dose of 160 mg/m²/day was declared to be the MTD for the 7-days-on, 7 days off schedule. The second schedule (4 days on, 10 days off) was completed without reaching the MTD. In the Phase II-part of the study additional pancreatic cancer and melanoma patients were treated with 140 mg/m²/d according to the second schedule.

There were 2 SAEs which were considered as possibly related (1 SAE of gastrointestinal hemorrhage, 1 SAE of pyrexia, both with outcome recovered). The first related SAE was a case of gastrointestinal hemorrhage which occurred in a 65 year old male patient with pancreatic cancer. The event occurred 4 days after end of the first treatment cycle with 140 mg/ m²/day (4 days on, 10 days off). Gastroduodenoscopy showed an acute duodenal varix bleeding. The bleeding was stopped by sclerotherapy and the outcome was reported as recovered. The second related SAE was a case of pyrexia (verbatim term fever and chills) which occurred in a 66 year old male malignant melanoma patient. Pyrexia (NCI CTC grade 1) occurred about 12 hours after start of the second treatment cycle of OT-101 with 140 mg/m²/day (4 days on, 10 days off) and got worse (NCI-CTC grade 2), requiring the patient's hospitalization on the third treatment day. The patient was treated with antibiotics (orally and intravenously) for suspected gastrointestinal infection and subsequently recovered from pyrexia. The patient had already suffered from gastrointestinal infection, from which he had recovered under antibiotic therapy prior to start of the second treatment cycle with OT-101. The investigator did not suspect the study drug to have caused pyrexia, but suspected that the treatment with OT-101 could have worsened these symptoms.

As thrombocytopenia has been identified as adverse drug reaction associated with trabedersen, the SOC Blood and lymphatic system disorders is looked at more closely. 12 of 61 patients (19.7%) suffered 14 AEs of either thrombocytopenia or platelet count decreased. All events were assessed as related/possibly related to trabedersen by the investigators and the vast majority (10 out of 14 events) was reported to have recovered without further treatment. None of these cases was assessed as serious. The intensity of 10 of the 14 occurrences was reported as mild or moderate whereas 4 AEs were reported to be of severe intensity. In 6 instances therapy with OT-101 was discontinued or temporarily interrupted. No bleedings were reported and no platelet transfusions became necessary. There were no signs of cumulative increases of thrombocytopenia in later treatment cycles.

Hence, thrombocytopenia is a known side effect of other phosphorothioate oligonucleotides and was identified as an expected adverse drug reaction during treatment with Trabedersen/OT-101. A close temporal relationship between the administration of Trabedersen/OT101 and the onset of the AE of thrombocytopenia and the fact that thrombocytopenia often occurs at the beginning of the treatment with trabedersen during cycle 1 or cycle 2 reinforce the existence of a causal relationship.

In PK analyses, the concentration time course of trabedersen/OT-101 was best described by a two-compartment model [14]. The distribution of trabedersen to tissues was not only rapid, but also extensive, with deep tissue penetration as demonstrated by the finding that the volume of the peripheral compartment far exceeded the total body water volume [14]. Plasma concentrations of trabedersen/OT-1011 began to be quantifiable up to 2 hours post start of infusion (SOI) for most subjects and generally fell below the lower limit of detection a few hours after the end of infusion, even for the high dose groups. PK profiles of trabedersen/ OT-101 show sustained plasma concentrations throughout the dosing period (4 or 7 days) with similar PK exposure parameters (Cmax and Area under curve (AUC)) between Cycle 1 and Cycle 2. Exposure was dose proportional for trabedersen over the 7-day dose range (40, 80, 160 and 240 mg/mg²/day) and the 4-day dose range (140, 190, 250, and 330 mg/m²/day) (Table 5).

