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A Usually Daptomycin-Resistant Gram-Positive Dermabacter Hominis: Rare Pathogen

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Abstract

Dermabacter hominis is a gram-positive, catalase-positive, glucose-fermenting rod, which, as it grows forms small greyish-white colonies with a characteristic pungent odor. Previously known as coryneform Centers for Disease Control and Prevention groups 3 and 5, it was catalogued as D. hominis in 1994. Various strains isolated in blood cultures, abscesses, or wounds in the 1970s were retrospectively characterized in referral centers as D. hominis. In this report we describe two patients with severe underlying pathology who developed bacteremias by D. hominis within the context of their clinical pictures.

INTRODUCTION

A 29-year-old woman was human immunodeficiency virus positive in stage B3 of AIDS. She was admitted to our hospital with left hemiparesia. A cerebral computer tomography scan showed a hypodense lesion in the left parietal lobe compatible with progressive multifocal leukoencephalopathy. A peripheral venous catheter was placed. While in the hospital the patient suffered a marked deterioration of her neurological situation, together with severe worsening of her general state, with alteration of pulmonary function, upper digestive tract hemorrhage, and fever peaks. Various antimicrobial therapies were administered, including clindamycin, ciprofloxacin, and the combination of fluconazole, meropenem, and sulfamethoxazole, in spite of the fact that no pathogenic microorganism was isolated in the various cultures requested, including the catheter culture. The patient's situation worsened and reached terminal stage, which led to the decision to suspend all medication and to maintain only sedation. Dermabacter hominis and Candida albicans were isolated from two blood cultures taken 48 h earlier. Treatment with fluconazole and vancomycin was initiated, but the patient died 24 h later.

A 65-year-old male patient had a personal history of cardiopathy and broncho-obstructive pneumopathy. He was brought to our Emergency Service with thoracic pain, for which he was admitted to the Cardiology Department. While in the hospital he suffered a cardiorespiratory arrest, from which he recovered after cardiopulmonary reanimation although he was left with irreversible neurological sequelae. During the following days he suffered episodes of fever peaks for which antibiotic treatment with cloxacillin plus gentamicin was administered. After 4 days without fever and without antibiotic treatment he presented with a febrile peak of more than 38°C. At this point two blood cultures were taken and the catheter was removed for culture. The patient's condition deteriorated, and he died suddenly 48 h later. That same day D. hominis grew in the two blood cultures. The catheter culture was negative.

In both patients small gram-positive coccobacilli with a coryneform appearance were detected in two blood cultures per patient, processed with the VITAL system (bioMerieux, Marcy l'Etoile, France). These grew, after 24 h of incubation in an atmosphere enriched with 5% CO2, in the subcultures

degree of wound contamination, and it is estimated that 50% of wounds contaminated by bacteria become clinically infected (6). These bacteria can seriously delay wound healing process by disrupting the normal clotting mechanisms and promoting disordered leukocyte function and poor quality granulate on tissue formation, reduce tensile strength of connective tissue, and impair epithelization.

Olax subscorpioidea.oliv is a shrub or tree which belongs to the family of Olacaceae. It is widely distributed in West African countries such as Nigeria, Zaire and Senegal (7). The Shrub or small tree is between 9 m and 10 cm in diameter, and is fairly common in the semi-deciduous forests and gallery forests. The branches are flexible and angular with elliptex leaves; lanceolate in its entire margin. The Leaf apex has a characteristic point (mucronate); petiole 2- 4 mm; blade about 10 x 3 cm, with about 7 pairs of looped laterals. Olax subscorpioidea Oliv. fruit is small globular, about 1.5 cm in diameter, and yellowish but turning red when ripe. It has a slash smell like garlic. Two other Olax species are possibly present as shrubs or small trees: O. mannii (smooth twigs) and O. gambecola (pustulate twigs). Their leaves slightly resemble those of Celtis because of the strong, ascending basal nerves (tri-nerved).

Figure: 1.4: Olax subscorpioidea Oliv

Medicinal plants play a significant role in human health as they make an awesome substitute for orthodox medicine. Olax subscorpioidea Oliv. is one of such medicinal plants that is used in folk medicine mostly for giving relief for several ailments.. Olax subscorpioidea Oliv leaves is traditionally used in the treatment of diabetic mellitus. The leaves either form part of traditional recipe or is singly used in diabetic conditions. Research has shown that the plant leaf has potential to alleviate postprandial hyperglycaemia by inhibiting α-amylase and α-glucosidase (8). Olax subscorpioidea (Olacaceae) is a major component of a recipe used traditionally in the management of Obesity. This plant (especially the root) has shown to have hypolipidemic property and probably is capable of reducing weight in human (9). Olax subscorpioidea is a plants used in Ivorian traditional medicine in the treatment of many diseases including jaundice and hepatitis. The plant is used in mixture with palm wine locally called "bandji" for their traditional therapeutic utilization (10). Researchers have proved that Olax subscorpioidea Oliv. has antidepressant effect on effect and this justified its use in the treatment of mental disorders (11).

MATERIALS AND METHODS

Collection of Plant Material

Olax subscorpioidea Oliv. roots were collected across farms in Akungba community in Ondo State. The plant part was authenticated at the herbarium of the Department of plant science and biotechnology Adekunle Ajasin University, Akungba Akoko, Ondo State. Nigeria.

Preparation of Extract

Olax subscorpioidea Oliv. Root was washed with sterile water and air dried for 10days, and then chopped into small pieces to increase surface area. 100 g of the root was soaked in 400 mL of ethanol for 7 days. After which it was filtered with whatman No. 1 filter paper. The solvent was recovered and the crude extract obtained using rotary evaporator. The crude extract was thus kept in refrigerator at 4oC for further screening (12).

Specimen Collection

Specimens were collected hospitals across Akoko region of Ondo state, Nigeria using standard collection techniques (Cheesebrough, 2006). Sterile cotton-wool swabs were used to collect samples from the wound of surgical patients with evident infection. The swabs were introduced gently into the wound sites and rotating the swab tips in the wound, taking care to avoid contamination of specimen with commensals from the skin, and then immersed immediately in a MacCartney bottle containing peptone water. The sample bottles were then transported to the Microbiology laboratory of Adekunle Ajasin University, Akungba-Akoko, Ondo state(12).

Isolation of Bacteria

At the laboratory, the inoculated peptone water was serially diluted to 107 and the bacteria isolated using pour plate method.Aliquot of 1 ml from 104 and 107 were dispensed into sterile petri dishes, molten agar was then added, swirled and allowed to solidify. The media used were MacConkey agar, Nutrient agar, Eosin Methylene Blue agar and Mannitol salt agar and were each prepared according to the instruction of the manufacturer. The agar plates were then incubated at 37oC for 24 hours.

Isolation of Pure Isolates

After 24 hours of incubation, the colonies were subcultured by streaking each colony on the surface of freshly prepared agar plate. The plates were then incubated at 37oC for 24 hours. Distinct colonies were then picked into nutrient agar slant and stored in the refrigerator for further use.

Biochemical Test

Microbact (24E) kit identification

This kit was used for biochemical tests with the range of simple, standardized system, for the rapid identification of Gram- negative bacteria. Preparation of 18-24 hours old pure culture of the organism to be identified, oxidase test was performed which must be negative or positive for 24E kit, selected isolate colony was emulsified in saline, test strip was placed in holding tray and the back seal was peeled and addition of 4 drops of bacterial suspension to each well, addition of 2 drops of mineral oil (MB1093A) to the black wells, the seal was replaced and incubated at 350C + 20C for 18-24 hours, the tray was removed from incubator and appropriate reagents was added. The results was then recorded and interpreted using the MicrobactTM identification software package (Balows et al., 1991; Oxoid limited). A purity check was performed by inoculating a purity plate with 1 drop of bacterial suspension and incubated at 350C + 20C for 24 hours,Well 13 was read at 24-48hours for Enterobacteriaceae and at 48 hours for MGNB, Well 24 was interpreted differently at 24 hours and 48 hours, A nitrate reduction test was done in well 7 after reading the ONPG reaction, Performance was monitored by testing appropriate control strains (Oxoid Limited)(13).