	7-days-on, 7-days-off schedule				4-days-on, 10-days-off schedule			
	40 mg/m ²	80 mg/m ²	160 mg/ m ²	240 mg/ m ²	140 mg/ m ²	190 mg/m ²	250 mg/m ²	330 mg/m ²
Cycle 1								
Tmax (h)	168	114	139	119	58.0	70.2	63.5	62.3
Cmax (µg/mL)	0.961	1.46	2.86	4.45	2.17	3.24	2.95	3.90
Tlast (h)	170	171	173	173	134	181	101	102.0
AUC (0-t) (μg*h/mL)	93.0	141	383	559	173	278	254	324
AUC (0-168) (μg*h/mL)	92.4	140	379	593	160	240	249	316
T1/2 (h)	0.594	-	1.70	1.60	1.10	1.23	1.25	1.53
Vd (mL/m²)	1902	-	7855	6783	5644	5310	7682	9029
Cl (mL/m ²)	2222	-	3309	2940	3728	2904	5126	4076
Cycle 2								
Tmax (h)	169	114	119	-	66.3	47.9	95.0	59.8
Cmax (µg/mL)	0.482	1.36	2.38	-	2.43	3.31	3.68	3.50
Tlast (h)	170	171	179	-	138	124	236	102
AUC (0-t) (μg*h/mL)	53.1	127	356	-	193	271	523	305
AUC (0-168) (μg*h/mL)	52.4	125	354	-	185	253	298	298
T1/2 (h)	-	-	1.49	-	1.17	1.23	1.63	1.78
Vd (mL/m²)	-	-	6944	-	5097	4999	6327	11233
Cl (mL/m ²)	-	-	3076	-	3118	2802	2801	4364

Table 5: Pharmacokinetic Parameters of Trabedersen after Intravenous Infusions of Trabedersen, by Cohort

The mean Cmax values on Schedule 1, Cycle 1 were 0.96 μ g/mL at 40 mg/m²/day, 1.46 μ g/mL at 80 mg/m²/day, 2.86 μ g/mL at 160 mg/m²/day, and 4.62 μ g/mL at 240 mg/m2/ day. The recovery phase at the end of the profiles declined in a linear manner for most subjects. The terminal elimination half-life (T1/2)was well determined from the recovery phase of most trabedersen profiles, and was approximately between 1 and 2 hours. The pharmacokinetic profile of trabedersen was in line with the short half-life expected for this class of agents. Observed plasma T1/2 was between 1 to 2 hours. This naturally requires a prolonged infusion as employed for this trial to ideally achieve extended target suppression, as for first-generation oligonucleotides a prolonged tissue half-life cannot be expected.

Role of TGF-β in ARDS, SARS, COVID-19

ARDS is characterized by severe pulmonary edema caused by an endothelial cell damage with increased capillary permeability contributing to accumulation of a protein-rich viscous fluid in alveoli and interstitium combined with an impaired alveolar fluid clearance incapable of remove the edema from alveoli owing to dysregulated Na+ and fluid transport across the alveolo-capillary barrier caused by a functional defect of the epithelial sodium channel (ENaC). Alveolar barrier dysfunction in ARDS leads to more pulmonary edema and to the systemic release of biological mediators from the lung, contributing to the failure of other organs and potentially a multi-organ failure [15].

TGF-β has been implicated as an important pro-inflammatory cytokine in the pathophysiology of acute lung injury and ARDS that contributes to both increased permeability and failed fluid reabsorption in lungs leading to persistent and severe pulmonary edema [16-21]. Importantly, lavage fluid (BAL) samples from ARDS patients collected within 2 days after intubation showed higher TGF-B levels when compared to BAL samples from non-ARDS controls [22]. TGF-β can increase alveolar epithelial permeability [20] and pulmonary endothelial permeability by promoting adherens junction disassembly [23] as well as inhibiting pulmonary endothelial proliferation [24]. SARS-CoV has been shown to up-regulate pro-inflammatory cytokines, including TGF-β and TGF-β levels were markedly elevated in SARS patients with ARDS [25]. SARS-associated coronavirus (SARS-CoV) nucleocapsid (N) protein has

been shown to potentiate TGF- β signalling via a Smad3-dependent induction of TGF- β 1 expression [26]. Further, the papain-like protease (PLpro) of SARS-CoV, a deubiquitinating enzyme and virulent factor in SARS pathogenesis, has been reported to trigger TGF- β production via ubiquitin proteasome, p38 MAPK, and ERK1/2-mediated signaling [27, 28].