Gram Stain

A loopful of sterile distilled water was dropped on a clean grease free slide by using a sterile inoculating loop after which an inoculum from the culture was mixed with the water on the slide. The smear was air dried and heat fixed by passing it quickly over a Bunsen flame. The smear was flooded with crystal violet solution for 60 seconds (one minute) and rinsed with water. The smear was again flooded with Lugol's iodine for 30 seconds and rinsed with water, 70% alcohol was poured on the slides for 15 seconds until the crystal violet had been completely washed off. It was then counterstained with Safranin for 60 seconds and allowed to dry. The slides were then observed under oil immersion objective lens (x100). Gram positive cells remained purple while Gram negative cells appeared red or pink (14).

Catalase Test

A drop of hydrogen peroxide solution was placed on a clean grease free slide. A flamed inoculating loop was used to place a loopful of an inoculum on the slide and gently mixed after which it was observed for bubbles or effervescence which is an indication of catalase positive organism (14).

Oxidase Test

A piece of filter paper was placed in petri dish and three (3) drops of freshly prepared oxidase reagent was added. Using a sterile glass rod, a colony of test organisms was removed from a culture plate and it was smeared on the filter paper. Oxidase positive organisms gave blue colour within 5 to 10 seconds, and in oxidase negative organisms, colour did not change (15).

Coagulase Test

A loopful of normal saline solution was placed on each glass slide and was emulsified. Human plasma was added to one of the suspension and was stored properly for 15 minutes while the other was left as control. Coagulase positive was indicated by clumping which did not re- emulsify (15).

Antibiotic Susceptibility Test and Antimicrobial Assay of the Extract

Antibiotic susceptibility tests were performed using Kirby-Bauer's disc diffusion method on Mueller-Hinton agar. The inocula were prepared from the stock cultures which were maintained in nutrient agar slant at 4^0C and subculture in nutrient broth using a sterilized wire loop. 1 ml of 10^{-4} normal saline dilution of a 24 hours broth culture was mixed with 19 ml of the agar in a sterile universal bottle and poured into sterile petri dish. The agar plate was left to solidify and the antibiotic disc were thus placed on the agar surface at minimum of 22 mm apart and were incubated overnight at 37°C for 24 hours.The diameter of zones of inhibition were measured and recorded in millimeter and the results interpreted according to the Clinical Laboratory Standard Institute (16) guidelines.

The antimicrobial screening ofthe Olaxsubscorpioidea Oliv. Root extract against the bacterial isolates was carried out using the agar well diffusion method. A stock concentration of 100 mg/ml was constituted by dissolving 1 g each, of the extracts in 10 ml of Dimethyl sulfoxide (DMSO) diluted with sterile distilled water in ratio 1:3. 50 mg/ml, 25 mg/ml and 12.5 mg/ml concentrations of the extracts were the prepared using dilution formula (C1V1=C2V2). 1 ml of10^{-4} normal saline dilution of a 24 hours broth culture was mixed with 19 ml of the agar in a sterile universal bottle and poured into sterile petri dish. The agar plate was left to solidify and wells were bored on them using 6 mm cork borer. 50 µl of each concentration of the extracts was poured into each well and incubated at 37°C for

24 hours.The diameter zones of inhibition were measured and recorded in millimeter and the results interpreted according to the Clinical Laboratory Standard Institute (16) guidelines. Piperacillin/tazobactam (0.125 mg/ml) and Demethyl sulfoxide (DMSO) were used as positive and negative controls respectively.

Minimal Inhibitory Concentration and Minimal Bacteriocidal Concentration

The minimal inhibitory concentration (MIC) was determined using the tube dilution method. Graded concentrations of the extract was prepared using Mueller Hinton broth medium into differenttest tubes. The concentrations were 100mg/ml, 50mg/ml, 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.125mg/ml. standardized inoculum of 24 hours broth culture was inoculated into the test tubes was incubated at 37oC for 24 hours. After incubation, the test tubes were examined for sign of growth (turbidity) and the minimal concentration with no growth was recorded as the MIC (17).

The minimal bacteriocidal concentration (MBC) was determined by streaking out samples from the test tubes with no growth on the surface of freshly prepared nutrient agar. The plates were then incubated at 37oC for 24 hours, after which plates were observed for any bacterial growth. Again, the minimal concentration with no growth was taken as the MBC(17).

Secondary Metabolite (Phytochemical) Screening of OlaxsubscorpioideaOliv. root

Qualitative Method of Analyses

Preliminary test / Preparation test

Plant filtrates were prepared by boiling 20 g of the fresh plant in distilled water. The solution was filtered through a vacuum pump. The filtrates were used for the phytochemical screening for flavonoids, tannins, saponins, alkaloids, reducing sugars, anthraquinones and anthocyanosides.

Test for Alkaloids

About 0.2gram was warmed with 2% of H2SO4 for two minutes, it was filtered and few drops of Dragendoff's reagent were added. Orange red precipitate indicates the present of Alkaloids(18).

Test for Tannins

One milliliter of the filtrate were mixed with 2m1 of FeC1, A dark green colour indicated a positive test for the tannins (18,19).

Test for Saponins

One milliliter of the plant filtrate were diluted with 2 ml of distilled water; the mixture were vigorously shaken and left to stand for 10min during which time, the development of foam on the surface of the mixture lasting for more than 10mm, indicates the presence of saponins (20).

Test for Anthraquinones

One milliliter of the plant filtrate was shaken with 10ml of benzene; the mixture was filtered and 5 ml of 10 % (v/v) ammonia were added, then shaken and observed. A pinkish solution indicates a positive test (20).

Test for Anthocyanosides

One milliliter of the plant filtrate was mixed with 5 m1 of dilute HCI; a pale pink colour indicates the positive test (21).

Test for Flavonoids

One milliliter of plant filtrate was mixed with 2 m1 of 10% lead acetate; a brownish precipitate indicated a positive test for the phenolic flavonoids. While for flavonoids, I m1 of the plant filtrate were mixed with 2m1 of dilute NaOH; a golden yellow colour indicated the presence of flavonoids(21,22).

Test for Reducing Sugars

One milliliter of the plant filtrate was mixed with Fehling A and Fehling B separately; a brown colour with Fehling B and a green colour with Fehling A indicate the presence of reducing sugars (23).

Test for Cyanogenic glucosides

This was carried out subjecting 0.5g of the extract 10ml sterile water filtering and adding sodium picrate to the filtrate and heated to boil (24).

Test for Cardiac glucosides

Legal test and the killer-kiliani was adopted, 0.5g of the extract were added to 2ml of acetic anhydrate plus H2S04(25)

Quantitative Method of Analyses

Saponins

About 20grams each of dried plant samples were ground and, put into a conical flask after which 100 ml of 20 % aqueous ethanol were added. The mixture were heated using a hot water bath. At about 55OC, for 4 hour with continuous stirring, after which the mixture were filtered and the residue re-extracted with a further 200 ml of 20% ethanol. The combined extracts were reduced to 40 ml over a water bath at about 90°C. The concentrate was transferred into a 250 ml separatory funnel and 20 rnl of diethyl ether were added and then shaken vigorously. The aqueous layer were recovered while the ether layer was discarded. The purification process was repeated three times. 60 rnl of n-butanol were added. The combined n-butanol extracts were washed twice with 10 m1 of 5% aqueous sodium chloride. The remaining solution was heated in a water bath. After evaporation, the samples were dried in the oven to a constant weight; the saponin content was calculated as percentage of the starting material (25,26).

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Flavonoids

About 10 g of the plant sample were extracted repeatedly with 100 ml of 80% aqueous methanol, at room temperature. The whole solutions were filtered through Whatman filter paper No 42. The filtrate were later transferred into a crucible and evaporated into dryness over a water bath; the dry content was weighed to a constant weigh (27).

Cardiac glucosides

Legal test and the killer-kiliani wwas adopted, 0.5g of the extract were added to 2ml of acetic anhydrate plus H2S04(28).