The contributing factors for the increased TGF-B levels in ARDS also include the activation of the complement signalling pathway with production of the complement cleavage product, C5a that triggers the formation of neutrophil extracellular traps (NETs) that are capable of activating platelets to release TGF^β. A recent study by Peters et al. demonstrated that TGF-β profoundly impacts alveolar ion and fluid transport by regulating the epithelial sodium channel (ENaC) activity and trafficking via a Tgfbr1-mediated unique signalling pathway [29]. TGF- β was identified as the exclusive master regulator of ENAC internalization by alveolar epithelial cells and its upregulation in ARDS causes an ENAC trafficking defect with marked reduction in the cell-surface abundance of ENaC on lung epithelial cells thereby rapidly and substantially impairing alveolar fluid reabsorption in ARDS patients and contributing to the persistence of their pulmonary edema [29]. A soluble recombinant TGF- β receptor protein capable of sequestering TGF- β has effectively attenuated the severity of pulmonary edema in experimental models of ARDS [20, 30]. Likewise, the anti-inflammatory isoflavone Puerarin has been shown to reduce the ARDS-associated inflammatory process in the lungs by inhibiting the expression of TGF-β [31]. Notably, a significant negative correlation existed between TGF-β levels in bronchoalveolar lavage fluid (BAL) samples from ARDS patients and ventilator-free days and ICU-free days. Furthermore, lower TGF-β levels correlated with better survival outcome indicating that that patients with higher TGF- β levels may have a higher and faster case mortality [22]. Notably, OT-101 exhibited nanomolar in vitro potency against both SARS-CoV and SARS-CoV-2 and it was at least 1 log superior to the antiviral agent Remdesivir (Table 6, Figure 1).

Table 6. An	tiviral activity (01-10	1	
Com-	Virus	EC	CC	SI
pound		50	50	
OT-101	SARS-CoV-1	7.6	>1000	>130
	(Urbani strain) ¹	(1.24 μM)		
OT-101	SARS-CoV-1	26	>1000	>38
	(Urbani strain)	(4.23 μM)		
OT-101	SARS-COV-2	2.0	>1000	>500
	USA_ 2 WA1/2020	(0.33 μM)		
RSV	SARS-COV-2 USA_ 2 WA1/2020	620.0	>1000	>1.6
M128533	SARS-COV-2 USA_ 2 WA1/2020	0.012	>10	>830
Remde- sivir	SARS-COV-2	1.06	>10	>830
	(Wang M. et al., 2020, Cell Res. 30:269-271)	(1.76 μM)		
RSV- Negative (control antisense/M	128533- po	sitive contro	

RSV- Negative control antisense/M128533- positive control EC50: 50% effective antiviral concentration (in μg/ml) / CC50: 50% cytotoxic concentration of compound without virus added (in μg/ml) / SI = CC50/EC50

¹Source: Centers for Disease Control Stock 809940 (200300592) ²Source: The World Reference Center for Emerging Viruses and Arboviruses (WRCEVA) at UTMB



Figure 1. Trabedersen is a Nanomolar Inhibitor of SARS-CoV-2 Replication. The in vitro antiviral activity against SARS-CoV-2 (USA_WA1/2020 strain) was tested in Vero 76 cells in collaboration with Dr. Brett Hurst at Utah State University that is part of the NIAID Antiviral Testing Consortium. The cytotoxicity of OT101 against Vero 76 cells was also tested in the absence of SARS-CoV-2. See Table 6 for further details.

The correlation of lower BAL fluid TGF- β levels with improved survival of ARDS patients taken together with the potent antiviral activity of Trabedersen/OT101 against SARS-CoV-2 further support the concept of reducing TGF- β levels with Trabedersen/OT-101 in COVID-19 patients with ARDS [22].