Tannins

About 500 mg of the plant sample were weighed into a 50 ml plastic bottle. 50 ml of distilled water was added and shaken for 1 hour on a mechanical shaker. This was filtered into a 50 ml volumetric flask and made up to the marked level. Then, 5 ml of the filtrate was transferred into a test tube and mixed with 2 ml of 0.1 M Fecl in 0.1 M Hcl and 0.008 M potassium ferrocyanide. The absorbance was measured at 120 nm within 10 minutes. The tannins content was calculated using a standard curve of extract (29).

Alkaloids

Five grams of the plant sample were weighed into a 250 ml beaker and 200ml of 10% acetic acid in ethanol was then be added, the reaction mixture were covered and allowed to stand for 4 hour. These were filtered and the extract will be concentrated on a water bath to one-quarter of the original volume. Concentrated ammonium hydroxide was added drop-wise to the extract until the precipitation is complete. The whole solution were allowed to settle and the precipitate was collected, washed with dilute ammonium hydroxide and then filtered; the residue being the alkaloid, which was dried and weighed to a constant mass (30,31,32).

Phlobatannins

About 0.5grams of each plant extracts were dissolved in distilled water and filtered. The filtrates were boiled in 2% Hcl, red precipitate show the present of phlobatannins (31).

RESULTS

Table 3.1 shows the result of preliminary tests carried out to characterize the twenty six (26) bacterial isolates recovered, six (23.1%) were Gram positive and twenty (76.9%) were Gram negative. Also, from a single surgical site more than two bacteria were isolated. Twenty two (22) of the isolates were rod shaped, three (3) are tiny rods and 0ne (1) is cocci shaped. They are predominantly Gram negative rods except six (6) which are Gram positive. Eighteen (18) of the isolates are oxidase positive while eight (8) of the isolates are oxidase negative. Nineteen (19) of the isolates are catalase positive while seven (7) of the isolates are catalase negative.

Table 3.2 shows the identity of the eight (8) Gram negative bacterial isolates as revealed by Microbact 24E identification kit. Fifty percent 50% were oxidase-negative with Escherichia coli most prevalent (25%). Other organisms include Klebsiella pneumoniae, Enterobacter agglomerans, Pseudomonas aeruginosa, Pseudomonas fluorescens, Burkholderia pseudomallei and Mannheimia haemolytical. Most of the SSI-associated bacteria were comprised of Staphylococcus aureus and Pseudomonas aeruginosa.

Table 3.1: Gram stain, microscopic examination and preliminary biochemical tests carried on the recovered isolates

ISOLATE	GRAM STAIN	SHAPE	OXIDASE	CATALASE
MAC 1a	+	Rods	+	-
MAC 1b	-	Tiny rods	+	+
MAC 2	-	Tiny rods	+	+
MAC 4	+	Cocci	+	+
MAC 6	-	Rods	-	-
MAC 8	-	Tiny rods	-	-
MAC 9	-	Rods	+	+
MAC 12	+	Rods	+	+
MAC 14	-	Rods	+	+
MAC 16	-	Rods	+	+
MAC 17	-	Rods	+	+
MAC 18	-	Rods	+	+
NA 1	-	Rods	+	+
NA 4	-	Rods	-	+
NA 5	-	Rods	-	-
NA 10	-	Rods	+	+
NA 13	-	Rods	-	+
NA 14	-	Rods	+	+
NA 18	-	Rods	+	+
EMB 2	-	Rods	+	-
EMB 13	-	Rods	+	-
EMB 11	-	Rods	+	+
EMB 18	-	Rods	+	+
MAN 5	+	Rods	-	+
MAN 10	+	Rods	-	-
MAN 18	+	Rods	-	+

Table 3.2: Identification of the Gram negative bacteria using Microbact 24E identification kit

ISOLATE	IDENTIFICATION CODE	PROBABLE ORGANISM
MAC 12	507041776	Mannheimia haemolytica
EMB 2	46610776	E. coli
MAC 18	707145776	Burkholderia pseudomallei
EMB 11	46710776	E. coli
NA 10	703134001	Pseudomonas aeruginosa
NA 13	43471776	Klebsiella pneumonia
MAC 14b	703134100	Pseudomonas fluorescens
MAC 6	07061776	Enterobacter agglomerans

Table 3.3 reveals the antibiotic susceptibility profile of the bacterial isolates. Ciprofloxacin (10µg) was active against 50% of the isolates with diameter of zones of inhibition ranging from 15mm to 30 mm. pefloxacin, ofloxacin and streptomycin were also active against 3 isolates with lesser diameter of zones of inhibition (11mm-17mm). The most susceptible isolates were Pseudomonas aeruginosa, Enterobacter agglomerans and Pseudomonas fluorescens. About 70% of the isolates were resistant to all the antibiotics tested except ciprofloxacin. Ciprofloxacin was effective against 50% of the isolates. Ciprofloxacin was the most effective antibiotic against Gram negative bacteria with exception to Staphylococcusaureus. High sensitivity was observed among Listeria monocytogens, Pseudomonas aeruginosa, Enterobacter agglomerans and Pseudomonas fluorescens. However, placid effects of septrin were observed against Pseudomonas aeruginosa, Enterobacter agglomerans and Pseudomonas fluorescens.

Table 3.4 shows the antimicrobial activity of the Olax subscorpioidea Oliv. root extract against the isolates. The extract inhibited the growth of all the bacterial isolates with diameter of zones of inhibition ranging from 6mm to 25mm. At 100mg/ml the diameter of zones of inhibition ranges from 6mm to 9mm, at 50mg/ml the diameter of zones of inhibition ranges from 6mm to 10mm, at 25mg/ml the diameter of zones of inhibition ranges from 7mm to 9mm, at 12.5mg/ml the diameter of zones of inhibition ranges from 7mm to 9mm while the control (Tazobactam antibiotics) ranges from 7mm to 25mm. This table is well explained in Fig. 3.1 to 3.4 below in which the zones of inhibition of the organisms were represented by bar chart.

ORGANISM	SXT (30 µg)	CH (30 µg)	SP (10 µg)	CPX (10 µg)	AM (30 µg)	AU (30 µg)	CN (10 µg)	PEF (10 µg)	OFX (30 µg)	S (30 µg)
E. coli	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Mannheimia haemolytical	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
E. coli	0.0	0.0	0.0	30.0	0.0	0.0	0.0	0.0	0.0	0.0
Listeria monocytogens	0.0	0.0	0.0	25.0	0.0	0.0	0.0	0.0	0.0	0.0
Pseudomonas aeruginosa	0.0	0.0	0.0	15.0	0.0	0.0	0.0	15.0	12.0	12.0
Enterobacter agglomerans	0.0	0.0	0.0	15.0	0.0	0.0	11.0	12.0	12.0	12.0
Pseudomonas fluorescens	13.0	12.0	0.0	15.0	0.0	0.0	9.0	11.0	14.0	17.0
Klebsiella pneumonia	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Burkholderia pseudomallei	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Staphylococcus aureus	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0

KEY: SXT- Septrin, CH- Chloramphenicol, SP- Sparfloxacin, CPX- Ciprofloxacin, AM- Amoxicillin, AU- Augmentin,
CN- Gentamycin, PEF- Pefloxacin, OFX- Ofloxacin and S- Streptomycin

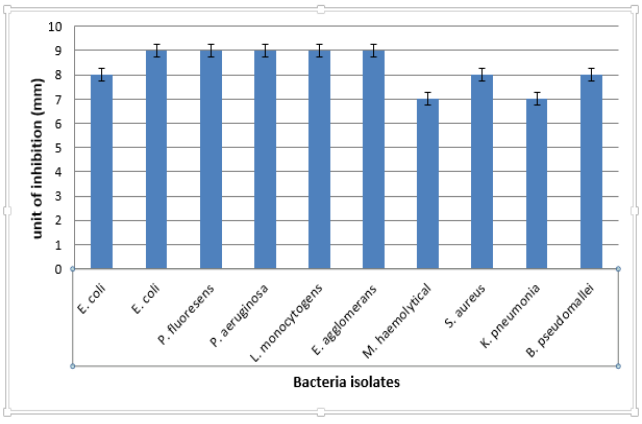

Fig 3.1:Antimicrobial screening of ethanol extract of Olax subscopioidea Oliv. Root at 100mg/ml.

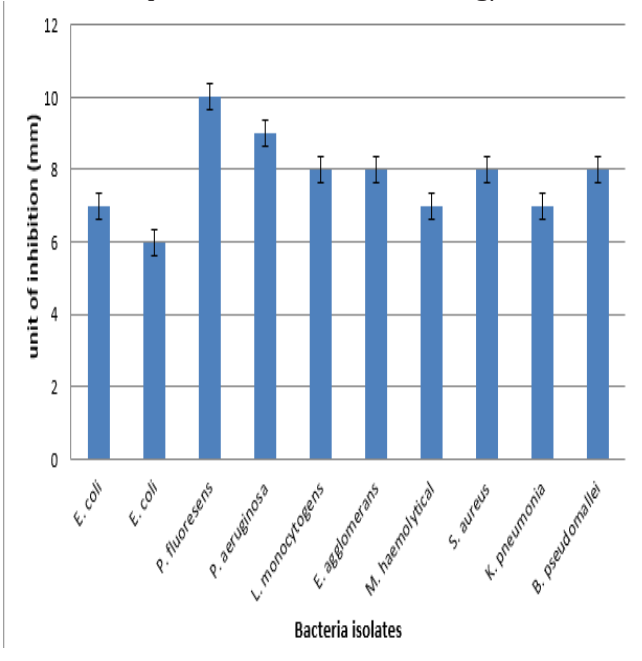

Fig 3.2:Antimicrobial screening of ethanol extract of Olax subscopioidea at 50mg/ml.

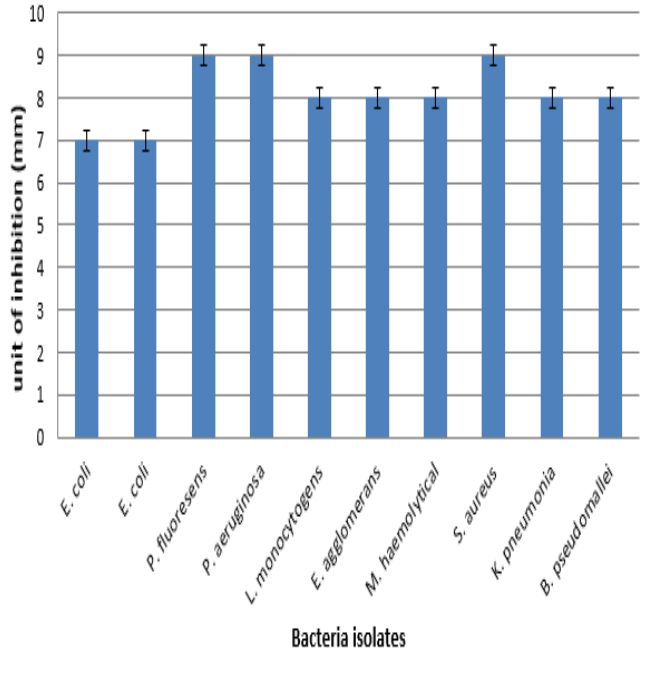

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Fig 3.3: Antimicrobial screening of ethanol extract of Olax subscopioidea Oliv. rootat 25mg/ml.

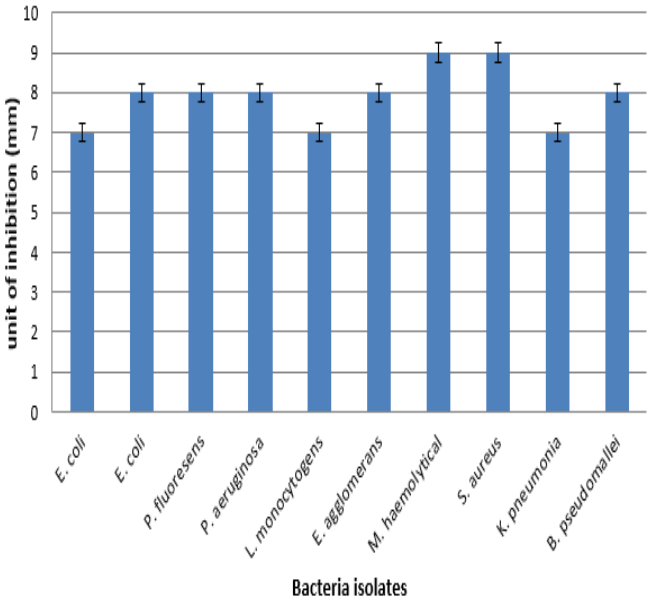

Fig 3.4:Antimicrobial screening of ethanol extract of Olax subscopioidea Oliv. rootat 12.5mg/ml.

Table 3.5 shows the minimal inhibitory concentration (MIC) and minimal bacteriocidal concentration (MBC) of the extract. MIC ranges from 25mg/ml to 100mg/ml while MBC was between 50mg/ml and 200mg/ml. This table is well explained in Fig. 3.5 to 3.6 below in which the zones of inhibition of the organisms were represented by bar chart.

Table 3.6 shows the qualitative phytochemical screening of the ethanol extract of Olax subscorpioidea Oliv. root. Flavonoid, tannins, saponins, alkaloids, reducing sugars, steroid, phenol, terpenoid, pyrrolozidine alkaloid, glycoside and cardiac glycoside were present while anthraquinones and volatile oil were not detected.

Table 3.7 present the quantitative phytochemical screening of the ethanol extract of Olax subscorpioidea Oliv. root. Glycoside and terpenoid were most present with a concentration of 14.01 while saponins were least present with a concentration of 3.21.

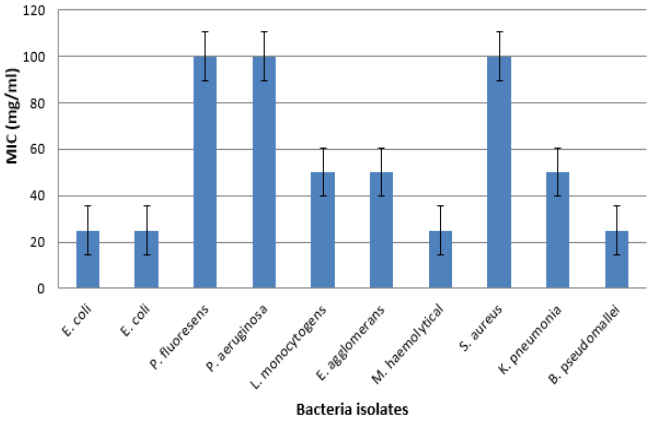

Fig 3. 5: Minimal inhibitory concentration MIC (mg/ml)

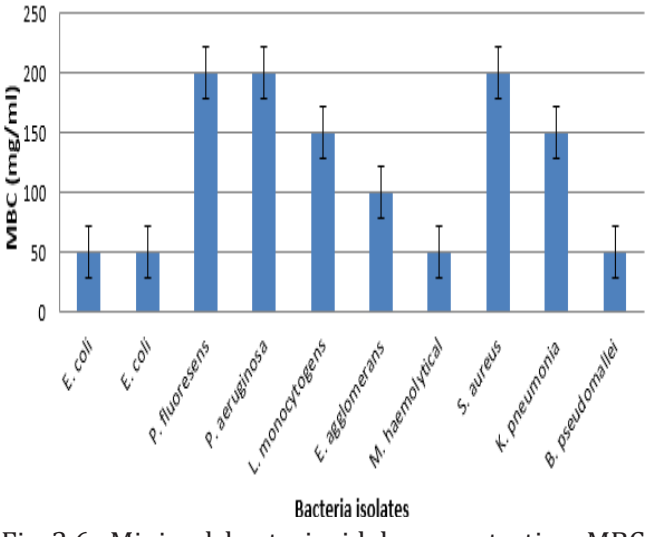

Fig 3.6: Minimal bacteriocidal concentration MBC (mg/ml)

Table 3.6: Qualitative phytochemical (secondary metabolite) screening of ethanol extracts of Olax subscorpioidea Oliv. root

Constituent	Presence
Alkaloids	+
Glycoside	+
Steroids	+
Anthraquinone	ND
Phenol	+
Tannins	+
Saponin	+
Flavonoids	+
Pyrrolizidine alkaloids	+
Reducing sugar	+
Terpenoid	+
Volatile oil	ND
Cardiac glycosides	+