TGF-β is also involved in the pathogenesis of lung tissue remodelling and lung fibrosis that follows ARDS. Specifically, TGF- β contributes to the development of lung fibrosis by stimulating the pro-liferation/differentiation of lung fibroblasts, accumulation of collagen and other extracellular matrix proteins in the pulmonary interstitial and alveolar space, leading to the occurrence and development of pulmo¬nary fibrosis [32, 33]. Wang et al. reported that miR-425 reduction in lung fibroblasts contributes to the development of lung fibrosis post ARDS through activation of the TGF-β signalling pathway [34]. Smad2, a key component of the canonical TGF-β signalling pathway was discovered to be regulated by miR-425 [34]. Therefore, inhibition of the TGF- β signalling pathway also has the potential to prevent development of pulmonary fibrosis following ARDS and improve the pulmonary healing process [31].

Clinical Development Strategy for COVID-19 Patients

Based on the clinical safety profile and PK of Trabedersen in cancer patients, we have designed our Phase I/II study in COVID-19 patients as a two-part study. Trabedersen will be used in combination with standard of care [35, 36] in COVID-19 patients with hypoxemic respiratory failure receiving either non-invasive positive pressure ventilation (NIPPV) or mechanical ventilation (MV). The eligibility criteria are listed in Table 7.

Table 7. Main Inclusion and Exclusion CriteriaA. Inclusion Criteria

1. Male or non-pregnant, non-lactating female subject with SARS-CoV-2 (previously known as 2019-nCoV) infection that is documented by an FDA-authorized diagnostic RT-PCR test (a) at or within 4 days of screening or (b) at baseline

2. \geq 18 years AND \leq 70 years of age

3. History of severe COVID-19 within the last 2 weeks prior to study enrollment as defined by: Respiratory distress \geq 30 breaths/ min, oxygen saturation \leq 93% at rest in ambient air; supplemental oxygen requirement to keep oxygen saturation \geq 90%

4. The patient or a legally authorized rep

resentative has provided written informed consent

5. The patient or the authorized representative is aware of the investigational nature of this study and willing to comply with protocol treatments, blood tests, and other evaluations listed in the informed consent form

6. Hospitalized with hypoxemic respiratory failure receiving either (a) non-invasive positive pressure ventilation (NIPPV) with Continuous Positive Airway Pressure (CPAP) or Bilevel Positive Airway Pressure (BiPAP) (clinical status: 3 on the 8-point ordinal scale) or (b) mechanical ventilation (clinical status: 2 on the 8-point ordinal scale)

7. History of worsening respiratory distress over the last 7 days

8. NIPPV or mechanical ventilation initiated ≤4 days prior to enrolment

9. Patients receiving mechanical ventilation must have acute respiratory distress syndrome (ARDS)

10. Patient's ARDS is MILD according to Berlin Definition with PaO2/FiO2 >200 AND \leq 300 mmHg while on positive end-expiratory pressure (PEEP) or CPAP \geq 5 cm H2O.

11. Bilateral opacities on a chest X-ray or Chest CT scan.

Ejection fraction on cardiac echo ≥40%
Grade 1 or less on CTCAE for red blood cells/haemoglobin, WBC and platelets.

B. Exclusion Criteria

1 Moderate or Severe ARDS by Berlin definition with PaO2/FiO2 \leq 200 mmHg on a PEEP or CPAP \geq 5 cm H2O; Extracorporeal membrane oxygenation (ECMO)

2 Uncontrolled hypertension (systolic blood pressure (BP) > 150 mmHg and/or diastolic BP > 100 mmHg), unstable angina, congestive heart failure (CHF) of any New York Heart Association (NYHA) classification, serious cardiac arrhythmia requiring treatment (exceptions: atrial fibrillation, paroxysmal supraventricular tachycardia), history of myocardial infarction within 12 months of enrollment;

3 Hypotension requiring vasoactive peptides, such as dopamine, norepinephrine, epinephrine, or dobutamine

4 Renal function impairment with Creatinine >2 mg/dL; Liver function impairment with Bilirubin >2 mg/dL; Platelet count <50,000/ μ L; Multi-organ failure

5 Documented active infection with a bacterial pathogen requiring parenteral systemic antibiotics; Bacterial or fungal sepsis

6 History of live vaccination within the last 4 weeks prior to study enrollment; Patients must not receive live, attenuated influenza vaccine (e.g., FluMist) within 4 weeks before enrolment or at any time during the study

7 History of an allergic reaction or hypersensitivity to Trabedersen/OT101 **REFERENCES**

1. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet. 2020;395:497-506. doi: 10.1016/S0140-6736(20)30183-5.

2. Young BE, Ong SWX, Kalimuddin S, Low JG, Tan SY, Loh J, Ng OT, Marimuthu K, Ang LW, Mak TM, Lau SK, Anderson DE, Chan KS, Tan TY, Ng TY, Cui L, Said Z, Kurupatham L, Chen MI, Chan M, Vasoo S, Wang LF, Tan BH, Lin RTP, Lee VJM, Leo YS, Lye DC; Singapore 2019 Novel Coronavirus Outbreak Research Team. Epidemiologic Features and Clinical Course of Patients Infected With SARS-CoV-2 in Singapore. JAMA. 2020 Mar 3. doi: 10.1001/ jama.2020.3204.

3. Zumla A, Hui DS, Azhar EI, Memish ZA, Maeurer M. Reducing mortality from 2019-nCoV: host-directed therapies should be an option. Lancet. 2020;395(10224):e35-e36. doi: 10.1016/S0140-6736(20)30305-6.

4. Maude SL, Barrett D, Teachey DT, Grupp SA. Managing Cytokine Release Syndrome Associated With Novel T Cell-Engaging Therapies. Cancer J. 2014 Mar-Apr; 20(2): 119–122. doi: 10.1097/ PPO.000000000000035

5. Davila ML, Riviere I, Wang X, Bartido S, Park J, Curran K, Chung SS, Stefanski J, Borquez-Ojeda O, Olszewska M, Qu J, Wasielewska T, He Q, Fink M, Shinglot H, Youssif M, Satter M, Wang Y, Hosey J, Quintanilla H, Halton E, Bernal Y, Bouhassira DC, Arcila ME, Gonen M, Roboz GJ, Maslak P, Douer D, Frattini MG, Giralt S, Sadelain M, Brentjens R. Efficacy and toxicity management of 19-28z CAR T cell therapy in B cell acute lymphoblastic leukemia. Sci Transl Med. 2014;6(224):224ra25.

6. Teachey DT, Rheingold SR, Maude SL, Zug

maier G, Barrett DM, Seif AE, Nichols KE, Suppa EK, Kalos M, Berg RA, Fitzgerald JC, Aplenc R, Gore L, Grupp SA. Cytokine release syndrome after blinatumomab treatment related to abnormal macrophage activation and ameliorated with cytokine-directed therapy. Blood. 2013;121(26):5154-5157.

7. Khadka RH, Sakemura R, Kenderian SS, Johnson AJ. Management of cytokine release syndrome: an update on emerging antigen-specific T cell engaging immunotherapies. Immunotherapy 2019; 11:10. https://doi.org/10.2217/imt-2019-0074

8. Garcia Borrega J, Gödel P, Rüger MA, Onur ÖA, Shimabukuro-Vornhagen A, Kochanek M, Böll B. In the Eye of the Storm: Immune-mediated Toxicities Associated With CAR-T Cell Therapy. HemaSphere 2019;3:2.

9. Schlingensiepen KH, Jaschinski F, Lang SA, Moser C, Geissler EK, Schlitt HJ, Kielmanowicz M, Schneider A. Transforming growth factor-beta 2 gene silencing with trabedersen (AP 12009) in pancreatic cancer. Cancer Sci. 2011; 102(6):1193-200. doi: 10.1111/j.1349-7006.2011.01917.x.