KEY: + = Present, - = Absent and ND = Not Detected

Table 3. 7: Quantitative phytochemical (secondary metabolite) screening of ethanol extracts of Olax subscorpioidea Oliv. root

Constituent	Quantity
Alkaloids	13.89
Glycoside	14.01
Steroids	9.71
Anthraquinone	9.75
Phenol	7.51
Tannins	7.60
Saponin	3.21
Flavonoids	4.21
Pyrrolizidine alkaloids	4.00
Reducing sugar	13.89
Terpenoid	14.01
Volatile oil	9.71
Cardiac glycosides	9.75

REFERENCES

1. Early Alzheimer's Disease: Patterns of Functional MRI Activation-The Neural Substrates of Semantic Memory Deficits. Am J Bra Dis and Tum. 2018; 1(1): 001-010.
2. H Chahal, S W D Souza, A J Barson and P Slater. How to develop human brain using magnesium of N-methyl-D-aspartate receptors, Am J Bra Dis and Tum. 2018; 1(1): 001-005.
3. F S LaBella, et al.Concepts and correlations related to general anaesthesia and cytochrome P450 oxygenases. Am J Anest and Pai med. 2018; 1(1): 01-05.
4. Hazim J Safi, et al. The long term method with the elephant trunk for the repair of aortic aneurysms. Am J Anest and Pai med. 2018; 1(1): 001-008.
5. Yoshitaka Fujii, et al. Diaphragmatic Fatigue is treated with Inhaled Aminophylline Therapy in an Experimental Canine procedure. Am J Anest and Pai med. 2018; 1(1): 001-003.
6. O Demirkiran, et al. Complications in patients with Crush syndrome after the Marmara earthquake. Am J Anest and Pai med. 2018; 1(1): 001-005.
7. Qi Wei, et al. Laparoscopic choledochotomy after Biliary drainage: Study. Am J Anest and Pai med. 2018; 1(1): 001-007.
8. Mark Palazzo, et al. Unilateral Babinski/Plantar Reflex - Acute Inflammatory Demyelinating Polyneuropathy. Am J Anest and Pai med. 2018; 2(1): 01-02.
9. Hakan Alfredson, et al. Achilles and patellar tendon operations performed in local anestesia, Am J Anest and Pai med. 2018; 1(1): 001-002.
10. Naemeh Nikvarz, et al. Evaluation The Analgesic Effect of Duloxtine Drug in Burn Pationts. Am J Anest and Pai med. 2019; 2(1): 01-07.
11. Chuandong Zheng, et al. Intravascular Plaque: Cause for Radial Arterial Catheterization Failure. Am J Anest and Pai med. 2019; 2(1): 01-05.
12. Laura Tyler Perryman, et al. Wireless Dorsal Root Ganglion Stimulation: An Introduction and Early Experience with the New Approach for Chronic Pain Management. Am J Anest and Pai med. 2019; 2(1): 01-04.
13. Lazraq Mohamed, et al. Pediatric Pre-Anesthesia Consultation: What are Parents Expectations?. Am J Anest and Pai med. 2019; 2(1): 01-02.
14. Alaa Ali M. Elzohry, et al. Safety and Efficacy of Intraperitoneal Irrigation of Levo-Bupivacaine plus Morphine in Patients Undergoing Major Abdominal Cancer Surgeries. Am J Anest and Pai med. 2019; 2(1): 01- 07.
15. Yıldız K, et al.Comparison between Anesthesia Methods In Orthopaedics Initiatives of Upper Extremity. Am J Anest and Pai med. 2019; 2(2): 01-03.
16. Jianming Liu, et al. The Analgesic Effects Nalbuphine Hydrochloride Combined With Sufentanil for Patients after Thoracoscopic Lobectomy. Am J Anest and Pai med. 2019; 2(2): 01-03.
17. Fudong Shi, et al. The Patient Controlled Intravenous Analgesia of Dezocine on the Elderly Patients After Orthopedic Surgery. Am J Anest and Pai med. 2019; 2(1): 01-04.
18. GE Meglia, et al. Investigation in blood Leukocytes and Neutrophils in Periparturient Dairy Cow. Sci J of Ani and Vet Sci. 2018; 1(1): 001-009.
19. G E Duhamel, et al.DNA Sequence Analysis of an Immunogenic Glucose-Galactose Mglb. Sci J of Ani and Vet Sci. 2018; 1(1): 001-009.
20. David G. White, et al. Chloramphenicol and Florfenicol Resistance in Escherichia Coli of Characterization . Sci J of Ani and Vet Sci. 2018; 1(1): 001-006.
21. N B Alhaji, et al. Anophthalmia and Choanal Atresia In Two Months Old Kid. Sci J of Ani and Vet Sci. 2018; 1(1): 001-004.
22. Christopher W Olsen, et al.Isolation and Characterization of H4N6 Avian and Influenza Viruses. Sci J of Ani and Vet Sci. 2018; 1(1): 001-0025.
23. Teresa Lopez-Arteaga, et al. Apathy as a Psychiatric Manifestation of Meningioma. Am J Bra Dis and Tum. 2018; 1(1): 001-004.
24. David R Murdoch, et al. The Use of Brain Natriuretic Peptide- Whole Blood can be Measured, Am J Bra Dis and Tum. 2018; 1(1): 001-003.
25. Stefan Brocke, et al. Antibodies to Integrin α4 and CD44, but not CD62L, Prevent CNS Inflammation and Experimental Encephalomyelitis by Blocking Secondary Leukocyte Recruitment. Am J Bra Dis and Tum. 2018; 1(1): 001-006.
26. Andrew J Saykin, et al. Early Alzheimer's Disease: Patterns of Functional MRI Activation-The Neural Substrates of Semantic Memory Deficits. Am J Bra Dis and Tum. 2018; 1(1): 001-010.
27. P Slater, et al.How to develop human brain using magnesium of N-methyl-D-aspartate receptors, Am J Bra Dis and Tum. 2018; 1(1): 001-005.
28. Clyde W Hodge, et al. The Paraventricular Nucleus Interactively Modulate Ethanol Consumption -Norepinephrine and Serotonin Receptors, Am J Bra Dis and Tum. 2018; 1(1): 001-005.
29. Paulo C Carvalho, et al. Bioinformatics grid application in simple - Squid. Sci J Biome and Biost. 2018; 1(1): 001-004.
30. Mahmoud A E Abdelrahman, et al. On The New Exact Solutions for the Nonlinear Models Arising In Plasma Physics. Sci J Biome and Biost. 2018; 1(1): 001-004.
31. Weicheng Shen, et al. Based on Personal Identification- Automated Biometrics. Sci J Biome and Biost. 2018; 1(1): 001-002.
32. V Prasathkumar, et al.Fingerprint Biometric System - Using of Personal Authentication. Sci J Biome and Biost. 2018; 1(1): 001-003.
33. SavitaChoudhary, et al. Software Development Environment : Design of Biometric Based Transaction System. Sci J Biome and Biost . 2018; 1(1): 001-003.
34. D J Lawrence, et al. Measuring the effectiveness in reliability and validity of a visual function outcomes instrument in cataract surgery. Sci J Biome and Biost. 2018; 1(1): 001-004.
35. Z Suvakovic, et al. Evaluation of early detection of gastric cancer requries more than gastroscopy. Anna of Can Ther and Phar. 2018; 1(1): 05.
36. Ho GY, et al. Informing and involving personalised computer based data for cancer patients. Anna of Can Ther and Phar. 2018; 1(1): 001-005.
37. Ray Jones, et al. Prostate Cancer Risk is associated with Polymorphism of Insulin gene. Anna of Can Ther and Phar. 2018; 1(1): 001-005.
38. : Jean-Pierre J. Issa, et al. Role of DNA Methylation in Tumor Suppressor Gene Silencing in Colorectal Cancer. Anna of Can Ther and Phar. 2018; 1(1): 001-008.
39. Jules J Berman, et al. Histological classification of tumour and molecular analysis meets Aristotle. Anna of Can Ther and Phar. 2018; 1(1): 001-005.
40. Kafil Akhtar, et al. Tuberculosis of the Tongue with Coexistent Squamous Cell Carcinoma: An Interesting Case Presentation, Anna of Can Ther and Phar. 2018; 1(1): 001-002.
41. Serafin Morales Murillo, et al. Vitamin D as A Prognostic Factor in Triple Negative Breast Cancer. Anna of Can Ther and Phar. 2019; 2(1): 01-08.
42. Ahmet Fuat, et al. A Qualitative Study of Accurate Diagnosis and Effective Management of Heart Failure in Primary Care. Am J of Card and Cardiovas Disc. 2018; 1(1): 01-05.