10. Oettle H, Seufferlein T, Luger T, Schmid RM, Wichert G, Endlicher E, Garbe C, Kaehler KK, Enk A, Schneider A, Rothhammer-Hampl T, Grosser S, Kiessling P. Final results of a phase I/II study in patients with pancreatic cancer, malignant melanoma, and colorectal carcinoma with trabedersen. Journal of Clinical Oncology 2012; 30 (15): 4034. DOI: 10.1200/jco.2012.30.15_suppl.4034 Journal of Clinical Oncology 30, no. 15_suppl (May 20, 2012) 4034-4034.

11. Hwang L, Ng K, Wang W, Trieu V. Treatment with trabedersen, an anti-TGF-beta 2 antisense, primed tumors to subsequent chemotherapies. [abstract]. In: Proceedings of the 107th Annual Meeting of the American Association for Cancer Research; 2016 Apr 16-20; New Orleans, LA. Philadelphia (PA): AACR; Cancer Res 2016;76(14 Suppl):Abstract nr 3742.12.

12. Uckun FM, Qazi S, Hwang L, Trieu VN. Recurrent or Refractory High-Grade Gliomas Treated by Convection-Enhanced Delivery of a TGF β 2-Targeting RNA Therapeutic: A Post-Hoc Analysis with Long-Term Follow-Up. Cancers (Basel). 2019 Nov 28;11(12). pii: E1892. doi: 10.3390/cancers11121892.

13. Uckun FM, Trieu VN. Monotherapy of High-Grade Gliomas with a TGF-beta2 Targeting RNA Therapeutic. Cancer Clin J. 2019; 1(1): 1007

14. Wang W, Ng K, Nam D, Trieu V, Hwang L. Population pharmacokinetic model for OT-101 - A TGF- β 2-specific antisense oligonucleotide in cancer patients [abstract]. In: Proceedings of the American Association for Cancer Research Annual Meeting

2017; 2017 Apr 1-5; Washington, DC. Philadelphia (PA): AACR; Cancer Res 2017;77(13 Suppl):Abstract nr 5043. doi:10.1158/1538-7445.AM2017-5043

15. Imai Y, et al. Injurious mechanical ventilation and endorgan epithelial cell apoptosis and organ dysfunction in an experimental model of acute respiratory distress syndrome. JAMA 2003; 289(16):2104–2112.

16. Frank J, et al. Transforming growth factor-β1 decreases expression of the epithelial

sodium channel alphaENaC and alveolar epithe lial vectorial sodium and fluid

transport via an ERK1/2-dependent mechanism. J Biol Chem 2003; 278(45):43939–43950.

17. Fahy RJ, et al. The acute respiratory distress syndrome: A role for transforming

growth factor-beta 1. Am J Respir Cell Mol Biol 2003; 28(4):499–503.

18. Willis BC, et al. Modulation of ion conductance and active transport by TGF- β 1

in alveolar epithelial cell monolayers. Am J Physiol Lung Cell Mol Physiol 2003; 285(6):

L1192-L1200.

19. Jenkins RG, et al. Ligation of protease-activated receptor 1 enhances $\alpha(v)\beta6$

integrin-dependent TGF- β activation and promotes acute lung injury. J Clin Invest 2006;

116(6):1606–1614. 20. Pittet JF, Griffiths MJ, Geiser T, Kaminski N, Dalton SL, Huang X, Brown LA, Gotwals PJ, Koteliansky VE, Matthay MA, Sheppard D. TGF-beta is a critical mediator of acute lung injury. J Clin Invest 2001; 107:1537–1544

21. Frank JA, Matthay MA. TGF-β and lung fluid balance in ARDS. PNAS 2014; 111: 885-886

22. Budinger S, Chandel NS, Donnelly HK, Eisenbart J, Oberoi M, Jain M. Active transforming growth factor-b1 activates the procollagen I promoter in patients with acute lung injury. Intensive Care Med 2005; 31:121–128. DOI 10.1007/s00134-004-2503-2

23. Hurst VI, Goldberg PL, Minnear FL, Heimark RL, Vincent PA. Rearrangement of adherens junctions by transforming growth factor-beta1: role of contraction. Am J Physiol 1999; 276:L582–595

24. Das SK, White AC, Fanburg BL. Modulation of transforming growth factor-beta 1 antiproliferative effects on endothelial cells by cysteine, cystine and N-acetylcysteine. J Clin Invest 1992; 90:1649– 1656

25. Lee C-H, Chen R-F, Liu J-W et al., Altered p38 Mitogen-Activated Protein Kinase Expression in Different Leukocytes with Increment of Immunosup pressive Mediators in Patients with Severe Acute Respiratory Syndrome. J Immunol 2004; 172:7841-7847 26. Zhao X, Nicholls JM, Chen Y-G. Severe Acute Respiratory Syndrome-associated Coronavirus

Nucleocapsid Protein Interacts with Smad3 and Modulates Transforming Growth Factor-Signaling. J. Biol. Chem. 2008; 283: 3272-3280

27. Li SW1, Yang TC, Wan L, Lin YJ, Tsai FJ, Lai CC, Lin CW. Correlation between TGF- β 1 expression and proteomic profiling induced by severe acute respiratory syndrome coronavirus papain-like protease. Proteomics. 2012 Nov;12(21):3193-205. doi: 10.1002/pmic.201200225. Epub 2012 Oct 5.

28. Wang CY, Lu CY, Li SW, Lai CC, Hua CH, Huang SH, Lin YJ, Hour MJ, Lin CW. SARS coronavirus papain-like protease up-regulates the collagen expression through non-Samd TGF- β 1 signaling. Virus Res. 2017 May 2;235:58-66. doi: 10.1016/j.virus-res.2017.04.008.

29. Peters DM, Vadasz I, Wujak T et al., TGF- β directs trafficking of the epithelial sodium

channel ENaC which has implications for ion and fluid transport in acute lung injury. PNAS 2013; E374–E383www.pnas.org/cgi/doi/10.1073/ pnas.1306798111

30. Bossman M, Ward PA. Protein-based Therapies for Acute Lung Injury: Targeting Neutrophil Extracellular Traps. Expert Opin Ther Targets. 2014 June ; 18(6): 703–714. doi:10.1517/14728222.201 4.902938.

31. Hu X, Huang X. Alleviation of Inflammatory Response of Pulmonary Fibrosis in Acute Respiratory Distress Syndrome by Puerarin via Transforming Growth Factor (TGF-b1).

Med Sci Monit, 2019; 25: 6523-6531

32. Shimbori C, Bellaye PS, Xia J et al: Fibroblast growth factor-1 attenuates TGF-beta1-induced lung fibrosis. J Pathol, 2016; 240(2): 197–210

33. Nithiananthan S, Crawford A, Knock JC et al: Physiological fluid flow mod¬erates fibroblast responses to TGF-beta1. J Cell Biochem, 2017; 118(4): 878–90

34. Wang L, Liu J, Xie W, Li G, Yao L, Zhang R, Xu B. miR-425 reduction causes aberrant proliferation and collagen synthesis through modulating TGF- β / Smad signaling in acute respiratory distress syndrome. Int J Clin Exp Pathol 2019;12(7):2604-2612 35. Ranieri VM et al., Acute respiratory distress syndrome: the Berlin Definition. JAMA 2012; 307(23):2526-33

36. Papazian L et al. Formal guidelines: management of acute respiratory distress syndrome. Annals of Intensive Care 2019; 9:69

37. Zhang Y, Li J, Zhan Y, Wu L, Yu X et al., Analysis of Serum Cytokines in Patients with Severe Acute Respiratory Syndrome. Infection and Immunity 2004; p. 4410–4415