Cite this article: A Usually Daptomycin-Resistant Gram-Positive Dermabacter Hominis: Rare Pathogen. Am J Micro and Bioche. 2019; 1(1): 001-011.

43. Jesús Millán Núñez-Cortés, et al. Prescription Habits for Statins in Patients with Impaired Glucose Metabolism. Results of a program with Focus Groups to Assess the Selection Criteria. Am J of Card and Cardiovas Disc. 2019; 1(1): 01-04.
44. G D Kolovou, et al. Evaluation of Postprandial hypertriglyceridaemia in patients withTangier disease. Am J of Card and Cardiovas Disc. 2018; 1(1): 01-04.
45. Brian O rourke, et al. Determination of The Mitochondrial Redox Waves and Subcellular Metabolic Transients in Heart Cells. Am J of Card and Cardiovas Disc. 2018; 1(1): 01-04.
46. Shuixiang Yang, et al. Radiofrequency Ablation Treating Atrial Fibrillation Can Reverse the Changes of Mirnas Regulating Ion Channel Proteins. Am J of Card and Cardiovas Disc. 2018; 1(1): 01-08.
47. Hadi abdulsalam Abo Aljadayel, et al. Penetrating War Cardiac and Great Vessels Injury, Surgical Outcome Analysis in 24 Patients. Am J of Card and Cardiovas Disc. 2018; 1(2): 01-05.
48. Hatice Yorulmaz, et al. Assessment of the Death Anxiety and Death Depression Levels of Cardiac Patients. Am J of Card and Cardiovas Disc. 2019; 2(1): 01-06.
49. Camara Abdoulaye, et al. Cardiomyopathie Du Peripartum Compliquee D'accident Vasculaire Cerebral Cas D'une Guinéenne De 19ans : Cas Clinique. Am J of Card and Cardiovas Disc. 2019; (1): 01-03.
50. Sergio F. Estrada-Orihuela, et al. Lasalocid, Interrupts and Reverses, Within One Minute, The Myocardial Damage Caused By Coronary Anoxia Reperfusion in Rat Heart. Am J of Card and Cardiovas Disc. 2019; (1): 01-05.
51. Jesus Millan Nunez-Cortes, et al. Prescription Habits for Statins in Patients with Impaired Glucose Metabolism. Results of a program with Focus Groups to Assess the Selection Criteria. Am J of Card and Cardiovas Disc. 2019; 1(1): 01-06.
52. Federico Cacciapuoti, et al. The Dilemma of Diastolic Heart Failure. Am J of Card and Cardiovas Disc. 2019; 1(1): 01-03.
53. Elad Boaz, Bowel Ischemia and Vascular Air-Fluid Levels. Anna Cas Rep and Ima Surg. 2018; 1(1): 001-00.
54. Sinisa Franjic, et al. A Patient With A Maxillofacial Problem. Anna Cas Rep and Ima Surg. 2018; 1(1): 001-004.
55. Davidson W, et al. Case Presentation: Hantavirus pulmonary syndrome [HPS]. Anna Cas Rep and Ima Surg. 2018; 1(1): 001-005.
56. Farid ZM, et al. Uropathy Secondary Chronic obstructive to Ureter Inguinal Herniation. Anna Cas Rep and Ima Surg. 2018; 1(1): 001-002.
57. De Letter DJ, et al. Cornual Molar Ectopic Pregnancy Diagnosis and Treatment. Anna Cas Rep and Ima Surg. 2018; 1(1): 001-003.
58. Ameni Touati, et al. Silver Russell Syndrome: Case Reports from North Africa and Review on The Literature. Anna Cas Rep and Ima Surg. 2019; 1(1): 001- 004.
59. Kunst WM, et al. Case Reports and Review of Spontaneous Rupture of Hyperreactive Malarial Splenomegaly [HMS]. Anna Cas Rep and Ima Surg. 2018; 1(1): 001-005.
60. F Hanefeld, et al. A Review of The Literature an Emerging Community Pathogen methicillin-Resistant Staphylococcus. Anna Cas Rep and Ima Surg. 2018; 1(1): 001-0011.
61. Page W Caufield, et al. Evidence for a Discrete Window of Infectivity. Am J Den and Ora Car. 2018; 1(1): 001-006.
62. Robert T Dirksen, et al. Dihydropyridine Receptors and Ryanodine Receptors: Bi-Directional Coupling . Am J Den and Ora Car. 2018; 1(1): 001-009.
63. IJ Jacobs, et al. Cancer and Intraepithelial Neoplasia-Tissue-specific apoptotic effects of the p53 codon 72 polymorphism . Am J Den and Ora Car. 2018; 1(1): 001-003.
64. Iain L C Chapple, et al. Human Immunodeficiency Virus disease in oral health significances. Am J Den and Ora Car. 2018; 1(1): 001-007.
65. H Larjava, et al. Activity of αvβ6 Integrin in Oral Leukoplakia. Am J Den and Ora Car. 2018; 1(1): 001-005.
66. Siddharth Kothari, et al. Effectiveness of Counselling and Home Care Self-Management Strategies in Reducing Masticatory Muscle Pain: A Review. Am J Den and Ora Car. 2019; 2(1): 001-007.
67. Betania Maria Soares, et al. Use of Blue LED and Curcumin for Photosensitization of Candida Albicans. Am J Den and Ora Car. 2019; 2(1): 001-005.
68. Jing Guo, et al. Advances in Methods of Maxillary Transverse Expansion. Am J Den and Ora Car. 2019; 2(1): 01-05.
69. Dario C. Altieri, et al. Cell division by p34cdc2 phosphorylation of survivin- Regulation . Sci J of Der and Ven. 2018; 1(1): 001-005.
70. Axel Trautmann, et al. Eczematous dermatitis: T cell and keratinocyte apoptosis plays a key pathogenetic . Sci J of Der and Ven. 2018; 1(1): 001-007.
71. JD Fine, et al. Epidermolysis bullosa Cardiomyopathy in inherited . Sci J of Der and Ven. 2018; 1(1): 001-004.
72. NE Fusenig, et al. Human Skin Angiogenic Switch Occurs Squamous Cell Carcinomas . Sci J of Der and Ven. 2018; 1(1): 001-007.
73. Tapani Tuomi, et al. Water- Damaged Building and Mycotoxins in Crude Building Materials. Sci J of Der and Ven. 2018; 1(1): 001-005.
74. John S Davies, et al. The Use of Social Media among Doctors Under taking a Post-Graduate Endocrinology Diploma. Sci J Endo and Meta. 2018; 1(1): 001-004.
75. Juan J Gagliardino, et al. By Short-Term Dietary Manipulation: The Endocrine Pancreas Activity of Tyrosine Hydroxylase. Sci J Endo and Meta. 2018; 1(1): 001-005.
76. Colin A. Leech, et al. The Glucose Dependent in Pancreatic β-Cells : Voltage-Independent Calcium Channels Mediate Slow Oscillations of Cytosolic CalciumPancreatic β-Cells. Sci J Endo and Meta. 2018; 1(1): 001-009.
77. Colin A. Leech, et al. The Voltage-Independent Activation of Inward Membrane Currents and Elevation of Intracellular Calcium in HIT-T15 Insulinoma CellsPituitary Adenylate Cyclase-Activating Polypeptide Induces. Sci J Endo and Meta. 2018; 1(1): 001-008.
78. Suhail AR Doi, et al. Making Use Of Combined Criteria - Diagnostic Criteria For Diabetes. Sci J Endo and Meta. 2018; 1(1): 001-006.
79. Maria I Borelli, et al. Effect Of Endogenous Islet Catecholamines Possible Modulatory On Insulin Secretion. Sci J Endo and Meta. 2018; 1(1): 001-005.
80. Louis Irwin, et al. Effect of exercise in combination with dietary nopal and zucchini on chronic and acute glucohomeostasis in genetically obese mice. Inte J Expe Bio. 2018; 1(1): 001-005.
81. Vijaya Saradhi Settaluri, et al. Validation of Non Essential Amino Acids and Total Protein Content in Different Categories of Tea. Inte J Expe Bio. 2018; 1(1): 01-04.
82. Patrick D Craig, et al. T Antigen: Polyomavirus Middle of Natural Biology. Inte J Expe Bio. 2018; 1(1): 001-007.
83. Yoshinori Ohsumi, et al. The HIV Coreceptor CCR5 - Recycling and Endocytosis. Inte J Expe Bio. 2018; 1(1): 001-008.
84. Marino Zerial, et al. Elicitation of the Angiogenic Phenotype1: Transforming Myc Protein for In Vivo. Inte J Expe Bio. 2018; 1(1): 001-008.
85. Zhang Y, et al. Odorant Receptor In Mammali : The Caenorhabditis Elegans Seven-Transmembrane Protein ODR-10 Functions on Cells. Inte J Expe Bio. 2019; 1(1): 001-008.
86. Kazuo Maeda, et al. Improved Outcome with Novel Studies in Fetal Monitoring. Sci J of Gyne and Obste. 2019; 2(1): 001-004.

Cite this article: A Usually Daptomycin-Resistant Gram-Positive Dermabacter Hominis: Rare Pathogen. Am J Micro and Bioche. 2019; 1(1): 001-011.

87. Sunil J. Wimalawansa, et al. Vitamin D Deficiency-Related Reproductive Consequences. Sci J of Gyne and Obste. 2019; 2(1): 001-006.
88. Munch A, et al.Investigation in blood Leukocytes and Neutrophils in Periparturient Dairy Cow. Sci J of Gas and Hepa. 2018; 1(1): 001-006.
89. Jie Song Hua, et al. Primary Helicobacter Pylori Resistance to Clarithromycin and Metronidazole in Singapore. Sci J of Gas and Hepa. 2018; 1(1): 001-003.
90. Paul Moayyedi, et al. A Systematic Review and Economic Analysis: Proton Pump Inhibitors in Nonulcer Dyspepsia Efficacy. Sci J of Gas and Hepa. 2018; 1(1): 001-003.
91. Zhen-Ning Wang, et al.Gastric Cancer: Collagen IV Expression and Biological Behavior. Sci J of Gas and Hepa. 2018; 1(1): 001-002.
92. Zhen-Ning Wang, et al. A Possible Pathophysiologic Contribution to Necrotizing Enterocolitis: Human Intestine Inflammation. Sci J of Gas and Hepa. 2018; 1(1): 001-006.
93. Paul M Wassarman, et al. Egg Interaction during Mammalian Fertilization in the Molecular Basis of Sperm . Sci J of Gyne and Obste 2018; 1(1): 001-006.
94. Mary Lou Moore, et al. Breastfeeding Benefits Support -Research. Sci J of Gyne and Obste 2018; 1(1): 001-002.
95. Pepita Gimenez-Bonafe, et al. Preservation of Fertility in Patients with Cancer. Sci J of Gyne and Obste 2018; 1(2): 001-006.
96. Yueyang F Fei, et al. Non-Hemorrhagic Unilateral Adrenal Infarct In Pregnancy: A Case Report. Sci J of Gyne and Obste. 2019; 1(1): 001-002.
97. Karen Pierre, et al. Protein-Energy Adequacy of Dialysis Patients in Trinidad and Tobago. Am J of Nep and Ther. 2018; 1(1): 01-05.
98. Balakrishna N, Tenckhoff Catheter Surgical under Local Anesthesia. Am J of Nep and Ther. 2018; 1(1): 001-003.
99. J T Ohlsson, et al. Man in angiotensin and noradrenaline inhibits the Endothelin. Am J of Nep and Ther. 2018; 1(1): 001-005.
100. David J, et al. Apoptosis and Ischemic Renal Injury Reduce the Guanosine Supplementation. Am J of Nep and Ther. 2018; 1(1): 001-005.
101. R W Baldeweg, et al. Tumor-induced osteomalacia : Cloning and characterization of Fibroblast Growth Factor 23. Am J of Nep and Ther. 2018; 1(1): 001-006.
102. Amitabh Arya, et al. Post Pyeloplasty Follow Up In Children And Adolescents: Diuretic Renography Or Renal Ultrasonography? Am J of Nep and Ther. 2019; 2(1): 001-005.
103. Amitabh Arya, et al. Post Pyeloplasty Follow Up In Children And Adolescents: Diuretic Renography Or Renal Ultrasonography? Am J of Nep and Ther. 2019; 2(1): 001-005.
104. Richard Lechtenberg, et al. Tau Interferon in Multiple Sclerosis. Amer J Neur & Neurophysi. 2018; 1(1): 001-002.
105. Eva Guy Rodriguez, et al. Discussion of the differential diagnosis of bilateral thalamic lesions-Bilateral thalamic infarcts due to occlusion of the Artery of Percheron. Amer J Neur & Neurophysi. 2018; 1(1): 001-004.
106. Yhashi Chang, et al. IVIg for Miller Fisher syndrome: Cerebral infarction. Amer J Neur & Neurophysi. 2018; 1(1): 001-002.
107. Fredrick J. Seil, et al. T cell responses to Myelin Antigens and Antimyelin Antibodies. Amer J Neur & Neurophysi. 2018; 1(1): 001-005.
108. Y Niimi, et al. Embolization of Spinal Cord AVMs: Neurophysiologic Provocative Testing. Amer J Neur & Neurophysi. 2018; 1(1): 001-002.
109. Ameni Touati, et al. Some Reducibility Results for Differentiable Sets. Amer J Neur & Neurophysi. 2019; 1(1): 001-005.
110. Chrisostomos Sofoudis, et al. Sofoudis C. Septic Abortion Accompanied with Dessiminated Intravascular Coagulation and Acute Cardiomyopathy Presentation of a Rare Case and Mini Review. Am J Nur & Pract. 2018;1(1): 001-00.
111. Nick Jones, et al. Nurse Practitioners and Family Physicians Ethics Health Care Services. Am J Nur & Pract. 2018; 1(1): 001-005.
112. Thomas R A, et al. Human Infants Learning by Prenatal and Postnatal Flavor . Am J Nur & Pract. 2018; 1(1): 001- 006.
113. D. J Wise, et al. A Randomized, Double-Blind, Placebo-Controlled - Milk Production in Mothers of PremaTure Newborns Domperidone Drug effect. Am J Nur & Pract. 2018; 1(1): 001-005.
114. Bronagh Bufton, et al. Effects of Nursing Homes Ownership Compromise the Quality of Care. Am J Nur & Pract. 2018; 1(1): 001-005.
115. Kerstin Ekberg, et al. How Physicians Deal With the Task of Sickness Certification in Cause-Based and Comprehensive Disability Systems – A Scoping Review. Am J Nur & Pract. 2019; 2(1): 01-10.
116. Michael J. Vives, et al. Factors in Choosing the Surgical Approach: Cervical Spondylotic Myelopathy. Am J Orth and Rhe. 2018; 1(1): 001-004.
117. M. Runge, et al. Geriatric Patients in Balance Training and Exercise. Am J Orth and Rhe. 2018; 1(1): 001-003.
118. Ukoha Ukoha Ukoha, et al. Nutrient Foramina in Long Bones : Study. Am J Orth and Rhe. 2018; 1(1): 001-003.
119. Zhiquan An, et al. Human Humeral Diaphysis of the Nutrient Foramina : Anatomical Study : Study. Am J Orth and Rhe. 2018; 1(1): 001-007.
120. K-P Günther, et al. Hip Replacement in Rates: International Variation: Study. Am J Orth and Rhe. 2018; 1(1): 001-005.
121. Saeed Taj din, et al. Level of Physical Activity among Diabetic Patients of Rural and Urban Areas. Am J Orth and Rhe. 2019; 2(1): 001-004.
122. Carolina Caleza Jiménez, et al. Breastfeeding, Bed-Sharing and Early Childhood Caries. Is There An Association? A Review of the Literature. Am J Pedi and Heal care. 2018; 1(1): 001-004.
123. Katarzyna Niewiadomska-Jarosik, et al. Lipid Profile in Children Born As Small for Gestational Age . Am J Pedi and Heal care 2018; 2(1): 01-03.
124. Mustafa Aydin, et al. Antibiotic Susceptibility Pattern and Clinical Features of Klebsiella Sepsis in Newborn Infants. Am J Pedi and Heal care 2019; 1(1): 01-04.
125. H Dele Davies, et al. Necrotizing Fasciitis- Flesh-Eating Bacteria Disease. Am J Pedi and Heal care 2019; 1(1): 01-06.
126. Marie Westwood, et al. The diagnosis of urinary tract infection (UTI) in children under five years: Rapid tests and urine sampling techniques. Am J Pedi and Heal care 2019; 1(1): 01-09.
127. Folkert Fehr, et al. What Entrustable Professional Activities Add To a Primary Care Residency Curriculum. Am J Pedi and Heal care 2019; 2(1): 01-06.
128. Sonya Martin, et al. Spatially Modulated Illumination Microscopy□D measures the size of Biological Nanostructures . Ann of Phar Nano Tech and Nanomedi. 2018; 1(1): 01-05.
129. Sonya Martin, et al. Genetic analysis of Fis interactions with their binding sites. Ann of Phar Nano Tech and Nanomedi. 2018; 1(1): 01-07.
130. John H Reif, et al. Nucleation assembly of DNA tile complexes is directed by barcode-patterned lattices. Ann of Phar Nano Tech and Nanomedi. 2018; 1(1): 01-07.
131. Thomas H LaBean, et al. Self assembly of DNA nanotubes from triple-crossover tiles as templates for conductive nanowires. Ann of Phar Nano Tech and Nanomedi. 2018; 1(1):

Cite this article: A Usually Daptomycin-Resistant Gram-Positive Dermabacter Hominis: Rare Pathogen. Am J Micro and Bioche. 2019; 1(1): 001-011.

01-05.
132. Ulrich Kettling, et al. Dual-Photon Fluorescence Coincidence Analysis: Rapid quantification of Enzyme activity. Ann of Phar Nano Tech and Nanomedi. 2018; 1(1): 01-05.
133. Ahmed R. Gardouh, et al. Design, Optimization and In-Vitro Evaluation of Antifungal Activity of Nanostructured Lipid Carriers of Tolnaftate Ann of Phar Nano Tech and Nanomedi. 2019; 2(1): 01-05.
134. Mohammed Khalid, et al. Khalid M. Predictors of Prognosis in Pulmonary Hypertension. Anna Pul and Crit Car Med. 2018; 1(1): 001-004.
135. Abdullah Alsaeedi, et al. The Prevalence of Smoking among sample of Kuwait Asthmatics and its impact on the response of the treatment, Anna Pul and Crit Car Med. 2018; 1(2): 001-002.
136. Nicolau Beckmann, et al. Resolving the Oedematous Signals Induced by OVA Challenge in the Lungs of Actively Sensitised Rats. Anna Pul and Crit Car Med.. 2018; 1(1): 01-06.
137. Thomas J walsh, et al. Investigate the performance of non-invasive diagnostic tests such as galactomannan enzyme immunoassay and quantitative Caspofungin in the early diagnosis of invasive aspergillosis (IA). Anna Pul and Crit Car Med.. 2018; 1(1): 01-06.
138. Charles B. Huddleston, et al. Lung Transplantation in pediatrics. Anna Pul and Crit Car Med.. 2018; 1(1): 01-05.
139. Jeffrey P. Lamont, et al. Comparision of valved vs non-valved implantable ports for vascular access:A randomized trial. Anna Pul and Crit Car Med.. 2018; 1(1): 01-03.
140. D Inwald, et al. Risk and relevance of open lung biopsy in Nonneonatal extracorporeal membrane oxygenation (ECMO) patients. Anna Pul and Crit Car Med.. 2018; 1(1): 01-04.
141. Guillaume Mortamet, et al. Does Esophageal Pressure Monitoring Reliably Permit To Estimate Trans pulmonary Pressure In Children?. Anna Pul and Crit Car Med.. 2018; 2(2): 01-05.
142. Yang Jin, et al. Extracellular Vesicle-Shuttling MicroRNAs Regulate the Development of Inflammatory Lung Responses. Anna Pul and Crit Car Med.. 2018; 1(2): 01-04.
143. Nicola Clemente, et al. Pneumonectomy As A Salvage Therapy: A Rare Indication For A Gastric Malt Lymphoma Disseminated To The Lung. Anna Pul and Crit Car Med.. 2018; 1(2): 01-04.
144. Nicola Clemente, et al. Pneumonectomy As A Salvage Therapy: A Rare Indication For A Gastric Malt Lymphoma Disseminated To The Lung. Anna Pul and Crit Car Med.. 2018; 1(2): 01-04.
145. Victor Chew, et al. Pulmonary Cement Embolism. Anna Pul and Crit Car Med. 2019; 2(1): 01-02.
146. Victor Chew, et al. An Unusual Cause of a Tension Pneumothorax. Anna Pul and Crit Car Med. 2019; 2(1): 01-03.
147. Mark C. Lavigne, et al. A Performance Summary of Agents Used in Oral Care for Non-Ventilated and Mechanically-Ventilated Patients. Anna Pul and Crit Car Med. 2019; 2(2): 01-34.
148. Elisangela Hermes, et al. Psychomotricity in Vestibular Dysfunction Therapy (VDT): A Collective Health Question. Am J Rhin and Otolo. 2018; 1(1): 001- 005.
149. Ramtej J Verma, et al. Diethanolamine-Induced Hepatic Injury and Its Amelioration by Curcumin. Am J Toxi and Res. 2018; 1(1): 001-004.
150. Chee Kong Yap, et al. A Preliminary Screening of Cd and Pb Concentrations in the Some Traditional Chinese Herbal Medicines Bought From Selected Shops in Peninsular Malaysia. Am J Toxi & Res. 2018; 1(1): 001-004.
151. Geza Bozoky, et al. Acute Silent Non-Massive (submassive) Pulmonary Embolism. Am J Ang and Surg . 2018; 1(1): 001-003.
152. Muhammad Imran Qadir, et al. Is Hunting Lovering Associates with Pulse Rate. Am J of Viro and Dis. 2019; 1(1): 01.
153. Mujahid Rasheed, et al. Relation of Blood Group with Motion Sickness. Am J of Viro and Dis. 2019; 1(1): 02.
154. Mujahid Rasheed, et al. Views of University Paramedical Students about Causes of Pharyngitis, Its Transmission and Medicinal Control. Am J of Viro and Dis. 2019; 1(1): 02.
155. Kainat Rafaqat, et al. Views of University Paramedical Students about Causes of Pharyngitis, Its Transmission and Medicinal Control. Am J of Viro and Dis. 2019; 1(1): 02.
156. Sajid Ullah, et al. HCV Prevalence in the Volunteer Blood Donors in District Bajaur Khyber Pakhtunkhwa Pakistan. Am J of Viro and Dis. 2019; 1(1): 02.
157. Rabbia Aslam, Analogue of Breathing With Lizard Fright Am J of Viro and Dis. 2019; 1(1): 01.
158. Hurain Shaukat, et al. Linkage of Body Temperature with Exercise Am J of Viro and Dis. 2019; 1(1): 01.
159. Mariyam Javed, et al. How Breathe Rate Relates With Cricket Likeness? Am J of Viro and Dis. 2019; 1(1): 02.
160. Hakan Alfredson, et al. Achilles and patellar tendon operations performed in local anestesia, Am J Anest and Pai med. 2018; 1(1): 001-002.
161. Richard Lechtenberg, et al. Tau Interferon in Multiple Sclerosis. Amer J Neur & Neurophysi. 2018; 1(1): 001-002.

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